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# THE GOLGI APPARATUS AND THE VACUOME IN PROTOZOA—SOME MISCONCEPTIONS AND THE QUESTION OF TERMINOLOGY.

(A Review)

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1940 a

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Received May 13, 1937

## *Introduction*

In a previous communication (Subramanian, 1937), some of the criticisms offered by some of the cytologists against the existence of the Golgi apparatus in the cell have been discussed and it was shown that these criticisms, if accepted, would render critical, histological and cytological studies impossible. It was pointed out that the reasons for denying the existence of the Golgi apparatus were—(1) Some cytologists were paying more attention to the technique allowing its morphology, position in the cell, size, constitution and function to fade into the background, (2) a tendency to consider the apparatus more as a substance than as a living inclusion, and (3) the introduction of personal bias resulting in a mistaken—though unintentional—interpretation of the results of other workers. Nowhere is this more true than in the study of the Golgi apparatus in Protozoa. This state of affairs has reached such a stage that we feel a clearing of some of the issues may be highly useful. Hence, in this communication, we shall deal with some of the anomalies in interpretation of results and also the contention of some that the vacuome and Golgi apparatus are one and the same in Protozoa. While we were going through the literature on Protozoa, Patten and Beams' (1936) paper on the use of the ultra-centrifuge on some living flagellates appeared. These authors came to the conclusion that the identity of Golgi material in Euglenoid flagellates is still uncertain. We were not at all surprised at this statement in view of the enormous confusion in the field.

## *Instances of Confusion*

Bowen (1926 δ) in his review of the Golgi apparatus, comes to the conclusion from an analysis of various observations, that there is no *de novo* origin of the Golgi apparatus. At the same time Gatenby and Nath (1926)

arrived at an identical conclusion. However, they consider that the phylogenetic origin of the Golgi apparatus, whether from the nucleus or from the cytoplasm, a different problem. Thus a *de novo* origin of the Golgi being not accepted by leading cytologists, we were surprised to read Bowen's (1928*d*) statement that a logical review of the methods for demonstration of the Golgi apparatus in various kinds of Protozoa is rendered impossible by the fact that no basis for the identification of what may or may not be the Golgi apparatus has yet been agreed upon. According to Bowen in sporozoa alone is there any agreement. If the Golgi apparatus does not arise *de novo*, the Golgi apparatus of metazoa should have evolved from that of Protozoa. If there is agreement as to what is the Golgi apparatus in any one group of Protozoa, say the sporozoa, the fact will be admitted that confusion in other groups should be more due to other causes than due to technique, unless cytoplasmic behaviour in sporozoa and other Protozoa can be said to be different.

The suggestion made in some quarters (King and Gatenby, 1923) that the terminal bead of the flagellum of some primitive flagellate represents the Golgi apparatus has not contributed to a clearing of the position. Gatenby supposes that the outer layer of the bead might have been differentiated to form a lipid store-house or elaborator of energy yielding materials necessary for the nutrition of the locomotor organ. From its primitive position in the metazoan cell, always associated at some time with the centrosome-centriole complex, Gatenby believes that in the early history of the cell the Golgi apparatus and the centrosome were evolved side by side or the apparatus from the centrosphere in some way. This speculation which is given undue prominence in the review of King (1927) probably led to the doubt expressed by Bowen. Even after Hirschler's (1927) and Brown's (1930) demonstration of the Golgi in Sarcodina, Hill in her review (1933) says that the parabasal hypothesis was supported by much practical evidence put forward by Duboscq and Grasse (1924, 1925 *a*, 1925 *b*, 1927, 1928). The theory was based on the suppositions (1) that the chromophobic part of the Golgi bodies is related to the sphere in some way and that the relation between the central apparatus and the Golgi is not merely topographical, (2) that the flagellates were the most primitive Protozoans, and (3) that the Golgi apparatus originated in primitive flagellate-like Protozoa. The fact that workers in Protozoa have tried to prove the existence of the Golgi apparatus shows that everybody feels that such a structure should occur in all Protozoa. Instead of trying to prove or disprove the theory of Gatenby, Protozoologists seem to have been searching for the material composing the apparatus than the actual structure which is a living inclusion. That

this is the case will be seen from Hall's (1930 a) criticism that since little is known about the Golgi in Protozoa, the Golgi material lacks the venerable antiquity necessary to the foundation of any concept of a 'classical' Golgi apparatus in Protozoa.

*Criteria for the Golgi and its Non-Application by Workers*

King and Gatenby (1923) gave the following criteria according to which they consider certain bodies seen in the cytoplasm of a coccidian to be the Golgi apparatus. (1) Its staining and fixing reactions are identical with those of the Golgi bodies of metazoan. (2) It occupies an eccentric, juxtap-nuclear position and spreads out in the cell cytoplasm as does the Golgi apparatus in many metazoan cells, e.g., the egg and the nerve cell. (3) It consists mostly of the very characteristically shaped crescents and beads known in the case of metazoan cell as dictyosomes. (4) As in the metazoan cell, these protozoan dictyosomes can be found dividing among themselves in the ground cytoplasm. (5) During division of the cell it becomes sorted out into sub-equal groups around each nucleus and each ultimate daughter nucleus has gathered near it a part of the original apparatus. This definition is not given due importance in the reviews of King (1927) and Hill (1933) and it is clear that application of the above criteria would rule out the suggestion that the parabasal bodies, the contractile vacuole and the stigma can by themselves constitute the Golgi apparatus. Thus having lost sight of criteria that define the Golgi apparatus each Protozoologist seems to have his or her criteria as to what constitutes the Golgi apparatus. Thus Brown (1930 a) defines the Golgi apparatus as follows—(1) Consistent, not merely occasional, impregnation by osmotic methods, (2) Resistance to the usual methods of bleaching after osmication, (3) Consistent impregnation by silver methods, (4) Occurrence in Protozoa generally and not merely in certain species, (5) General similarity of form in different Protozoa. A comparison of the above criteria with those of King and Gatenby (1923) will show that some of the important ones have been left out. The definition of Brown is also in essence that of Hall (1931) who considers that neutral red stains the Golgi apparatus and treats the Golgi apparatus in Protozoa as having peculiar staining reactions. That this is the case will be seen from the following statement by Hall (1930 a) "On the other hand, it must be admitted that there are able cytologists who are not in accord with Bowen on this point, and who even go so far as to believe that the 'classical' Golgi apparatus may be a fusion product of discrete inclusions, either neutral red stainable globules (Covell and Scott, 1928; Cowdry and Scott, 1928), or in part of modified mitochondria (the 'lepidosomes of Parat')." If this is so, we have not been able to understand why Hall should be particular to

call his neutral red staining globules Golgi apparatus and Golgi bodies. Applying these criteria Hall describes certain inclusions in *Arcella* (Nigrelli and Hall, 1930). He says that these inclusions of *Arcella* are obviously more similar to the so-called 'vacuome' (Parat, 1928) than to the Golgi apparatus. "Hence it would seem that, if vacuome and Golgi material are separate and distinct cell constituents, *Arcella* possesses only inclusions which apparently belong to the former category." Hall's conception of the vacuome is not apparently Parat's conception. In *Peranema* (1929) Hall found certain neutral red staining globules and unstained vacuoles in the cytoplasm. These neutral red staining bodies according to him resemble those in sporozoans (Joyet-Lavergne, 1926; Cowdry and Scott, 1928) but differ from the vacuoles of *Euglena* (Grasse, 1925), *Ceratium* and *Peridinium* (Danegard, 1929) which are stained by neutral red and cresyl blue. Hall rejects the idea of the vacuoles in *Peranema* being homologous to the vacuoles described by Grasse and Danegard because the staining was irregular, and only few vacuoles were stained in any one specimen, the staining was always very faint and furthermore these vacuoles showed irregularity of reaction to osmotic impregnation which was not observed in the small globules. Having thus shown the doubtful nature of the bodies we shall now show that his criticisms of other workers' observations appear to be not logical at all. The Golgi apparatus has been shown by Hirschler to be vesicular and also semi-lunar in shape and these bodies were shown to have a duplex structure. In none of Hall's papers is there any reference to any duplex structure. Hall (1930 a, 1930 b, 1931) attempts to compare the 'osmophilic globules' of Protozoa seen by him with those of metazoa because of similarity in general form, intra-cellular distribution and blackening in osmotic and silver impregnation. The comparison is out of place because shape and structure described in metazoan Golgi as well as that described by Hirschler in *Endameba* and gregarines (1927) are not certainly the globules of Hall. Hall saw in *Chromulina* (1930 a) blackened crescents or almost complete rims of small vacuoles bordering a less densely impregnated substance. He dismisses these appearances because Parat (1928) showed that incomplete fixation followed by osmotic impregnation resulted in appearance of such 'crescents' while longer periods of fixation produced only uniformly impregnated globules. In Da Fano preparations of *Trichameba* (1930 b) also, he found crescents and rings, but he dismisses these, following Parat, as artifacts as longer fixation produced homogeneously blackened globules. Parat's observations and opinions are accepted by Hall and collaborators only to a certain extent. Full acceptance would have meant acceptance of some of the bodies as modified mitochondria or lepidosomes. This attempt at reconciliation

between two schools of thought divided by a wide gulf becomes all the more difficult to understand when we consider some of the arguments put forward by Hall. Bowen asserted that the Golgi apparatus in metazoa is not stained by neutral red. Bowen in his review of *Protozoa* (1928 d) says "In gregarines (*Gregarina cuneata*, *G. polymorpha* and *Steinhsa ovalis*) Joyet-Lavergne (1926 b) finds that after the slow action of a sufficiently dilute neutral red solution, small red arcs, strongly coloured, appear in the cytoplasm, each bordering an ovoid mass weakly coloured by neutral red. These bodies correspond exactly with the morphology, size and location of Golgi bodies demonstrated by methods of fixation and staining, and are similarly interpreted." This sentence is misquoted by Hall in support of his contention that neutral red stains Golgi bodies thus "Joyet-Lavergne (1926) finds that after the slow action of a sufficiently dilute neutral red solution, small red arcs (and also granules), strongly coloured, appear." The important point of the arcs bordering the ovoid mass being omitted gives a very different meaning to the quotation from Bowen. If small red arcs appear in the cytoplasm there is no reason to accept Parat's suggestion that the ereseent-shaped bodies are artifacts. On the other hand, if they are artifacts, Hall (1931) should not have used Joyet-Lavergne's (1926) evidence of the arcs staining with neutral red and blackening with osmic acid and silver nitrate, to argue against Beams and Goldsmith (1930), who said that bodies stained by neutral red and subsequently impregnated with osmic acid may be due to the direct chemical action between osmic acid and neutral red, instead of the action of osmic acid upon pre-formed contents of the vacuole. Hall's answer is that Joyet-Lavergne described osmophilic inclusions in a number of Protozoa based on material impregnated by the usual methods without previous treatment with vital dyes. "In a later investigation, Joyet-Lavergne (1926) stained certain gregarines vitally with neutral red and was so impressed with the similarity between the neutral red globules and the previously demonstrated osmophilic globules that he concluded the two sets of inclusions must be identical." We have not been able to reconcile the observation of Joyet-Lavergne in sporozoans of "small red arcs strongly coloured—each bordering an ovoid mass weakly stained with neutral red" with Hall's statement of "neutral red globules and osmophilic globules".

Having shown thus far that the attitude taken by Hall is a mistaken one, we shall now show that Hall's papers do not give an idea of the 'vacuome' or Golgi being considered as living inclusions. When terming any set of inclusions, Golgi apparatus or vacuome, Hall should have realised that the cytologists dealing with these inclusions have been considering them as living inclusions. Gatenby even in 1919 has shown that the Golgi have the

power to assimilate, grow and divide. Even 'vacuome' "appear never to arise *de novo* (?) but are permanent elements of the cell handed on in cell division and in reproduction and are presumed to have a status comparable to that of the nucleus, plastidome and cytome" (Bowen, 1927). That being so, during cell division these 'globules' should divide equally between the daughter cells and as the number thus becomes reduced they should exhibit vegetative division during cell growth. In the whole series of papers Hall refers to fluctuations in numbers only in *Vorticella* (Hall and Dunthue, 1930) and *Chlamydomonas* (Hall and Nigrelli, 1931). Even here he does not deal with these fluctuations as caused by division as the following observations of his would show. In different specimens of *Vorticella*, he says, the number and to some extent, the size of the neutral red globules vary. Ciliates from one culture may show very few globules as compared with specimens from another culture. In one instance, specimens from a three-day culture containing many dividing forms, showed very few globules as compared with material from older cultures. After a week or more however, this difference was not noticeable. "It seems that there is little accumulation of neutral red globules in *Vorticella* during the first few days after a culture is started, and it is not until the ciliates begin to divide less rapidly that they contain many neutral red globules. These observations might suggest that the vacuome bears some relation to the storage of food materials or the accumulation of waste products in the cytoplasm." In *Chlamydomonas* (Hall and Nigrelli, 1931), the larger flagellates usually contain more globules than the smaller ones and the larger the globules, the fewer they are in any one specimen. The appearance of large globules according to Hall, cannot be due to conditions encountered in sealed slide preparations since such variations were seen in flagellates from fresh stock cultures. "These variations suggest the possible occurrence of fusion or growth of smaller globules and breaking up of larger ones." Thus Hall and collaborators have no clear cut ideas of the division of Golgi bodies.

In coming to a conclusion from the above analysis we feel that many of the Protozoan workers do not seem to have applied the criteria which define the Golgi apparatus in metazoa. This indefiniteness about what constitutes the Golgi apparatus, and an acquaintance, in several cases, with papers on Protozoan cytology alone, have led to the mistaken belief that the vesicles and rings are artifacts. The duplex structure of the Golgi bodies is not considered at all and hence any attempt to bring the observations of vital staining in Protozoa with those in metazoa is impossible because of the fact that no distinction into chromophilic and chromophobic portions have been distinguished by many of the workers who assert that neutral red

stains Golgi bodies. Joyet-Lavergne admits that the staining does not remain long in the elements. This evidence of temporary staining of the Golgi bodies has been used to demonstrate the Golgi material and has led to the mistaken belief that in Protozoa at least the vacuome and the Golgi apparatus are one and the same thing. Criticisms have been offered by many that these neutral red staining bodies are volutin, etc. Leaving aside such criticisms we shall now consider whether the terms 'vacuome' and Golgi apparatus could be used for the same structure.

According to Hall and Dunilue (1931) the usage of the terms 'vacuome' by some workers and 'Golgi apparatus' by some others for the neutral red globules seems to be merely a question of terminology rather than anything else, since neutral red globules apparently satisfy the essential requirements for either classification. Such a statement raises some fundamental issues such as (1) what is the Golgi apparatus and (2) what is the vacuome? As the Golgi apparatus was first demonstrated in metazoan cells and as modern conceptions about shape, structure and function of the Golgi apparatus have been largely based on the studies of cells from both vertebrates and invertebrates, any structure if it has to be labelled the Golgi apparatus should show staining reactions and behaviour as noticed in metazoan cells unless it can be clearly proved that Golgi in Protozoa have a different structure, behaviour and function. Hall and Dunilue after quoting Gatenby's (1929) statement that as the vacuome is not consistently argentophilic the view that the vacuome is the Golgi apparatus is not tenable, remark that whether or not this objection of Gatenby is valid for metazoan cells, it fails to hold in Protozoa since a number of different workers have already shown that neutral red globules react consistently to silver as well as osmotic impregnation. Such a statement does not carry conviction and it is at best a very curious argument.

In order to understand Hall's position we have to go back to his paper on "Osmophilic Inclusions Similar to the Golgi Apparatus in the Flagellates *Chromulina*, *Chilomonas* and *Astasia*" (1931 a). There he mentions that Bowen himself had changed his opinion regarding the stainability of the Golgi material from 'certainly not' to 'probably not' and that other able cytologists go so far as to believe that the 'classical' Golgi apparatus may be a fusion product of either neutral red stainable globules or in part of modified mitochondria. He continues that whatever may be the status of vital or supra vital staining in the demonstration of the metazoan Golgi apparatus there appears to be no such problem in Protozoa. Bowen in his survey of the structure of the Golgi apparatus in metazoa (1926 b) came to the conclusion that "the important thing is that the Golgi apparatus is a substance, the exact modelling of which in the cell is purely a matter of

secondary interest". In his "Introduction to the Methods for the Demonstration of the Golgi Apparatus Part I" (1928 *a* May) while speaking generally of the silver technique for the demonstration of the Golgi, he mentions that the lack of specificity in staining reaction will doubtless seem a very serious drawback, and that in inexperienced hands, sometimes in experienced ones, the dangers of misinterpretation are very real. However, as in the case of silver methods in neurological studies, he feels that there is no reason to discount the real value of the technique. Rather is it our problem he says, to seize upon the results which come to us and make the most of them, and that such by-products of efforts to demonstrate the Golgi apparatus have sometimes proved more valuable than would have been the result had a more specific stain been obtainable. The moral of all this is that one should be careful and that the "criteria for identification of the Golgi apparatus in a given case must be based upon its morphology and behaviour, not upon its staining capacity".

In his paper on osmotic impregnation (1928 *c*, July) he comes to the conclusion that "of all known methods for demonstrating Golgi material, that of osmotic impregnation is by far the best". Hall confuses all these statements of Bowen and comes to the conclusion "This final decree leaves only 'morphology and behaviour' as the fundamental criteria for identification of Golgi material". It will be seen from the chronological order in which Bowen's conclusions are given that the decree attributed by Hall to Bowen is not final at all, for it is in a later paper and not in the same that Bowen says that osmotic acid is the best known substance for the demonstration of the Golgi material. Thus Hall leaves out the staining with osmotic acid and applying the criteria of 'morphology and behaviour' comes to the conclusion that the criterion of behaviour fails to offer any parallel between sporozoa and metazoa. According to him if one accepts the 'Golgi apparatus' of sporozoa as homologous with that of the metazoa in spite of the scanty evidence offered by Bowen's single available criterion 'morphology', then the logical deduction seems to be that any set of granular, globular, elongated or crescent-shaped inclusions in the Protozoa may be considered Golgi apparatus provided first, that they are blackened in osmotic impregnation (and withstand the usual bleaching methods) and second that they may be distinguished from chondriosomes. An additional criterion, that of vital staining with neutral red may, according to him, be used to identify the Golgi material of other Protozoa with that found in sporozoa. Having shown previously that Bowen's stress upon 'morphology and behaviour' did not weaken his assertion that osmotic acid is the best known substance for the demonstration of the Golgi apparatus, we shall now show

that 'morphology and behaviour' are as important. According to Hall if one accepts the contention of Bowen that the staining capacity must not be relied upon, the only other criterion left is the resemblance in shape between the sporozoan Golgi elements and the discrete metazoan Golgi bodies which vary both in size and form. The above statement cannot be accepted since King and Gatenby (1923) and Joyet-Lavergne (1924) compare the behaviour of the Golgi bodies in various sporozoans to the behaviour of the same cell organ in some eggs, sperms and nerve cells.

In somatic or germ cells which are in a quiescent condition the Golgi bodies in metazoans generally occupy a place near the centriole and just before cell division they get scattered throughout the cell and also increase in number. When the daughter cells separate they have been known to be equally divided between the two daughter cells. It is this characteristic that has been used as one of the criteria by King and Gatenby (1923) to identify some bodies seen in *Adelie* as the Golgi apparatus. Hall (1930 a) himself mentions that the Golgi apparatus exhibits certain types of behaviour in spermatogenesis, secretory cells, etc. A critical analysis of these fundamental characteristics has been made by Gatenby even as early as 1920 which led him to classify the Golgi apparatus among living 'protoplasmic inclusions', as distinguished from 'deutoplasmic' or non-living inclusions. Further Gatenby comes to the conclusion (1919) that a uniform distribution of the mitochondria and Golgi apparatus during cell division has something to do with the control of cell metabolism. Growth and division of Golgi bodies have been observed during increase in size of the cell just prior to division and during intense cellular activity. Bowen (1928 d) apparently seems to have included the characteristics which led the Golgi apparatus to be included as 'protoplasmic inclusions' under the terms 'morphology and behaviour'. Another contention of Hall (1930 a) seems to be that the dispersed sporozoan Golgi apparatus is not even remotely similar in morphology to the original "apparato reticolare interno" of Golgi and since little is known about this phase of Protozoan cytology the Golgi material lacks the venerable antiquity necessary to the foundation of any concept of a 'classical' Golgi apparatus in the Protozoa. It should be pointed out here that even though Golgi and his pupils conceived of the Golgi apparatus as essentially network-like in shape, Weigl was the first to show that in invertebrates they had a scattered distribution. Weigl's corrosive osmotic fixation followed by post-osmication are being used to-day and many workers have succeeded in demonstrating the Golgi apparatus by Weigl's technique in secretory cells of both vertebrates and invertebrates where though they have different shapes, they have an identical function.

(see especially Bowen, 1926 a) When it is remembered that Golgi's original technique and that of Weigl show networks in vertebrate somatic cells and discrete bodies in invertebrates and germ cells, when different somatic cells of vertebrates themselves show discrete bodies (lipoid or skin glands) and networks and when networks themselves break up into discrete bodies during cell division, the usage of the term Golgi apparatus seems to have been based on staining reactions, morphology and behaviour. Even in Protozoa Hall's statement that the Golgi material lacks antiquity seems to be unjustified.

Hirschler even in 1914 demonstrated in sporozoa bodies which showed identical structure and behaviour as those found in metazoan cells investigated by him and hence any work on Protozoa on the Golgi bodies should proceed on the fundamental criteria employed by Hirschler, in terming bodies seen by him in sporozoa as the Golgi apparatus.

Golgi's "apparato reticolare interno" has been designated by his disciples and followers as the Golgi apparatus and this procedure is followed by other students of cytology according to priority of nomenclature. Proceeding on this basis any particular cell inclusion in Protozoa should be termed Golgi apparatus if it satisfies the conditions according to which a host of workers have styled a particular organ of the cell as the 'Golgi apparatus' in metazoa. Hall hints that these discrete inclusions mentioned by him in *Chromulina*, *Astasia* and *Chilomonas* (1930 a) may not be the Golgi bodies at all in the metazoan sense and that there is a probability of their being merely metabolic products which show the characteristics of Golgi material. "Until more is known about this aspect of Protozoan cytology, such a question cannot be settled one way or another. In the meantime, whether we accept the rather convincing arguments of Parat or follow the somewhat bewildering dictates of Bowen, there is no reason why these discrete inclusions of Protozoa may not be accepted as true Golgi material, since, so far as the present criteria for identification of metazoan Golgi apparatus extend, these inclusions of Protozoa satisfy all essential requirements." We have not been able to understand this particular attitude of Hall. It is surprising that after stating the point of view of Parat that the classical Golgi apparatus may in part be modified mitochondria Hall suggests the acceptance of the discrete bodies as 'true Golgi material'. As pointed out previously the fact that neutral red does not probably stain the Golgi material in metazoa itself shows that the discrete bodies shown by Hall do not satisfy all the essential requirements. We should like to mention here that temporary staining of the chromophobic portion of the Golgi vesicles in *Meretrix* eggs have been noticed by us (Subramaniam, 1937), but never could the results be compared with what has been observed by Hall in various

Protozoa Neutral red vacuoles in *Meretrix* arise as segregation products and its segregation by the Golgi apparatus is comparable to the secretion of fat, fatty yolk and yolk in eggs

Hall's (1931) reasons for considering the vacuome as the classical Golgi apparatus seem to be that (1) the vacuome is consistently blackened in osmotic impregnation (2) it is resistant to bleaching after osmication, (3) it is consistently impregnated by silver methods, (4) so far as available observations indicate, form and intra-cellular distribution of the elements of the vacuome are in general similar throughout Protozoa, and (5) a vacuome has been demonstrated in representative species of the major groups of Protozoa

It is surprising that Bowen (1928 d), King (1927) and Hill (1933) who noted the disagreement as to what constitutes the Golgi apparatus in Protozoa have not defined the criteria on which are based the identification of Golgi material in metazoa. Such a clear analysis of the criteria of staining, structure, shape and morphology of the Golgi apparatus in the metazoa together with what constitutes the vacuome will, we believe, go a long way in clearing the position for a study of what constitutes the Golgi material in Protozoa

#### *Definition of the Golgi Apparatus*

- 1 The Golgi apparatus is a permanent structural element in the cytoplasm universally present in all cells
- 2 In quiescent cells generally it has an excentric juxta-nuclear position
- 3 It possesses the power of independent growth and multiplication and is passed on from cell generation to cell generation by division processes which have been shown to be of special interest
- 4 The Golgi apparatus arises only from pre-existing Golgi and does not arise *de novo*
- 5 The apparatus assumes various forms and in invertebrates and germ cells generally it is constituted by discrete bodies having a duplex structure
- 6 The apparatus has a lipoidal basis and may also have a second constituent which is protein in character
- 7 The Golgi material in many cases is set off rather sharply from the cytoplasmic background and could sometimes be seen in fresh material under ordinary and even dark ground illumination
- 8 The apparatus is consistently impregnated by osmotic and silver technique and is resistant to the usual methods of bleaching after osmication.

9 The apparatus shows an increase in size following an increase in volume of the cytoplasm or during active synthesis of visible granules from raw materials in eggs, sperms and secretory cells. This hypertrophy is by an increase in number mainly in invertebrates and germ cells—also in vertebrate lipoidal cells—and by an increase in complexity and size in those cells where networks occur.

*Parat lays down the following Points for the guidance of Workers in the Identification of the Vacuome*

1 There are present in all cells two and only two kinds of formed elements in the cytoplasm. The elements in question are (1) the chondriome or the mitochondria, stainable with Janus green and (2) the vacuome stainable with neutral red. Thus Janus green and neutral red (especially Krall's Microcolor) are considered specific in a narrow sense.

2 The chondriome is a lipoidal phase and the vacuome a watery one.

3 The Golgi material of animal cells as usually described does not exist. The appearance of a Golgi apparatus results from the fact that within, around and between the vacuoles of a cell osmic acid or silver nitrate is reduced to form an entirely artificial structure which going now this way, now that, produces that illusion of a rambling network. Thus the network produced has no fixity of shape.

To summarise, the first essential characteristic of the vacuome is its specificity for neutral red and the second, its constancy. The third is the nature of the vacuolar contents which are never lipoidal (see Parat and Bergeot, 1925) but always watery with a reaction 'franchement acide' and probably made up in most cases of a solution of crystalloids.

*Facts against Considering the Vacuome as the Golgi Apparatus*

- 1 Vacuoles are not universally present in animal cells.
- 2 There are two types of neutral red staining structures (a) artificial and (b) pre-existing. Hence neutral red cannot be considered specific.
- 3 Osmic acid does not cause deposits within animal cell vacuoles.
- 4 Parat's explanation of the artificial nature of the Golgi apparatus as seen after silver and osmic technique does not cover the discrete bodies of invertebrates. His interpretation that the rod-shaped bodies of *Helix* are modified mitochondria is based on staining with Janus green and if this has to be admitted, there is also the fact that the vacuome may often be stained with Janus green which strikes at the very root of his theory.
- 5 Even in lipoid gland cells of different kinds, the Golgi material appears in various forms always characteristic for a given cell type. As

Parat has based his hypothesis on the results obtained in plant cells it may be mentioned here that though the vacuoles in the pro-meristem cells blacken, they have not been observed to have the remotest resemblance to a Golgi network. Moreover no precipitations have been observed between the vacuoles either in plant or gland cells.

6 The demonstration of the osmophilic platelets (Bowen, 1927) and their being considered as homologous to the animal Golgi apparatus have shaken the fundamental basis of Parat's theory.

As would appear from the criteria defining the vacuome, staining with neutral red is the most important criterion. If this is so one wonders why the food vacuoles in Protozoa should not be termed vacuome. Bearing in mind Parat's postulate it would appear that the food vacuoles in specimens studied by Hall satisfy more of the conditions than the osmophilic bodies, for (1) they are all stained by neutral red, (2) often osmic acid and silver nitrate are precipitated inside these vacuoles, and (3) their contents are not generally lipoidal!

A persual of the above fundamental though elementary facts will show that the terms vacuome and Golgi apparatus cannot be used for the same structure. If a body is to be considered as the vacuome, it cannot be termed Golgi apparatus for it is not lipoidal and if a body is considered the Golgi apparatus it cannot be considered the vacuome for it is not an acidic solution of crystalloids.

In addition to the above fundamental differences between the vacuome and the Golgi apparatus, the test of cell nomenclature is how it works when applied to different categories of cells. Considering the Golgi apparatus and the vacuome as essentially the same leads to enormous confusion making it difficult to reach any point of agreement between the different schools of thought. Use of terminology is a matter where there cannot be serious differences of opinion and an agreed terminology together with criteria defining what constitutes the Golgi apparatus will clear the field for a correct understanding of the shape, structure and function of the Golgi bodies in Protozoa.

Analysing the results obtained in Protozoa according to the above criteria it seems to us that the contractile vacuole, stigma and parabasal bodies could not be considered as the Golgi apparatus because they possess no power of independent growth and vegetative multiplication. In *Jania annulans* Duboscq and Grasse (1928) particularly emphasise the secretory activity of the parabasal during division. They saw chromophile vesicles each composed of a cap of chromophile substance surrounding chromophobe

material being expelled into the cytoplasm Chatton and Grasse (1929) observed in *Polykrikos schwartzii* osmophile vesicles Three streamer-like parabasals have also been seen by them and they are inclined to think that the vesicles are the products of granules liberated by the parabasal reminiscent of the secretion of very similar structures by the parabasal in *Jenia*

Proceeding on the criteria given above for the identification of the Golgi apparatus the osmophile vesicles together with the parabasals constitute the Golgi apparatus in *Jenia* for only such a concept enables even the parabasal to be considered as Golgi as postulated by Grasse It is surprising that osmophile vesicles described by these authors which have identical reactions as the parabasal itself in *Jenia* are interpreted by them as secretion, whereas they have to be considered as Golgi bodies Hill (1933) in her review misinterprets Brown's observations on *Microjenia* and suggests that the vesicles and batonettes described by Brown may be mere vesicles secreted by the parabasal She forgets that in metazoa no case of a Golgi body secreting another has ever been noticed Taking both together, the osmophile vesicles and the parabasal, they are seen to satisfy most of the conditions laid down for the identification of Golgi material in *Jenia* If this is accepted, formation of osmophile vesicles is not by secretion but by fragmentation It is not quite necessary that all parabasals should react to Golgi technique—and thus be considered parts of the apparatus—for the parabasals alone cannot constitute the Golgi apparatus Similarly the contractile vacuole alone cannot be accepted as the Golgi apparatus, for as in the case of parabasals and stigma it also does not show growth and vegetative multiplication though it may divide during cell division Hall (1930 a) records some interesting results in flagellates In a random example of 100 flagellates, 39 had both the contractile vacuole and the *globules* (which according to him stain with neutral red and are considered as Golgi) were blackened, 15 in which the *globules*, contractile vacuole and other cytoplasmic vacuoles were blackened, 9 in which the *globules* and cytoplasmic vacuoles but not the contractile vacuole remained blackened, 25 in which only the *globules* remained blackened, 3 in which the *globules* and cytoplasmic vacuoles were bleached completely, while the contractile vacuole was only partly bleached, and 9 in which the *globules* and vacuoles were completely bleached

Hall comes to the conclusion that as the *globules* were blackened in 88 per cent they alone are the Golgi bodies The same result offers another interpretation If the *globules* are to be accepted as the Golgi bodies, it will be seen that in the largest number of cases (39 per cent) with no artifacts both the contractile vacuole and the *globules* were blackened which

leads to the logical conclusion that both together should be considered as Golgi apparatus in the particular flagellates.

On the whole from a critical reading of the available literature it appears that what is described in many cases as Golgi apparatus in Protozoa, excepting probably the sporozoa, does not answer to the definition of the Golgi apparatus and that further critical studies, bearing in mind that some of the Golgi elements may become modified as the contractile vacuole, parabasal or stigma in some species, may offer a new approach to the study of the Golgi apparatus in Protozoa.

#### BIBLIOGRAPHY

Baker, Clinton L. "Studies on the Cytoplasmic Components of *Endamoeba gracilis* Klebs," *Arch. Protist.* 1933, **80**, 434-468.

Beams, H. W., and Goldsmith, J. B. "Golgi bodies, Vacuome and Mitochondria in the Salivary Glands of Clitostomous larva," *J. Morph.*, 1930, **50**, 497-515.

Bowen, R. H. "Studies on the Golgi Apparatus in Gland Cells. Parts I, II, III & IV," *Quart. Journ. Micr. Sci.*, 1926 a, **70**, 75-113, 193-217, 395-451.

\_\_\_\_\_. "Golgi Apparatus—Its Structure and Functional significance," *Anat. Rec.*, 1926 b, **32**, 151-193.

\_\_\_\_\_. "The Golgi Apparatus and the Vacuome," *Anat. Rec.*, 1927, **35**, 309-335.

\_\_\_\_\_. "The Methods for the Demonstration of the Golgi Apparatus I. Intra Vital Observations, Vital Staining Fresh Preparations and Tissue Culture," *Anat. Rec.*, 1928 a, May, **38**, 293-321.

\_\_\_\_\_. "II. Silver and Gold Methods," *Anat. Rec.* 1928 b, June, **39**, 86-136.

\_\_\_\_\_. "III. Methods of Osmic Impregnation," *Anat. Rec.* 1928 c, July, **39**, 231-276.

\_\_\_\_\_. "VI. Protozoa, The Vacuome, Plant Tissues," *Anat. Rec.*, 1928 d, **40**, 225-276.

Brown, V. E. "Hypermastigote Flagellate from the Termite *Recticulitermes*," *Univ. Calif. Pubns. Zool.* 1930 a, **36**, 67-80.

\_\_\_\_\_. "The Golgi Apparatus of *Amoeba proteus* Pallas," *Biol. Bull.*, 1930 b, **59**, 240-247.

\_\_\_\_\_. "Cytoplasmic Inclusions of *Endamoeba gracilis* Klebs," *Z. Zellf. u. mikr. Anat.*, 1930 c, **11**, 244-254.

Chatton, E., et Grasse, P. "Le Chondriome, le Vacuome, les vesicules osmophiles, le parabasal, le trichocystes et les endocystes du Dino-flagelle *Polykrikos schwartzii*, Butschli," *C. R. Soc. Biol.*, 1929, **100**, 281-285.

Covell, W. P., and Scott, G. H. "An Experimental Study of the relation between granules stainable with neutral red and the Golgi Apparatus in Nerve Cells," *Anat. Rec.*, 1928, **38**, 377-399.

Cowdry, E. V., and Scott, G. H. "Etudes cytologiques sur le paludisme III. Mitochondries, granules colorable au rouge neutre et appareil de Golgi." *Arch Inst Pasteur (Tunis)*, 1928, 17, 233-252

Danegard, P. "Coloration vitale de l'appareil vacuolaire chez les Peridimens marins," *C R Acad Sci*, 1923, 177, 978-980

Dawson, A. B. "The Segregation Apparatus of the Amphibian Erythrocyte and its possible relation to the Golgi apparatus." *Anat Rec*, 1928, 39, 137-151

Duboscq, O., et Grasse, P. "Sur les protistes parasites des Termites de France. Sur l'évolution de *Janickiella Grasse*," *C R Soc Biol*, 1924, 90, 547-550

\_\_\_\_\_ "L'appareil parabasal et son évolution chez *Holomastigotes elongatum*," *C R Soc Biol*, 1925 a, 92, 154-156

\_\_\_\_\_ "L'appareil des flagelles et sa signification," *C R Acad Sci*, 1925 b, 180, 477-480

\_\_\_\_\_ "Sur la division mitotique de *Trichonympha chattoni*," *C R Soc Biol*, 1927, 96, 92-94

\_\_\_\_\_ "L'appareil parabasal de *Jewia annexans*," *C R Soc Biol*, 1928, 99

\_\_\_\_\_ "L'appareil parabasal des flagelles avec remarques sur le Trophospunge, L'appareil Golgi, les Mitochondries, et le Vacuome," *Arch sool Exp et Gen*, 1933, 73, 381-621

Dunshue, F. W. "The Vacuome and the Neutral Red Reaction in *Paramcromus conditum*," *Arch Protistenk*, 1931, 78, 475-487

Gatenby, J. B. "The Cytoplasmic Inclusions of Germ Cells V. The Gametogenesis and Early Development of *Limnaria stagnalis* with special reference to the Golgi Apparatus and the Mitochondria," *Quart Journ Microsc Sci*, 1919, 63, 445-493

\_\_\_\_\_ "Study of the Golgi Apparatus and the Vacuolar System in *Canna*, *Helix* and *Abraxas* by Intra Vital Methods," *Proc Roy Soc London, (B)*, 1929, 104, 302-322

Gatenby, J. B., and Nath, V. "The Oogenesis of certain Invertebrates with Special reference to *Lumbriacus*," *Quart Journ Microsc Sci*, 1926, 70, 371-391

Gatenby, J. B., and Woodger, J. H. "On the Relationship between the formation of Yolk and the Mitochondria and the Golgi Apparatus during Oogenesis," *J Roy Microsc Soc*, 1920, 129-156

Grasse, P. "Vacuome et appareil de Golgi des Euglenes," *C R Acad Sci*, 1925, 181, 482

\_\_\_\_\_ "L'appareil parabasal des flagelles est-il un organite sécréteur?" *C R Soc Biol*, 1925 b, 93, 1097-1099

\_\_\_\_\_ "Contribution à l'étude des flagelles parasites," *Arch. sool Exp et Gen*, 1926 a, 65, 345-602.

\_\_\_\_\_ "Sur le stigma ou appareil parabasal des Euglenes," *C. R. Soc Biol*, 1926 b, 94, 1012-1014

Hall, R. P. "Reactions of certain Cytoplasmic Inclusions to Vital Dyes and their Relation to Mitochondria and the Golgi Apparatus in the flagellate *Peranema trichophorum*," *J. Morph.*, 1929, 48, 105-121

— "Osmophilic Inclusions similar to the Golgi Apparatus in the Flagellates *Chromatina*, *Chilomonas* and *Astasia*," *Arch. Protistenk.*, 1930 a, 69, 7-22

— "Cytoplasmic Inclusions of *Trichomonas* and their relation to Vital Dyes and Osmic and Silver Impregnation," *J. Morph.*, 1930 b, 49, 139-153

— "Cytoplasmic Inclusions of *Menardium* and *Euglena* with special reference to the 'Vacuome' and Golgi Apparatus of Euglenoid Flagellates," *Ann. Protistol.*, 1930 c, 3, 57-68

— "Vacuome and the Golgi Apparatus in the ciliate *Stylonychia*," *Zeit. zell. u. mikr. Anat.*, 1931, 13, 770-782

— "Cytoplasmic Inclusions of Phytomastigida," *Bot. Rev.*, 1936, 2, 85-94

Hall, R. P., and Alvey, C. H. "The Vacuome and the so-called Canalicular System of Colpedita," *Trans. Amer. Micr. Soc.*, 1933, 52, 26-32

Hall, R. P., and Dunthorne, F. W. "On the Vacuome and Food Vacuoles in *Vorticella*," *Trans. Amer. Micr. Soc.*, 1931, 50, 196-205

Hall, R. P., and Loeffler, J. B. "Studies on *Euglypha* I. Cytoplasmic Inclusions of *Euglypha alveolata*," *Arch. Protistenk.*, 1930, 72, 365-376

Hall, R. P., and Nigrelli, R. F. "Relation between Mitochondria and Food Vacuoles in the ciliate *Vorticella*," *Trans. Amer. Micr. Soc.*, 1930, 49, 54-57

Hill, J. C. "The Vacuome of the Flagellate *Chlamydomonas*," *J. Morph.*, 1931, 51, 527-541

Hirschler, J. "The Golgi Apparatus of Protozoa," *J. Roy. Micr. Soc.*, 1933, 227-247

— "Über Plasmastrukturen (Golgischen Apparat, Mitochondrien u. a.) in den Tunicaten, Spongien und Protozoen," *Anat. Anz.*, 1914, 47, 289-311

— "Studien über die sich mit Osmium schwarzenden Plasmakomponenten (Golgi Apparat, Mitochondrien) einiger Protozoenarten, nebst Bemerkungen über die Morphologie der ersten ihnen im Thierreiche," *Zeit. zell. u. mikr. Anat.*, 1927, 5, 704-786

Joyet-Lavergne, P. "L'appareil de Golgi dans les schizozoïtes d'un Aggregatidé," *C. R. Soc. Biol.*, 1924, 90, 680

— "Recherches sur le cytoplasme des sporozoaires," *Arch. Anat. Microsc.*, 1925, 22, 1-128

— "Sur la coloration vitale au rouge neutre des éléments de Golgi des Gregarnes," *C. R. Soc. Biol.*, 1926, 94, 830-832

King, S D "The Golgi Apparatus of Protozoa—A Review," *J Roy Microsc Soc*, 1927, 342-355

King, S D, and Gatenby, J B "Golgi Bodies of a Coccidian," *Quart Journ Micro Sci*, 1923, 67, 381-389

Nassonov, D "Der Exkretionsapparat (Kontraktile Vacuole der Protozoa als homologon des Golgiischen Apparats der Metazoenzellen," *Arch Mikr Anat*, 1924, 103, 437-482.

— "Zur Frage über den Bau und die Bedeutung des Lipoiden Exkretionsapparats bei Protozoen," *Zent Zellf u mikr Anat*, 1925, 2, 87-97

Nigrelli, R F, and Hall, R P. "Osmiophilic and Neutral Red stainable Inclusions of *Arcella*," *Trans Amer Micro Soc*, 1930, 49, 18-25

Parat, M "Contribution à l'étude morphologique, et physiologique du cytoplasme, chondriome, Vacuome (appareil de Golgi encalves, etc)," *Arch Anat Microsc*, 1928, 24, 73-357

Parat, M, et Bergot, P. "Sur le pretendu contenu lipoidique de l'appareil de Golgi," *C R Soc Biol*, 1925, 92

Patten, R, and Beams, H W "Observations on the effect of Ultra-Centrifuge on some Living Flagellates," *Quart Journ Micro Sci*, 1936, 78, 615-637

Subramanian, M K "Oogenesis of *Meristix casta* with a Note on the Nature of the Contents of Neutral Red Vacuoles," *J Morph*, 1937, 61

— "An Analysis of Certain Criticisms against the Existence of the Golgi Apparatus," *Proc Ind Acad Sci*, 1937, 5, (2), 48-56

## CLIMATIC CONDITIONS IN SIND.

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Received May 4, 1937

(Communicated by Prof. D. N. Wadia, M.A., F.R.G.S., F.A.S.S.)

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### *I. Introduction*

PARTS of India form some of the warmest regions of the earth. In this tropical country, having a climate of great diversity and range, Sind is found to be the driest and hottest of all its provinces, the distribution of solar radiation playing a prominent part in it. Aridity is its main characteristic, with Jacobabad (mean maximum temperature 113° F and mean humidity 41 per cent) as the hottest and driest place in India. The tropic of Cancer passes a little below its southern boundary and so the solar heat is great, coupled with longer hours of sunshine in the hot season. The climate is reminiscent of the Sahara type, and of that prevailing in hot and dry lowlands. It presents a great contrast with Assam at the other end of the country, as regards humidity—a contrast greater than that between Egypt and the British Isles. It is rather cold in winter for the latitudes within

which it falls (viz.,  $23^{\circ} 35'$  and  $28^{\circ} 30' N$ ), parts of it being cooler than the Punjab, situated higher up and encompassed between latitudes  $29^{\circ}$ - $31^{\circ} N$

The thermal equator (the line of the greatest mean heat of the globe) passes through Sind. "It has long been known that what is termed, the thermal equator, does not follow the course of the equinoctial line, where, on the average of the year, the sun's action is most direct and intense, but in the longitude of India bends northwards, running up through Ceylon and the peninsula to beyond the tropic and *passing through Sind*, crosses thence to the Arabian peninsula."<sup>1</sup>

In the matter of rainfall, another important factor of climate, the region also suffers greatly. It is said to be "between the two monsoons", known as the S.W. and N.E. It manages to miss the influence of the former, while the latter does not extend much beyond the Ganges basin. The average rainfall in the valley is meagre and precarious. Contrast with this the region of Western Deccan, which receives almost the whole of its annual rainfall in summer and in abundance, and the N.W.F. Province with its main share of rains in winter. The mountains on the west of Sind are not sufficiently high to catch the S.W. monsoon currents in one part of the year or to prevent the cold blasts from the Iranian Plateau from entering the region, in the other (See Plate I.)

Thus the chief features which affect the climate of Sind are:—

- (1) The tropical zone in its southern half and the temperate zone in its northern one
- (2) The position and direction of mountains, e.g., the Himalayas which prevent the northern cold and induce the southern rains
- (3) The Thar Desert Region in the neighbourhood
- (4) Nearness or remoteness from the Arabian Sea
- (5) Presence or absence of forests.

Within the region of Sind there are some marked climatic contrasts. There is an uneven distribution of rainfall from one end of it to another. Jacobabad has an average rainfall of 3 inches, while Nagar Parkar has 15 inches. There is almost no rainfall in some parts of the desert area ordinarily, but in years of heavy downpour a whole season's rainfall is received at some places in Sind in a couple of days, it being as heavy as 8 inches per hour, e.g., on 17th June 1912, when 7 inches of rainfall were recorded at Karachi.<sup>2</sup> Places nearer the sea get somewhat better and regular rains. The diurnal and seasonal variations of temperature are also great in many parts of Sind. The thermometer drops below the freezing point in winter with frequent cold spells and frost in North Sind, while heat waves with

whirlwinds and dust storms are common in summer throughout the Province. There are no great forests in Sind to render the rainfall more effective except in a few narrow strips along the river banks. Dry land winds blow most of the year. "It is this persistence of land winds which explains the aridity of the Indus Valley, for the south-west direction, that prevails in the Valley itself, is merely that which is given to the current by the trend of the hills that bound it."<sup>10</sup> This dry wind system prevails over the greater part of the Province, except near the coast, where the influence of the monsoon winds is felt somewhat. But the chief causes of this draught in Sind are the deficient relief of land and the very high temperatures and low humidity, jointly operating to reduce the chances of rain or even of cloud formation (Ref. Simpson's theory in Section IV). On the whole, the climate of Sind can be called a Desert climate—a cold dry winter, alternating with a hot summer, with very meagre and precarious rainfall.

## *II. General Climatic Conditions of India Affecting Sind*

### *A. The Monsoons.*

India is one of the monsoon lands of Asia. The word 'Monsoon' is derived from the Arabic *mausim*, season or seasonal wind, the wind direction being the chief factor in the practice of navigation of those days. This wind depends upon the principle that it always blows from places which have greater barometric pressure to where it is less. This wind pressure gradient in its turn, depends upon the rate of heating and cooling between land and sea.

The very nature of the peninsular projection of India into the Indian Ocean helps this. During the months of March, April and May the sun's rays fall almost perpendicularly on Northern India and the barometric pressure in this region is the lowest, while over the Indian Ocean and lower down, the air is comparatively cool and the pressure high. A flow of air is consequently generated inland from the seas, and, crossing them for nearly 4,000 miles, it is charged with considerable moisture. Also the hotter the air, the more moisture it carries in an invisible form. But this current of air does not flow directly from south to north, due to the rotation of the earth, the air currents in the Northern Hemisphere circulate anti-clockwise, hence the S E trade winds. On crossing the equator, they are caught in the circulation around the low pressure system existing over the N W corner of India and Sind, and move, over the region of the Arabian Sea on one side and the Bay of Bengal on the other, deflected as the S W monsoon, according to Ferrel's Law. At the equator due to the rotation of the earth, the monsoon has acquired a high velocity of hundreds of miles per hour;

but as it goes up to higher latitudes it reaches the air currents which move slower and hence overtake them, deflecting itself eastwards

Over Sind, there is in summer a persistent depression with an oval isobar of only 29 inches round about Jacobabad, and an isotherm of about 100° F co-terminous with it. There is, therefore, a difference of nearly an inch of barometric pressure between South India and this region, which at once becomes a theatre of great atmospheric depression, breeding thunderstorms and air disturbances. In this respect the contribution of Sind to the whole country of India and its rainfall is really very great.

On entering India, this moisture is precipitated as the air advances northwards over the country, because the general pressure distribution over Asia and the Indian Ocean forces it into this box or *cul-de-sac*, out of which it can only find a way by rising over the sides."

*Causes of Precipitation*—But though, the chief cause of the rainy season is this pressure distribution over land and sea, actual precipitation of rain is due to several other circumstances. The orographical structure of the whole country is mainly responsible for it. At the very outset the Western Ghats obstruct the path of the monsoon. Such an obstruction is beneficial, as the air currents are pushed upon a height of nearly 5,000 ft. and the vapour is condensed. Also, as the currents blow across the whole country, they find no openings in the north, north-west and north-east being blocked by the extra-peninsular mountain ranges of insurmountable heights. They, therefore, must rise higher and higher. This ascensional movement, thousands of feet above the sea-level, causes complete condensation and rainfall for the country.

*Branches of the SW Monsoon*—It should be noted that these ascensional movements flow in two different directions (1) Those flowing over the Arabian Sea crossing the Ghats and penetrating the Narbada and Tapti Valleys called the Arabian Sea branch and (2) Those blowing from the Bay of Bengal along the Burmese Yomas and the Assam Hills called the Bay branch of the summer monsoon. Further still, as the Bay current is checked by the Himalayan barrier, it bends westwards as the S E monsoon and passing up the Gangetic Valley, it reaches the eastern limits of the Punjab in a very weak state.

Those parts of India, which do not lie within the paths and ranges of these two SW and S E monsoon currents, those which lie in the rain-shadow areas, and those, again, which have low or no mountain barriers, receive scanty rainfall in this season. Thus Mahabaleshwar, 4,500 ft. on the windward side of the Western Ghats, receives an average rainfall of 300 inches, Poona and Bangalore on the leeward side of the Western Ghats

receive only 25 and 10 inches respectively, while Peshawar in the extreme NW gets less than 15 inches and Jacobabad in Sind less than 5 inches. Most of Sind lies outside the path of the SW monsoon and it has no high mountain barriers to catch it. The other branch of the summer monsoon, viz., the Bay current, giving as much as 500 inches (average) to the Assam Hills (Cherra Punjee), does not reach parts of Sind and the Punjab, as the currents get weaker and weaker as they travel further westward thus Peshawar gets only 4½ inches and Sukkur 2 inches during the whole summer season. Even the cyclones, developed in the Bay of Bengal during the monsoon period, do not usually travel as far as the Indus basin.

This wet season lasts for about four months from June to September, at the end of which period the moisture is exhausted and the currents become greatly weakened. As a rule three-fourths of the total annual rainfall in India is caused by this monsoon current.

*N E. Monsoon*—By the end of September, the meteorological conditions in India are reversed. The rays of the sun are also aslant in these months. While the SW monsoon is still hanging over the country, particularly over Bengal, the air over the greater part of Central Asia becomes cold and an anticyclone, exceeding 31 inches and resting on Tartary, is developed. North of this region, the barometric pressure slopes away to the Arctic Ocean and south of it, it falls away to the equator, thus causing the Central Asiatic anticyclone to be persistent for some time.

On account of these changed conditions, the SW monsoon current, still flowing into the Bay of Bengal, cannot enter this high pressure region, but retreats and goes round the area of low pressure in South India, as what is called, the *retreating monsoon*. This is also called N E monsoon. "After September, the region of chief indraught gradually travels southwards, the moisture-laden winds from the Indian Ocean no longer penetrate so far in a northerly direction. Accordingly, in October the pressure in the Bay of Bengal is in general the lowest in the area, lying off the Madras coast, and from this time onwards, rain occurs in the Peninsula chiefly in connection with a series of depressions or areas of low pressure, which form in the Bay of Bengal and strike the coast. The rain-bearing winds at such times are north-easterly and hence the rains of the N E monsoon."<sup>4</sup> This current does not affect Sind. It gets gradually exhausted in its passage westwards and there is a break in the rains for about a month or so, viz., October.

*Winter Rains in Northern India*—By November-December the temperature at Jacobabad falls as low as 50° F and though the pressure gradient

is also low, steady winds begin to flow from the N.W. These are mainly continental winds and therefore, dry. But where they pass over portions of the seas, such as the Persian Gulf, they carry some moisture which is condensed and precipitated by the northern winds. Thus we have snows on the Himalayas and winter rains in Northern India.

It should be borne in mind that the belt of high pressure and the anti-cyclonic conditions in winter are not local but stretch over a wide area of varying physical features from the Mediterranean to Central Asia as far as N.E. China, corresponding to the belts of high latitudes in the Atlantic and the Pacific Oceans. As a result of this, shallow or feeble storms pass successively across Central Asia and occasionally also over Northern India. Thus lightly clouded skies and occasional showers of rain are the predominant weather conditions in winter in the Indus basin.

#### *B Other External Causes*

Apart from the regular monsoon currents, such cyclonic storms or disturbances as cross the whole region are a very important feature of the climate of Sind. Dr B. N. Banerjee<sup>4</sup> has classified the meteorological conditions prevailing over the section of the Persian Gulf to Karachi, under two distinct types, the winter and the summer, western depressions and eastern depressions respectively. These storms differ from the regular cyclones, which are winds of great destructive violence but not bearing much rain. The cyclonic storms, though milder, are very extensive and bring about winter rains in Northern India. The difference of barometric pressure or gradient too is very small, e.g., about  $\frac{1}{10}$  inch. "Also they are more lasting than the cyclones, and at times bring torrential rains. They travel over hills without any loss of energy unlike cyclones, which are dissipated by mountains." Thus floods are caused when a shallow low disturbance is superimposed on the general regional flow. It has been noticed that the Makran Coast from Gvadar to Karachi is disturbed more by summer weather storms than by winter ones, while in the interior of Sind and the Punjab it is the western (winter) disturbances which count more.

*Winter Conditions: Western Depressions*—From December to February, depressions originate as far as the Atlantic Ocean and the Mediterranean Sea and periodically pass through the region of high pressure or the anti-cyclone belt created in Central Asia in the cold season. Whenever they pass over masses of water, e.g., the Persian Gulf, they carry moisture which is ultimately precipitated as rain elsewhere. There are original or parent depressions, which are formed at the meeting place between the polar and the equatorial air, and depending upon this "juxta-position of two different

air masses, having different characteristic properties, such as temperature and humidity". Then there are secondary depressions, which are formed generally "when the members of the family are moving over hilly regions or are nearing the end of their activity", and which having been thus formed, behave like other western depressions. These depressions generally move north-eastwards along a big front. "Older members dying out at one end and new ones appearing at the other".\*

The nature of such a western disturbance is three-fold, that is to say, there are three distinct weather conditions, following one another, as it moves onwards. (1) The *warm front* stage, whereby an air mass warmer and more humid moves forward and there is a sudden change of wind direction with rise of temperature and cloudy air accompanied by squally weather with passing clouds and drizzles or fog. (2) The *warm sector* stage, by which weather becomes settled down but with very high temperature and humidity. It is the steamhouse of the depression. (3) The *cold front* stage, whereby a mass of cold air enters the area and there is a sharp change of wind direction with a rise of barometric pressure, reduced humidity and sharp convectional currents. The result is squalls, thunderstorms, dust-storms and showers of rain.

These western depressions generally occur several times during a winter month and depend for their intensity upon the degree of contrast existing between the two air masses, pointed out above.

Only a portion of these cyclonic storms or disturbances affects the Indus basin and Sind. On reaching Makran, they generally split themselves into three streams covering (1) Western Highlands, (2) The Punjab and Kashmir and (3) Sind. The last stream is rather feeble and so gives quite a little rainfall in the region of our study. On crossing the Kirthar Range the storms usually develop secondary depressions which also create disturbed weather in Sind.

*Summer Weather and Eastern Depressions*—From June to August, the low pressure area extends from N.W. India to Iran and Arabia and depressions arising in the Bay of Bengal or the Arabian Sea and moving across Central India pass through this low pressure region. Like the western disturbances, these storms are also caused by the encounter of air masses of different weather conditions, e.g., the S.W. monsoon winds and the N.E. winds from the Bay. At some times they die out when resting over Central India, but at other times fresh depressions arise from them over Rajputana. "At the meeting place of the fresh Arabian Sea monsoon air and the land air which is the old and modified air from the Bay." Whichever, such

a secondary depression travels westwards with fresh vigour, there is a chance for Sind to get rains " These eastern depressions, when they are over N.W. India, induce an inflow of the Arabian Sea monsoon current, which, because of its moisture contents, maintains the activity of the depressions, whereby cloudy weather, storm winds and rainfall occur in Sind ""

Also, while moving further towards the west into the Arabian Sea off Makran, such depressions cause rough seas and squally weather over the Pasni-Karachi section

*Transition Periods*—Between these two distinct periods of winter and summer depressions, i.e., between September and November and between March and May, which are called *transition periods*, the characteristics of both the depressions are noticed,—not extensive storms, but only *localised weather disturbances* without any regular sequence of phenomena. Thus feeble or strong, local or wide-spread disturbances of weather are likely to occur in Sind throughout the year

*Cold and Heat Waves*—The passage of cold and heat waves through our province can be understood easily, as the foreign disturbances described above are their cause. As they pass over Sind, the weather invariably becomes abnormal, i.e., abnormally hot at one time and abnormally cool at another. These waves are the distinct triple-weather conditions noticed above, in connection with the depressions and disturbances. Such weather spells are frequent in the winter months, e.g., about half a dozen in January. They have a mass of warm air in their southern and eastern quadrants and cool air in their northern and western quadrants. As soon as they approach a station, there is a rise in temperature which then drops to several degrees below normal, when the depression has passed away. The weather remains abnormal, until another depression from the west approaches. Side by side with these disturbances there is good rainfall or snowfall in the localities crossed by them. At times they travel eastwards as far as Bengal, Burma and Assam and even beyond, if they are strong enough.

One or two examples of such waves may be cited here. In January, 1935, a western disturbance passed over the Gulf of Oman, the Punjab, the United and the Central Provinces causing snow or rainfall in all these areas. The depression was followed by cold waves, which affected Makran and Sind, Nawabshah recording the lowest temperature, of 27° F., while at other stations in Sind the freezing point was passed

The following minimum temperatures were recorded at Manora and Drigh Road, though the disturbance did not directly affect the two localities —

January 1935	Manora	Drigh Road	Fall of Temperature
12th	55° F	51° F	
13th	48° F.	40° F	
14th	67° F	37° F	10° F -12° F.

Notice the great difference also between the temperatures of the two places, viz., as much as 10° F., though they are only 10 miles distant

There was another instance of a heat wave, followed by a sudden drop in the temperatures, recorded at Drigh Road, during the same winter season, viz., on the 12th November 1934

Hour	11th Nov	12th Nov	Fall of T
9-15 A.M.	79° F	75° F.	1° F.
10 A.M.	85° F	79° F	6° F
11 A.M.	91° F	83° F.	8° F
12 A.M.	91° F	85° F	9° F
1 P.M.	96° F	86° F	10° F
2 P.M.	98° F	86° F	12° F
3-45 P.M.	98° F.	86° F	12° F.

Similarly, spells of heat in summer months are also usual e.g.

May 1937	Manora		Drigh Road		Departure from the Normal
	Max T	Min. T	Max. T	Min T	
16th	89° F	80° F	91° F	80° F	
17th	87° F.	79° F.	102° F	81° F	
18th	94-6° F.	80° F.	111° F	81° F	+5° F -10° F.

Such instances can be easily multiplied, the weather conditions of places under the disturbance being invariably abnormal for instance, temperatures in the Nawabshah District are very uneven for places not very far from one another. The effects are definitely localised

*C. Local Storms.*

In addition to these storms, there are other local afternoon disturbances occurring in Sind. They are due to convectional currents and contrasts of weather such as high temperature, diurnal range of temperature and differences in humidity, which cause thunderstorms, dust-storms and squally weather, whenever there is the transition stage between the two seasons, *e.g.*, (1) March to May and (2) September to November. During these months India is not affected by storms from abroad and it becomes "an independent meteorological area". They occur in Sind and the Punjab, and the time is generally in the afternoon, when these convective movements are the strongest. At times, when the influence of a passing western disturbance from the Persian Gulf synchronises with the maximum convectional conditions of the afternoon, sharp thunderstorms and squalls may occur at a station but a neighbouring station may not be affected at all.<sup>8</sup> The results are *local* dew, fog, rain, etc., while the general weather both at sea and upper air layers is more or less settled.

*Flood Years and Flood Menace*—Due to such meteorological and other conditions, Sind gets abnormal floods in some years. The following are some of the notable flood years in Sind.

Before 1900	Since 1900
1841	1902
1858	1910
1874	1913
1882	1916
1892	1921
1895	1926
	1929

These floods are caused by two chief agencies.—

1. *Meteorological causes, as stated above*

2. *Physical features and conditions of drainage*—There are a number of Nais or hill-torrents from the western highlands bringing in torrents of water down into the valley below, *e.g.*, The Gaj, the Nari. Often they flood the Larkana and Dadu Districts.

On the other hand, the annual inundations of the Indus, which flows on a bed higher in level than the surrounding country, cause floods, which prove to be disastrous.

3. *Artificial bund breaches*—Due to heavy floods and the force of torrents not only from the Nais but also of the Indus, breaches are made in the various

protective bunds, causing considerable damage to the fields and human beings. Such breaches are now controlled by the Public Works Department.

4 *Conditions in the upper reaches of the Indus*—These are the breaking of glacier dams, e.g., Shyok glacier dam, snow-melting and river-damming by landslips.

### III Climatic Divisions of Sind, the Seasons

The ancient writers divided this province of the Lower Indus Basin into three parallel zones, viz., Siro (Upper), Vachelo (Middle) and Lar (Lower) Sind, and from the point of view of climate they are found appropriate, though one is tempted at present to retain only the two chief ones, the Upper and the Lower [See Plate I(a)]

That these divisions are suitable for practical purposes can be seen from the following data [Average of 30 years (1901-1930) *India Weather Reviews*—India Meteorological Department]

Condition	Siro	Vachelo	Lar
Mean Max. Monthly T. (June 1919)	117.4° F	110.8° F (1905)	96.8° F (1901)
Mean Min. Monthly T. (Jan. 1903)	38.2° F.	47.3° F (1925)	52.2° F. (1903)
Range of T. (Seasonal)	69° F	46° F	34° F
(Monthly—May)	49° F.	26° F	10° F
Mean Humidity (Annual)	57%	61%	73%
Mean Rainfall (Annual)	3.44"	7.92"	7.51"

#### Characteristics of the Climatic Sub-Divisions.

1. *Upper Sind*—Chief locality, Jacobabad; height above sea: 186 ft.; distance from the sea: 300 miles

The dry atmosphere of the neighbouring highlands as well as the orographical features of the North West Frontier affect this region; rainfall

is the lowest in Sind and temperature is the highest (e.g., the highest temperature recorded recently is 127° F in June 1919). The air is generally very dry. There is a great range of temperature in winter and frost is common. The climate resembles the continental.

2 *Middle Sind*—Chief locality Hyderabad; height above sea: 96 ft., distance from the sea: 125 miles

The S.W. monsoon winds have in this region an average speed of 11 miles per hour in June. Rainfall is slightly more than at Karachi in some years. The temperatures are lower here than in Upper Sind and humidity is moderate. The range of temperature is also much less than that at Jacobabad. Dry hot days and cool nights are summer characteristics of this sub-division.

3 *Lower Sind*—Chief locality Karachi, height above sea: 6 feet, distance from the sea: 5 miles

The winding coastline affects this region to some extent. This coastal and deltaic area is naturally damper, with smaller ranges of temperature and pressure. The S.W. winds in summer and N.E. winds in winter are the prevailing air currents. Rainfall is at times a little less than in Middle Sind. Humidity is the greatest and the temperature moderate throughout the year. Dampness causes muggy or oppressive weather in summer. The climate is maritime.

*Seasons*—In an arid sub-tropical region such as this, there are only three main seasons of nearly 4 months each.

- 1 S.W. monsoon season, from July to October (the retreating monsoon does not affect it appreciably)
- 2 Cold season from November to February
- 3 Dry hot season from March to June

In (1), the rains are not general as in Peninsular India, but only occasional, the wind direction is S.W. or S.E. and humidity is on the increase. The 15 days monsoon period is 9th July to 23rd July.

In (2), the rains are only cyclonic storms and the usual direction of the wind is N or N.E. (Trade wind). When it does not rain and the sun shines, the weather is fine and bracing. It gets cooler and cooler as we go from south to north from an average of 60° F at Karachi to 40° F at Jacobabad. This is the touring season of India.

In (3), the temperature of the air gradually rises to the maximum in June-July. The sun shines directly over Sind and the wind direction changes from N.W. to S.E. The isotherm of 85° F. passes through Karachi, while round about Jacobabad the mean temperature is 95° F.

*IV Chief Climatic Factors and Elements*

It is now necessary to consider the chief climatic data that are available, viz., temperature, barometric pressure, relative humidity, wind system and rainfall.

### 1 Temperature

Sind is uniformly very hot in summer and comparatively cool in winter, as the records of 30 years (1901-1930) clearly show (See Plate II)

Sind derives its heat from the following sources —

- (i) The sun, which shines for the greater part of the year in cloudless skies
- (ii) The rocks which are barren and the soil which is arid, sandy and saltish and which absorbs and radiates heat quickly
- (iii) The warm front of the wave of a cyclonic storm approaching Sind
- (iv) The warm N W winds from the Iran Plateau in summer

At times, there blows a harmful wind analogous to the Simoon of Arabia and the Sahara, while dust-storms with whirlwinds are common in the hot weather period. The eddies, as much as 10 ft wide in dusty areas, rise to a height of some 200 ft

There are at the same time many ways in which heat is lost and cold ensues

- (i) The air is still and the sky clear and there is little humidity or water vapour, in it to absorb heat
- (ii) The rocks being barren and not covered with grass, there is quick radiation of heat towards the close of the day
- (iii) Cold winds from the snow mountains in the north and the north-west blow during winter and reduce the temperature of Sind to a great extent

The month of January, for example, may be very cool from cold winds in one year and from nocturnal radiation of heat in another

*Mean Maximum Temperature* — The graphs drawn, for the mean maximum temperature, show that there is a steady rise of temperature at Karachi, from 76° F in January to 91.6° F in June, which is the hottest month in the year. It then begins to fall upto 86.7° F in August and remains more or less steady in September; then after a slight rise in October, it falls rapidly for November, December and January.

The graph for Hyderabad differs slightly from that for Karachi, in that the maximum temperature is reached in May, with a rapid fall up to

August In September and October there is a gentle rise and then a sudden fall again for the winter months of November, December and January.

Jacobabad is more irregular than either of the above stations. After the fall in winter there is a very steep ascent in February and March and a slight rise in April, then again a sudden rise for May till the maximum is reached in June. The fall after June is also sudden and rapid with practically no rise in October, as in Middle and Lower Sind (See Plate II)

The average annual maximum temperature for Karachi is 85.5° F., for Hyderabad 93.3° F and for Jacobabad 95.5° F

*Mean Maximum T.  
Average of 30 years (1901-1930)  
(India Weather Reviews )*

Month	Karachi	Hyderabad	Jacobabad
January	76.0	75.8	72.7
February	77.7	81.4	77.5
March	83.1	92.5	90.7
April	87.0	101.2	101.8
May	90.2	107.0	111.3
June	91.6	104.6	113.7
July	89.6	99.6	108.5
August	86.7	95.5	103.9
September	86.8	96.7	102.9
October	88.5	97.5	98.8
November	86.1	88.9	87.3
December	79.3	78.2	75.5
Annual	85.5	93.3	95.5

*Mean Minimum Temperature*—The graphs of the mean minimum temperatures of the two stations of Karachi and Hyderabad are smoother than those for Jacobabad. The coldest month for all three places is January

The fall of temperature is greater at Jacobabad in September, and again in November and December than at the other two stations

The annual average minimum temperature at Karachi, comes to  $71.2^{\circ}$  F., at Hyderabad to  $68.7^{\circ}$  F. and at Jacobabad to  $66.4^{\circ}$  F. only

*Mean Minimum T*  
Average of 30 years (1901-1930)  
(India Weather Reviews)

Month	Karachi	Hyderabad	Jacobabad
January .. ..	56.7	50.5	44.1
February .. ..	59.5	54.0	49.1
March .. ..	67.3	63.5	60.5
April .. ..	74.1	71.6	70.6
May .. ..	78.9	78.1	78.1
June .. ..	82.1	82.0	85.1
July .. ..	81.3	81.7	85.6
August .. ..	78.5	79.6	82.5
September .. ..	76.9	76.3	70.0
October .. ..	73.5	70.8	65.7
November .. ..	65.3	60.5	53.5
December .. ..	58.8	52.1	44.7
Annual .. ..	71.2	68.7	66.4

The above table shows that the coastal strip has a more equable climate than places further inland, Jacobabad faring the worst in this respect

*Fluctuations in the Three Decades (1901-1930)*—The graphs for the average temperatures (maximum and minimum) for the whole year indicate that they were more or less steady in all the decades for Hyderabad and Jacobabad, while those at Karachi slightly fluctuated. The following are the mean (mean of 30 years 1901-1930) temperatures at the three places:—

Station	Max T	Min T.	Average
Karachi	85.5° F	71.2° F.	78.3° F
Hyderabad	93.3° F.	68.7° F	81.0° F.
Jacobabad	95.5° F	68.4° F	80.9° F.

*Range of Temperature*—Both as regards the daily and monthly ranges of temperature, Hyderabad and Jacobabad differ from Karachi, where the range is small, owing to the influence of the sea. But even at a little distance beyond the coast-line, e.g., Drigh Road, it increases considerably.

The accompanying thermograms (see Plate II) of Karachi show the daily range near the sea very clearly. The daily range in summer months is much smaller than in November and December. The day's temperature is the highest in the afternoon, about 2 P.M., and the lowest in the early morning at 5 A.M.

At Drigh Road the daily range is upto 35° F, at Hyderabad upto 40° F and at Jacobabad upto 45° F.

The following table gives an indication of the monthly ranges of temperatures in Sind.

*Mean Monthly Range of Temperatures*

Month	Karachi	Hyderabad	Jacobabad
January	27.1° F	25.3° F	42.8° F
February	19.8° F	26.5° F	49.5° F
July	9.0° F	17.8° F	36.7° F.
November	5.7° F.	28.5° F.	50.4° F

(*Gazetteer of the Province of Sind B Vols*)

The monthly range of temperature increases from the coast northwards till near the Upper Sind Frontier it goes to over 50° F in winter. It is also greater in winter than in summer. Owing to the SW monsoon current the range of temperature near the coast is not so great in summer.

### 2 Barometric Pressure

Indirectly due to these temperatures, the barometric pressure is the lowest at Jacobabad and the highest at Karachi. The difference between the lowest pressure in July and the highest in December is (1) at Karachi, .568 inches; (2) at Hyderabad, .621 inches and (3) at Jacobabad, .670 inches, as the following table of monthly average pressure shows —

*Barometric Pressure*  
Average of 30 years (1901-1930)  
(*India Weather Reviews*)

Month	Karachi	Hyderabad	Jacobabad	Remarks
January	30.090	30.006	29.921	
February	30.062	29.941	29.817	
March	29.935	29.834	29.733	
April	29.831	29.710	29.597	
May	29.723	29.589	29.460	
June	29.560	29.432	29.295	
July	29.522	29.391	29.260	Lowest in the year
August	29.599	29.467	29.353	
September	29.747	29.619	29.506	
October	29.902	29.796	29.690	
November	30.024	29.935	29.852	
December	30.000	30.012	29.930	Highest in the year.
Annual	29.830	29.728	29.621	

### 3 Relative Humidity

There is a great divergence also in the relative humidity of the three stations. The difference between Karachi and Jacobabad amounts to 35 per cent in the months of April and May. The driest month in Karachi is December, whereas in Hyderabad and Jacobabad it is April.

due to there being more winter showers. The greatest amount of humidity, however, is in August at all three stations due to the influence of the S.W. monsoon current. Hyderabad is distinctly drier than Karachi, but Jacobabad is drier still.

*Average Monthly Humidity*

Average of 30 years (1901-1930)

(*India Weather Reviews.*)

Month	Karachi %	Hyderabad %	Jacobabad %	Remarks
January	61	61	65	
February	67	58	54	
March	71	53	45	
April	76	52	41	Lowest in Upper and Middle Sind.
May	78	55	43	
June	79	64	57	
July	82	69	65	
August	81	72	71	Highest in all Sind.
September	82	70	68	
October	74	61	56	
November	62	55	56	
December	58	59	63	Lowest in Lower Sind.
Annual	73	61	57	

Thus in the interior the air is dry, though hot, in summer and hence not so distressing as on the coast; after sunset the humidity is reduced for a while, the heat from the ground radiating quickly, and nights are therefore cool. As there is a greater range of temperature in winter, there is also a greater increase or decrease of humidity.

*Frost*—Some parts of Sind are visited by frost in winter, the necessary conditions being fulfilled, viz., (1) a clear sky, so that radiation is not impeded by clouds and (2) a calm night, when the layers of air in contact

with the ground are cooled. Whenever the temperature of the ground falls below the freezing point, the air in contact with it is so much cooled that the water vapour in its layers immediately above it or in the voids among the soils is condensed into ice particles or frost. This kind of frost is found frequently in Upper Sind, but rarely in Lower Sind. Some years are notable for severe and prolonged frost, causing great damage to crops, e.g., 4th January to 4th February 1934.<sup>10</sup>

**Cloudiness**—The skies in Sind are cloudy during the monsoon season and also during the months of December and January, but in other months they are very clear. As a rule, afternoons are more cloudy than the rest of the day.

#### 4 Wind System

Average of 20 years (1901-1920)  
(*India Weather Reviews*)

Season	Wind Direction	Karachi Days	Hyderabad Days	Jacobabad Days	Remarks
Cold Winter	N W	35	26	29	(Predominantly calm weather in Upper Sind)
	Calm	25	42	173	
	N	33	69	21	
Dry Hot	N.E.	69	8	22	
	E.	16	2	24	
	S.E.	4	5	62	
Monsoon	S.	3	65	24	(Force of S.W. monsoon in Middle Sind)
	S.W.	50	134	8	
	W	130	14	3	
Total No. of Days..		365	365	365	

The above data show that the predominant wind direction in Karachi is westerly for over four months of the year. During the months of May to September, the general wind direction is S.W. and during the winter months of January, February, November and December it blows from the N.E., while in the intermediate months, before and after the summer season, the direction changes to N.W. It rarely blows from the south or east.

At Hyderabad the S.W. direction is the most conspicuous from April to October, while in winter the N.W. direction prevails.

At Jacobabad there is practically no wind for half the year. During the other half the direction swings from S E to N E in the hot season and from N E to N W in winter [See Plate I (a)]

The influence of the S W monsoon wind reaches nearly a hundred miles inland and so evenings are generally cool at Hyderabad in summer

*Wind Velocity (Miles per Hour)*

Average of 20 years (1901-1920)

(India Weather Reviews)

Station	Jan.	Feb.	Mar.	Apr.	May	June	July	Aug.	Sep.	Oct.	Nov.	Dec.	Annual
Karachi	0.6	7.4	9.2	11.4	13.1	14.0	14.9	13.7	10.7	7.5	5.9	5.2	10.3
Hyderabad	4.9	4.6	5.1	6.8	8.5	11.3	10.5	10.0	8.5	5.2	3.9	4.7	7.3
Jacobabad	1.7	2.3	3.1	3.5	3.7	4.4	4.7	3.8	3.3	1.8	1.3	1.3	2.9

The above table shows that near the seashore the wind velocity is greater, while in Upper Sind it is generally calm for nearly half the year. During the cold weather months the wind velocity varies from about 6 miles in Lower Sind to only about 1.5 miles in Upper Sind. It begins to increase from March and April, until it reaches its maximum velocity of about 15 miles in Lower Sind, about 11 miles in Middle Sind but only about 4-5 miles in Upper Sind. With the occurrence of dust storms and cyclonic rains, the wind velocity is naturally higher.

*Local Land and Sea Breezes* —No special study of winds prevailing locally at the different centres has yet been made except at Karachi.<sup>11</sup> Recently owing to the requirements of the Dighi Road Airship Base and Aerodrome and as a result of the facilities available at the Meteorological Observatory there, Dr. Ramdas<sup>12</sup> has produced a valuable paper on the local afternoon sea breeze at Karachi, occurring during the non-monsoon period of October to March. This breeze is calculated to be a result of the difference in temperature and pressure above land and sea and due to the difference in solar insolation. His finding is that "The breeze sets in during the afternoon and has some of the characteristics of a cold front. It sets in suddenly, is usually colder, more moist and has a greater velocity than the preceding land breeze." Thus land breeze, again, is part of the prevailing winds at lower levels over the whole of Sind, the normal direction being N W and N E and the actual direction and strength of the winds are determined by the position and intensity of the seasonal anticyclone over North West India.

The general characteristics of the breeze have been summarised as under : " The sea-breeze first sets in at Manora, then moves towards Drigh Road through the Karachi City. The time taken by the sea breeze to reach the Airship Base varies from 1 to 3 hours. Before the arrival of the breeze at the Airship Base, the previous land breeze, if any, drops to calm and the setting in of sea breeze is accompanied by a fall of temperature (about 5° F to 3.5° F ), rise of humidity (from 5% to 30% and above), shift of wind direction to west-south or south-west, and an increase of wind velocity . The transition from land breeze to sea breeze is more and more marked as the latter proceeds inland. The sea breeze front appears to be somewhat diffuse near the coast but by the time it reaches the Airship Base it becomes quite sharp owing to increased contrast with the land-breeze "

It has been also noticed that this sea-breeze is not the actual gradient wind, i.e., westerly and north-westerly breeze but owing perhaps to the hills to the north-west and north of Karachi City, it is deflected largely to the west-south-west and south-west direction

Thus it can be seen that in the climatic elements of temperature, barometric pressure, humidity and wind system, Sind in general has three distinct regions differing from one another. How far precipitation agrees with this analysis has to be seen now

### 5 Rainfall in the Lower Indus Basin

Sind is called an " Unhappy Valley ", perhaps on account of its scanty rainfall. "The Hindus of Sind," says R. F. Burton, "have a tradition that Hiranyakasipu, the demon king of Multan, was powerful enough to draw down Megha Raja, the cloud god from heaven and compel him to promise never to visit the valley of the Indus "

The orographic features of Sind lend support to such a belief. The average height of the surrounding hill ranges is only about 5,000 ft., while the Laki Hill lying in Kohistan just reaches a little above 2,000 ft. To catch the monsoon current a high mountain within the region is required. Again the alignment of the two ranges of mountains bordering the frontier of Sind, is not straight. There is a marked re-entrant angle in the north-west, which is very low. Any clouds that are caught by storms are again lost in them. Particularly in the case of Sind in winter, "the seat of maximum pressure is not the seat of maximum rainfall, nor indeed is the case of the great summer depression of Upper Sind and Lower Punjab of any precipitation whatever "<sup>14</sup>

But though the rainfall is meagre, it is a useful supplement to underground supply of water in many parts of Sind

*Annual Rainfall*—Being a portion of the *dry* monsoon belt, this region receives very little rainfall, it has very few rainy days, during the year, year after year, the average rainfall at any station, not amounting to 10 inches, as the following table shows—

*Rainfall Year by Year.*  
Average of 30 years (1901-1930)  
(*India Weather Reviews*)

Year	Karachi (Inches)	Rainy Days	Hyderabad (Inches)	Rainy Days	Jacobabad (Inches)	Rainy Days
1901	1.37		1.96		2.68	
1902	18.23		15.93		2.10	
1903	4.58		5.72		3.22	
1904	4.62		1.41		2.68	
1905	3.64		2.62		2.21	
1906	6.47		7.45		5.57	
1907	7.75		6.60		4.44	
1908	6.45		19.93		4.59	
1909	7.09		7.03		1.41	
1910	12.63		10.19		3.84	
1911	4.84		3.79		1.36	
1912	3.17		5.39		1.41	
1913	13.45		21.13		4.86	
1914	9.31		4.09		6.78	
1915	2.26		1.01		1.34	
1916	21.87		14.01		3.80	
1917	5.63		9.81		10.88	
1918	2.04		1.74		1.36	

## Rainfall Year by Year—(Contd.)

Year	Karachi (Inches)	Rainy Days	Hyderabad (Inches)	Rainy Days	Jacobabad (Inches)	Rainy Days
1919	3.39		7.76		3.23	
1920	1.97		1.70		2.26	
1921	16.90		13.78		3.58	
1922	1.99		2.66		0.13	
1923	5.57		5.21		4.73	
1924	3.69	9	12.07	10	2.11	5
1925	4.38	7	1.02	2	4.54	10
1926	20.04	11	12.11	13	4.28	10
1927	8.90	10	13.55	12	0.67	3
1928	2.39	5	2.14	5	0.90	3
1929	4.13	8	23.13	14	6.55	12
1930	16.70	12	2.77	6	5.67	4
Average ..	7.51	9	7.92	9	3.44	7

A special feature of the rainfall curves in Sind is that there are peaks of good rains and troughs of droughts at an interval of 5-6 years (See Plate III.) It is remarkable that while there are a number of years of great scarcity of rainfall, there are years of abnormal rains and floods. Also, these do not always coincide in all the three climatic divisions, e.g., while 1926 was a flood year for Karachi, there was only 4.28 inches of rainfall at Jacobabad. In 1929, Hyderabad received 23.13 inches but Karachi only 4.13 inches. Nearer the coast, rainfall is greater than in Upper Sind, where the average is only 3.44 inches.

*Distribution of Rainfall*—In the matter of distribution of rainfall in Sind, Nagar Parkar stands by itself. Not only does it receive the advantage of its height but it touches, to some extent, the fringe of the S.W. monsoon current. Middle Sind receives slightly more rain than Lower Sind in some

years while there is hardly any rain (less than 2 inches) in parts of Upper Sind. Even in years of great floods there is not more than 10-12 inches of rainfall in this the driest part of the Province.

*Mean Annual Rainfall  
(India Weather Reviews)*

	Place	Mean Annual Rainfall	Rainy Days
	Nagar Parkar	14.85	17
Lower Sind	Tatta	8.83	8
	Karachi	7.51	9
Middle Sind	Umerkot	8.96	12
	Hyderabad	7.92	9
	Mirpurkhas*	9.55	9
	Nawabshah*	6.05	7
	Naushero	4.26	6
	Sehwan	5.41	8
Upper Sind	Sukkur*	2.74	5
	Shikarpur	3.39	6
	Larkana*	3.55	5
	Jacobabad	3.44	7

Stations marked \* have the averages taken for 17 years (1914-1930). The rest have those for 30 years (1901-1930).

At Sukkur the average rainfall is the least, *i.e.*, less than 3 inches, while at Nagar Parkar it is the heaviest, *i.e.*, nearly 15 inches. At the intermediate stations it varies from about 5 inches to 10 inches. Also the average rain per day is a little less than 1 inch, the total number of rainy days amounting to 7 only (See Plate III).

*Mean Monthly Rainfall*—Though there are two distinct seasons of rainfall, *viz.*, summer and winter, the following table shows that, on the whole, it is not only precarious and fitful, in both of them, but also scattered thinly throughout the year. The amount is, however, more in summer than in winter. The actual monsoon period during the year is from 9th July (commencement) and 23rd July (withdrawal) (See Plate IV.)

*Mean Monthly Rainfall*  
Average of 30 years (1901-1930)  
(India Weather Reviews)

	Jan	Feb	Mar	Apr	May	June	July	Aug	Sept	Oct	Nov	Dec	Annual
Nagar Parkar	0.03	0.15	0.09	0.01	0.27	0.64	6.18	4.52	2.49	0.39	0.04	0.04	14.85
Tatta	0.11	0.37	0.21	0.04	0.21	0.61	4.48	2.09	0.58	0.02	0.04	0.03	8.83
Karachi	0.31	0.46	0.34	0.07	0.10	0.57	2.76	1.95	0.78	0.03	0.10	0.15	7.51
Umerkot	0.09	0.14	0.17	0.01	0.17	0.36	3.79	2.72	1.34	0.13	0.01	0.03	8.96
Hyderabad	0.12	0.30	0.28	0.08	0.15	0.36	3.43	2.12	0.93	0.02	0.03	0.10	7.92
Mirpurkhas*	0.08	0.07	0.10	0.18	0.23	0.28	1.86	2.87	0.71	0.04	0.05	0.08	9.55
Nawabshah*	0.07	0.08	0.07	0.03	0.16	0.19	1.96	2.47	0.87	0.06	0.04	0.05	6.05
Nashero	0.15	0.22	0.18	0.03	0.15	0.30	1.20	1.21	0.67	0.00	0.02	0.13	4.26
Sehwan	0.23	0.27	0.14	0.11	0.08	0.28	2.14	1.22	0.72	0.00	0.03	0.16	5.41
Sukkur*	0.04	0.15	0.14	0.03	0.17	0.06	0.86	0.90	0.18	0.05	0.05	0.11	2.74
Shikarpur	0.14	0.22	0.25	0.06	0.07	0.21	0.72	1.02	0.46	0.04	0.01	0.17	3.39
Larkana*	0.05	0.14	0.14	0.08	0.10	0.21	0.98	0.93	0.66	0.06	0.03	0.17	3.55
Jacobabad	0.23	0.20	0.26	0.20	0.13	0.36	0.82	0.70	0.28	0.06	0.02	0.18	3.44
Average for Sind	0.12	0.21	0.18	0.07	0.15	0.33	2.63	1.90	0.82	0.06	0.03	0.10	6.65

Stations marked \* have their averages taken for 17 years (1914-1930). The rest have those for 30 years (1901-1930).

*Rainfall in Kohistan and the Desert Province*—In the matter of the distribution of rainfall in Sind the data for Kohistan are very important, as they give an indication of the periodic supply of rain water in the Malir Basin, which is the main source of Karachi's water supply. Here too the peaks of good rainfall alternate with the years of drought. At Khadeji the record is as follows—

Year	Khadeji R.G. No. 2 Inches
1914	5.02
1915	3.53
1916	15.60
1917	11.48
1918	0.62
1919	10.05
1920	1.83
1921	12.17
1922	3.96
1923	6.61
1924	5.32
1925	0.75
1926	7.75
1927	8.19
1928	2.94
1929	0.37
1930	8.25
1931	1.24
1932	10.86
1933	15.70
1934	6.29
1935	4.40
Average	6.89

(Data supplied by the Office of the Karachi Municipality)

Thus, though the rainfall in Kohistan is usually scanty, there are years in which it increases to over 15 inches, e.g., in 1916 and 1933. It is even more at Khadeji than at Karachi in some years. There is, therefore, plenty of underground water secured by percolation and absorption in the deeper strata of rock in the dry river basin. (See Appendix III.)

Even in the region called the Thar Desert, there are good, though occasional showers and years of floods. The following is the average rainfall in the different Talukas of Thar Parkar.—

Average rainfall (average of 55 years)	Bunjaro	Mirpur khana	Jamesabad	Dighri	Khopro	Samro	hanghri	Umerkot
1910-1935	8.49	9.02	9.30	9.16	7.99	13.5	6.59	9.39

(Data from P W D, Sind)

A portion of this rainfall is due to the winter showers, which account for water existing in the number of Dhands or lakes in the Thar and the Pat Sections and affording camel pasture in the cool season

*Reasons for Scarcity of Rainfall*—Apart from the fact that Sind usually escapes the influence of both the monsoons, there are other reasons of such scarcity or irregularity of rainfall<sup>14</sup>

(1) In the first place, the arrival of the S W monsoon is rather late in Sind and the season is short. It does not burst here in May-June, when the land is the hottest (mean maximum temperature at Jacobabad being  $113.7^{\circ}$  F), but in July when the temperature is lower (mean maximum temperature being  $108.5^{\circ}$  F), and the chief pressure gradient is over the Peninsular and the Bay of Bengal. The reason is that the upper air current over N W India, flowing from the west and north-west during the season is warm and dry with low humidity and this *descends a couple of thousand feet against the S W monsoon current*

(2) The northern parts of India get very cold in winter, the Himalayan snows melt slowly in early summer and so the high pressure over the area *takes much time to turn into a low pressure system*, so essential for the necessary undraught. Hence the delay in the actual arrival of the rainy season in the Indus valley and the reduction in the number of rainy days. Also, the more the snowfall in one winter season on the mountains, the poorer the prospects of rains in the plains in the following summer

(3) As the monsoon season advances, and the lowest pressure is well within the angle between the Kirthar and the Himalayan Mountains, the air can only flow in here from the south and east and so *only a portion* of the monsoon current containing a little moisture can pass over the Province

(4) The land being exceptionally and increasingly hot from south to north, *e.g.*, mean T  $85^{\circ}$  F at Karachi to  $95^{\circ}$  F at Jacobabad, the humidity is *relatively reduced*, from 80% to 50% only. The high temperature is also further increased on account of the dry hot land and the absence of low clouds.

(5) The air current over the N W. *must rise some 3,000 ft above the sea-level* before any appreciable precipitation can take place. Such is not the case in Sind, and the upper layers, being warm and dry, do not

allow formation of low clouds over the land. While in North India and in the Ganges basin rainfall increases as we approach high mountains, in the Indus basin the reverse is the case, precipitation decreasing considerably on the western highlands.

*Variability of Rainfall*—Not only is the rainfall in Sind scarce but it is also irregular, so that for every one season of good rains, there are two or three or even more of drought. At times, again a whole season's rain falls within a couple of days and there is prolonged scarcity for the rest of the year. The result is terrific floods and destruction of crops at one time and severe drought at another. "Over the Indo-Gangetic Plain, variability\* is at a maximum in Central Sind, which is at once the seat of the most niggardly and also the most variable rainfall in the whole of India. The noticeable improvement in reliability is of little, if any, economical significance, for throughout Sind, Bhawalpur and Western Rajputana a variability of over 30 per cent combines with an average rainfall, which is nowhere greater than 15 inches and for the most part is considerably less."<sup>11</sup>

In Sind and Cutch the annual percentage deviation from the average is  $\pm 37$  (mean) + 124° (max) - 53° (min), the most unique in the whole of India.

Following table of comparative variability is significant.

Station	Average Annual Rainfall	Variability
Naoshera (Sind)	5"	53%
Cawnpore (U P)	34"	20%
Calcutta (Bengal)	65"	11%
Rohri (Sind)	6"	65%
Sirsa (Punjab)	12"	46%
Lucknow (U P)	35"	28%
Darbhunga (Bengal)	44"	20%
Barisal (Bengal) ..	62"	16%

\* Variability of rainfall is the degree in which the rainfall of any given year is likely to deviate from the local average either in excess or defect (Blandford).

This shows that while the normal yearly rainfall increases from a minimum in Sind to a maximum in Bengal, the variability is the reverse. "The province in which the annual variations as compared with the general average are greatest is Sind and Cutch."

In this region, again, there is variability of *summer rainfall* of 40 per cent to 80 per cent from the desert in the east to the highlands in the west [See Plate I(b)]

Even during the different months of the monsoon season, the variability is different in different regions of the Indo-Gangetic Plain, the worst sufferer being Sind —

Month	Region I	Region II	Region III A	Region III B	Region IV
June .	> 70	65-75	60-75	60-75	45-60
July	> 60	45-75	10-50	40-50	30-40
August	> 70	40-70	40-50	10-50	35-50
September .	> 90	> 80	60-75	60-75	15-60
October	> 100	> 100	> 100	> 100	> 75

Thus the rainfall in the Lower Indus basin is not only scanty but also irregular and unreliable; so one cannot speak of *normal* rainfall figures for any station in Sind. "Within the Middle and Lower Indus lowlands, the so-called *normal* rainfall figures are anything but indicative of conditions from year to year."

In the case of Karachi over a period of a third of a century, the normal rainfall was 6.3 inches though half the season's precipitation was no more than 4.3 inches. In three seasons, the rainfall amounted to a little over 20 inches.

#### V Cyclical Period

This leads us to the consideration of a cyclical period in Sind. There is no doubt that the floods come periodically but to establish a cycle of a certain number of years is a most difficult meteorological problem here. Blanford established a cycle of 11 years from certain meteorological calculations which, however, do not tally with the data collected in this region in recent years. He marked the relationship between weather and sunspots, the maximum of which occurred at intervals of 11 years. "But the actual amount of surface covered at each maximum is very irregular."

Another argument advanced in favour of a cycle of 11 years is that it tallies with the main daily range of magnetic declination, according to Prof. Balfour Stuart. But in this case also we have to deal with very irregular curves. It may be that there are other unknown influences working out a cyclical period. The temperature and rainfall in a particular area are caused by storms and depressions, which cannot be accurately gauged or forecast at present. But it can be said with a certain amount of accuracy that any alteration in the solar heat may *modify* the formation of cyclones and so a cyclical period.

There is, again, the possibility of connecting years of drought with the years of heavy snow-fall on the Himalayas, as too much snow may cause prolonged suspension of summer rainfall. Droughts may also be due to unseasonable persistence of dry land winds. Another strange coincidence is that a drought in the Peninsula is followed by one in North India, e.g., 1876 Deccan drought, 1877 Punjab drought.

So far as the region of our study is concerned, a cycle of 11 years is quite out of the question. In very recent years a cycle of 6-7 years is noticeable, viz., 1902, 1908, 1910, 1913, 1921, 1926, 1932 (See Plates III & IV).

#### VI Climatic Change

Is, then, the climate of Sind also changing? There are several records to show that Sind experienced a better and more congenial climate in prehistoric days. "It is related by the chronicles of antiquity that in days gone by, in ages that have long fled, Sind was a lovely land, situated in a delightful climate, a fertile plain, traversed by the beneficent Mihran with large flourishing and populous cities, orchards producing every kind of tree and fruit, and gardens that were the reflections of Iran and the envy of the seven Heavens."<sup>19</sup>

The remains of Mohen-jo-Daro and of other kindred sites in Sind distinctly indicate a wetter Sind in the pre-Aryan period.

The admirable researches of Sir Aurel Stein<sup>20</sup> in Baluchistan and Waziristan show that the climate of Central Asia must have been in the past more favourable, especially in Gedrosia where there were impenetrable forests.<sup>21</sup>

Again, a favourable climate is an essential condition of high civilisation. This has led Sir John Marshall<sup>22</sup> to believe that in the chalcolithic age of Mohen-jo-Daro the rainfall in Sind must have been 15-20 inches average annually. He thinks that the northern storm belt was then deflected by arctic pressure further south. At the same time a deflection of the S.W. monsoon current is also suggested. On the whole, all the dry Afrasian belt was covered by westerly rain storms in that age.

Dr. C. W. Normand, Director of the Meteorological Department of India, has opined that more rain in the summer season was possible in Sind and Baluchistan at the time of the Indus Valley Civilisation, "as a very much greater change in meteorological conditions is required to explain a copious rainfall in winter, unless a change in the orography of Sind's surroundings is simultaneously postulated."<sup>22</sup>

Mr. Majumdar is also of opinion that from the presence of many drains and pipes and burnt bricks of Mohen-jo-Daro, figures on seals of animals living in moist climate such as rhinoceros, tigers and elephants and of trees on painted pottery, it can be surmised that Sind must have seen wetter years in the millenniums before Christ.<sup>23</sup> Remains of elephants and rhinoceros are actually found on the eastern side of the Kirthar Range.

Burnt bricks and not baked ones were used in Sind in those prehistoric times. Even in the remains of Bahmanabad discovered by Bellasis, such burnt bricks were used suggesting greater rainfall.<sup>24</sup>

Raverty has noticed in *Arabian geographers* a reference to the green (*Hariana*) and cultivable lands now lying barren.<sup>25</sup> A big forest between Larkana and Khampur in Larkana District was actually utilised by the Daud-potras for their Shikargahs.

That the climate of Sind has become gradually drier to-day than at the time of Mohen-jo-Daro is also attested by Dr. Mackay from the rapid deterioration of bricks, which is now taking place and those beautifully preserved in the buried city. "This state of things was due to the fact, that the plain round the city was then well watered and consequently the destroying action of the salt was checked, although as we know the buildings needed frequent repairs."<sup>26</sup>

In this matter of climatic change, the climatic tables, of two sets of normals for the three stations Karachi, Hyderabad and Jacobabad, covering two long distinct periods, are also suggestive. (See Appendix II)

Whatever the meteorological conditions of Sind may have been in the past ages, the rainfall recorded for the past five years does not give any indication of a change of climate. The year 1933 was only a flood year for Karachi while in 1935 and 1936 the rainfall at the station was decidedly poor, while Hyderabad has fallen below the average.

There is a suggestion made by some writers that the Sukkur Barrage which is fully functioning since 1932, would help Sind to secure more rains. But such an expectation is too premature. Considerable data<sup>27</sup> must be collected and more time must elapse before the meteorologists of Sind can come to any definite conclusion. It is not impossible that more extensive water

*Latest Rainfall Statistics  
India Weather Reviews (1929)*

Year	Karachi	Hyderabad	Jacobabad
1931	0.73	2.01	1.11
1932	12.78	4.33	5.24
1933	20.11	8.60	5.71
1934	8.07	8.35	5.97
1935	4.51	3.29	5.30
1936	3.73	4.24	3.61
Average	8.32	5.13	4.49

supply, perennial irrigation, afforestation and continuous evaporation may cause slightly greater precipitation of rainfall in Sind in the years to come. Says Blanford "Of the influence of local evaporation in enhancing rainfall, one or two instances are given especially that of Yarkand and that of the irrigated tract bordering the Indus in Sind and the lower Punjab."

*Summary*

In no other region in India are the climatic conditions more unique, irregular and unstable than in the Lower Indus basin. It is the driest and hottest of all Indian provinces, aridity being its commonest feature, e.g., Jacobabad has a mean maximum temperature of 113° F and mean humidity of 41 per cent. The thermal equator passes through Sind. These conditions are accentuated by its physical features, the Thar Desert in the east, the sea in the south and the Hala-Suleiman (Kirthar) Mountains with their peculiar re-entrant angle, in the west and northwest. Thus the temperatures are exceptionally high in summer but in winter they are comparatively low. The diurnal range of temp. rature is also great (e.g., over 45° F in a single day at Jacobabad). Nearest the sea coast, the daily range of temperature is less. The monthly range increases from south to north and from summer to winter (e.g., Jacobabad has a summer range of temperature 42° F and winter range 50° F). The greatest humidity occurs in August, viz., 75, but the least in December, viz., 58 in Lower Sind and 47 in April in Upper Sind. Thus the weather is drier and hotter but more bearable in the north than in the south. The skies are generally clear and frost is not uncommon.

While in Upper Sind it is generally calm for nearly half the year, nearer the coast of Sind, the wind velocity is about 15 miles per hour in the monsoon season. Dust storms and squally weather are common in the beginning of the two chief seasons. The predominant wind direction at Karachi is westerly before the monsoon season begins, but in winter the direction is north-east or north-west. In Upper Sind, however, it varies from south-east to north-east in summer and from north-east to north-west in winter.

In the matter of rainfall, the region suffers the most. It is not only precarious and scanty but also the most variable in India, the orography of Sind being mainly responsible for it. What little precipitation it gets is often due to cyclonic storms, caused by eastern and western disturbances, particularly the former. The average rainfall is about 7 inches in Lower Sind and about 3 inches in Upper Sind, with only about 8 rainy days in the whole year, July being the rainiest month. The greatest peculiarity about the rainfall curve is that after gaps of six or seven years of scarcity, there are peaks of good and at times heavy rainfall, though the cyclical period is hard to determine. It is a question whether there is any real progressive climatic change in Sind, as the history of the Province shows that there might have been greater rainfall in the past when forests existed. It is equally difficult to say whether the Sukkur Barrage drainage system would influence future rainfall. Even with perennial irrigation, afforestation and continuous evaporation it is not certain whether Sind would see wetter days.

\* \* \*

The paper is a continuation of the *Geographical Analysis of the Lower Indus Basin (Sind)*, already published in the *Proceedings*\*. It is illustrated with necessary graphs, sketch maps, etc.

Reliable climatic data for many stations in Sind are hard to obtain. For the present Paper they have been collected from the *Gazetteer of the Province of Sind*, and from the publications of the India Meteorological Department, e.g., *India Weather Reviews*, access to which was kindly allowed by the Meteorological Office, Karachi, also from the records of the Development and Research Department, P W D, Sind, and the Office of the Karachi Municipality, by the courtesy of the chief executive officers.

My thanks are due to Professors E. G. R. Taylor, Dr H. A. Matthews, both of the University of London and to Professor D. N. Wadia, of the Geological Survey of India, for their guidance and helpful suggestions.

## APPENDIX I

*General Weather Conditions in Sind Month by Month*

## January

Coldest month of the year  
 Western disturbances affect the region  
 Cold waves, rain, squally and northerly wind expected  
 Dust storms and frost also are likely  
 Mean temperature-- Max 74.8° F , Min 50.4° F  
 Mean rainfall-- 12 inches  
 Mean Baro pressure-- 30.005  
 Mean humidity--60%  
 General wind direction- N E , N W

## February

Meteorological conditions similar to those in January  
 More windy month--Chilly weather  
 More westerly depressions--Rain expected  
 Mean temperature-- Max 78.8° F , Min 54.2° F  
 Mean rainfall--21 inches  
 Mean Baro pressure--29.950  
 Mean humidity--59%  
 General wind direction, N E , N W

## March

Dry weather, clear or partially clouded skies  
 Rather dusty month  
 Early mornings foggy near the coast  
 Western disturbances still occur with scattered showers  
 Mean temperature-- Max 88.7° F , Min 63.7° F  
 Mean rainfall--18 inches  
 Mean Baro pressure-- 29.835  
 Mean humidity--50%  
 General wind direction--N W

## April

Winter conditions disappear and summer conditions appear  
 S W wind begins to blow across the region  
 Dust storms at times, practically no rainfall  
 Mean temperature-- Max 96.6° F , Min 72.1° F.  
 Mean rainfall-- 7 inches  
 Mean Baro pressure--29.712  
 Mean humidity--50%  
 General wind direction--S E.

## May

Arabian Sea storms begin to affect Sind  
 (Tropical Sea storms of the Arabian Sea enter Sind Coast, once in 12 months )  
 Practically dry weather Cloudiness increases  
 Moderate to strong surface winds from S W  
 Dust storms expected  
 Mean temperature-- Max 102.8° F , Min 78.3° F.  
 Mean rainfall--15 inches.  
 Mean Baro pressure-- 29.590.  
 Mean humidity--60%  
 General wind direction--S E & S W

## June

Hottest month of the year  
Monsoon condition but with practically no rain  
Wind current stronger than in May  
Rough seas and swells  
Coasting steamers stop sailing  
Mean temperature—Max 103.3° F., Min 83.1° F  
Mean rainfall—33 inches  
Mean Baro pressure—29.429  
Mean humidity—63%  
General wind direction—S W & S E

## July

Rainiest month of the year  
Monsoon current reaches the interior  
Humidity is expected to increase  
Western depression on rare occasions, wind and rain  
Wind current strong  
Mean temperature—Max 99.2° F., Min 82.8° F  
Mean rainfall—2.58 inches  
Mean Baro pressure—29.391  
Mean humidity—72%  
General wind direction—S W, S E

## August

Milder weather prevails  
More cloudy and less rainy month  
Visibility is improved. Wind direction still, S W  
Eastern depressions sometimes occur  
Mean temperature—Max 95.3° F., Min 80.2° F  
Mean rainfall—1.86 inches  
Mean Baro pressure—29.473  
Mean humidity—75%  
General wind direction—S W, S E

## September

Receding monsoon  
Eastern disturbances expected sometimes  
Dust storms and squalls are fewer  
Wind velocity gets reduced  
Mean temperature—Max 95.4° F., Min 74.4° F  
Mean rainfall—82 inches  
Mean Baro pressure—29.624  
Mean humidity—73%  
General wind direction—S W, S E

## October

Dry and clear month  
Only rare Arabian Sea depressions  
Practically no rain  
Temperature rises slightly at times  
Wind direction now alters  
Sea is smooth and coasting steamers begin to ply  
Mean temperature—Max 94.9° F., Min 70.0° F  
Mean rainfall—.06 inches  
Mean Baro pressure—29.799  
Mean humidity—63%  
General wind direction—S W., N E

## November

Wind direction changes from W-SW to N-NW, N E  
 Pleasant month. Without any showers of rain  
 Western disturbances on occasions bring drizzles rarely  
 Mean temperature—Max 87.4° F, Min 59.7° F.  
 Mean rainfall—03 inches  
 Mean Baro. Pressure—29.937  
 Mean humidity—57%  
 General wind direction—N E, N W

## December

Squally weather expected  
 Western disturbances come in at times  
 Pleasant month  
 Wind directions changes to N and N E  
 Winter showers expected  
 Sea very smooth  
 Mean temperature—Max 77.6° F, Min 51.8° F.  
 Mean rainfall—10 inches  
 Mean Baro. pressure—30.010  
 Mean humidity—60%  
 General wind directions -N W, N E

*N.B.*—The above summary of the weather conditions for the whole of Sind is based upon the climatic data for the three main stations in Sind, utilised in the body of the paper viz. Karachi, Hyderabad and Jacobabad.

**APPENDIX II**  
**Temperature, Barometric Pressure and Humidity Normals**

*Climatic Conditions in Sind*

Karachi (Mauria)				Hyderabad				Jacobabad			
Temperature	Pressure	Humidity	Temperature	Pressure	Humidity	Temperature	Pressure	Humidity	Pressure	Humidity	Pressure
(Max.)	(Min.)	%	(Max.)	(Min.)	%	(Max.)	(Min.)	%	(Max.)	(Min.)	%
January	75.7	58.6	30.065	43	78.2	60.6	30.100	57	73.0	43.8	29.915
	76.6	56.7	30.050	61	75.6	60.6	30.066	61	72.7	44.2	29.922
February	78.2	61.3	30.047	67	79.5	62.9	30.053	65	72.8	44.2	29.925
	79.3	63.5	30.049	67	80.4	64.0	30.052	65	72.9	45.7	29.927
March	81.8	64.3	29.954	67	82.6	64.5	29.944	53	70.7	45.7	29.913
	82.7	67.3	29.948	71	82.5	65.3	29.944	53	70.3	45.7	29.913
April	85.3	74.0	29.836	79	82.0	72.2	29.818	47	99.6	69.3	41.4
	87.0	74.2	29.832	76	80.2	71.6	29.816	47	101.8	70.6	41.9
May	88.8	78.3	29.757	80	81.7	78.2	29.601	54	111.0	78.3	42.6
	90.2	78.9	29.723	75	80.7	76.1	29.569	55	111.3	78.3	43.0
June	91.0	82.4	29.677	80	81.4	81.7	29.472	62	84.3	59.6	39.460
	92.6	82.4	29.546	79	81.4	81.7	29.439	64	112.9	84.3	39.318
July	88.2	80.4	29.540	83	79.2	64.7	29.419	65	112.5	83.7	39.295
	89.6	81.2	29.523	62	80.6	64.7	29.391	69	108.5	81.6	39.290
August	85.5	77.7	29.651	65	80.9	73.0	29.468	70	104.6	81.9	39.361
	86.5	77.7	29.650	64	80.9	74.6	29.467	70	101.9	81.5	39.347
September	86.8	75.8	29.745	62	81.7	74.4	29.658	66	103.7	78.2	39.315
	88.6	75.8	29.747	62	81.7	75.6	29.639	70	102.9	78.2	39.306
October	88.6	77.0	29.918	73	97.8	70.5	29.809	56	99.2	81.2	39.719
	89.5	73.5	29.904	74	97.9	70.5	29.796	61	99.8	81.8	39.639
November	84.3	66.7	30.049	68	89.1	58.9	29.942	62	89.8	51.8	39.818
	85.7	65.3	30.074	67	88.9	60.2	29.943	62	89.8	51.2	39.817
December	78.2	58.6	30.100	58	79.0	62.3	30.025	65	78.7	44.3	39.934
	79.2	58.6	30.090	60	78.3	57.2	30.017	59	73.5	44.7	39.920
Annual	64.1	71.2	29.633	73	90.3	65.2	29.741	59	91.6	66.3	39.636
	65.5	71.2	29.639	73	90.3	65.2	29.736	62	91.6	66.4	39.627

The figures stated are monthly and annual normals of 30 years (1901-1930) obtained by me from the data gathered from India Weather Review (Mad. Met. Dept.), while the others are monthly and annual normals (Published in *Memoirs Ind. Met. Engg.*, Vol 22, Pt. III, Calcutta, 1914). The figures given at the top of each column show the number of years of which the latter are normals. At Karachi, the conditions are nearly the same except a slight rise in the maximum  $T_{10}$ . Humidity is doubtless increased at Hyderabad while at Jacobabad the range of  $T_{10}$  is reduced, so also the barometric pressure and humidity

## Rainfall Normals

Station	No. of years taken	Jan	Feb	Mar	April	May	June	July	Aug	Sept	Oct	Nov	Dec	Annual
Nagar Panar	14	0.03	0.22	0.10	0.06	0.06	0.42	5.70	5.27	1.20	0.64	0.10	0.03	13.22
30	0.03	0.23	0.10	0.07	0.01	0.37	6.64	6.18	4.52	9.49	0.39	0.04	0.04	44.65
32	0.28	0.42	0.13	0.18	0.07	0.68	4.00	1.83	0.54	0.60	0.14	0.07	0.24	8.24
Tata	20	0.11	0.27	0.22	0.04	0.21	0.61	4.48	2.09	0.93	0.62	0.04	0.05	8.85
Karachi (M. Adara)	31	0.63	0.48	0.23	0.17	0.08	1.03	2.95	1.19	0.51	0.00	0.17	0.15	7.79
30	0.21	0.46	0.34	0.07	0.10	0.57	2.76	1.65	0.76	0.63	0.20	0.15	7.51	7.51
Umerkot	33	0.17	0.13	0.15	0.09	0.06	0.68	3.00	2.22	1.90	0.00	0.05	0.04	7.33
30	0.09	0.14	0.17	0.02	0.17	0.46	3.19	2.77	1.34	1.33	0.07	0.03	0.02	8.95
Hyderabad	41	0.24	0.28	0.15	0.13	0.11	2.11	2.68	0.53	0.00	0.09	0.08	0.08	7.32
30	0.22	0.30	0.29	0.06	0.16	0.36	3.49	2.12	0.93	0.00	0.03	0.10	7.92	7.92
Nasikero	30	0.25	0.31	0.21	0.03	0.07	0.48	1.13	1.15	0.37	0.00	0.02	0.10	4.12
30	0.15	0.22	0.18	0.03	0.03	0.30	1.40	1.11	0.67	0.00	0.02	0.13	4.76	4.76
Selwas	31	0.16	0.32	0.12	0.19	0.15	0.53	1.66	1.31	0.91	0.03	0.08	0.06	5.18
30	0.23	0.27	0.14	0.14	0.08	0.48	2.14	1.29	0.72	0.00	0.03	0.06	5.42	5.42
Sukar (Kohn)	32	0.25	0.26	0.19	0.18	0.09	0.68	0.74	1.01	0.48	0.00	0.08	0.10	3.12
17	0.04	0.15	0.14	0.03	0.17	0.06	0.66	0.30	0.28	0.03	0.65	0.21	2.74	
Shikarpur	32	0.29	0.36	0.29	0.16	0.12	0.13	0.69	0.47	0.20	0.00	0.18	0.18	3.58
30	0.14	0.29	0.25	0.06	0.07	0.21	0.72	1.02	0.46	0.04	0.01	0.17	3.59	3.59
Jacobabad	42	0.33	0.32	0.28	0.16	0.13	0.15	1.13	1.17	0.19	0.01	0.08	0.11	4.06
30	0.23	0.30	0.26	0.20	0.13	0.36	0.62	0.70	0.28	0.05	0.02	0.05	3.44	

The figures tabulated are normals of more recent years, obtained by me from the data gathered from *India Weather Review*

*Ind. Met. Dept.*, while the others are those published in *Mem. Ind. Met. Dept.*, Vol. 22, Pt. 1 Calcutta, 1913.

It must be noted that there is a tendency of increasing rainfall at some stations in Lower and Middle Sind and that this increase is chiefly in the summer rains (especially in July).

## APPENDIX III

Comparison of Annual Rainfall at Karachi (Manora) and in Kohistan

(See Plate V.)

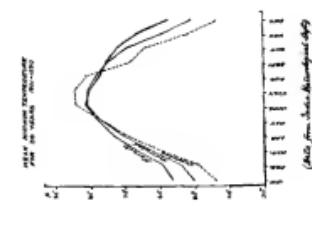
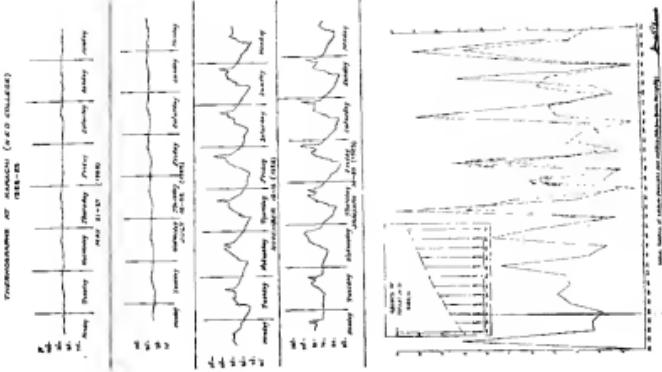
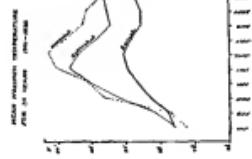
Year	Karachi and Manora	Kohistan
1896	11.60	1.38
1897	12.24	10.47
1898	4.03	4.08
1899	0.83	0.15
1900	2.14	8.05
1901	2.05	3.19
1902	20.76	16.15
1903	3.86	6.35
1904	4.59	3.21
1905	3.09	4.90
1906	8.93	9.70
1907	8.15	12.25
1908	7.03	23.09
1909	7.03	3.79
1910	12.63	12.75
1911	4.72	9.0
1912	0.39	3.35
1913	13.60	31.55
1914	9.43	6.26
1915	2.22	0.85
1916	22.32	1.338
1917	5.52	16.21
1918	1.87	2.01
1919	2.59	9.63
1920	1.44	3.16
1921	16.35	18.44
1922	1.98	3.65
Average	6.92	8.96

(Data obtained from the Gazetteer of the Province of Sind B Vol 1.)

## REFERENCES

- 1 Balfour, H. F., *A Practical Guide to the Climates and Weather of India, Ceylon and Burma* London, 1889, 96
- 2A *Ibid* 266-270
- 2 Balfour, H. F., "A full discussion of the Rainfall of India and cognate Subjects," *Ind Met Mem.* 3 122  
Also *Chronicles and Weather of India and Ceylon*, 133-36
- 3 Brooks, *The Weather*, 1927, 70
- 4 Hemraj and Walker, *The Oxford Survey of the British Empire* Ch. II, 69
- 5 Banerjee, B. N. *Meteorology of the Persian Gulf and Makran* Calcutta, 1931, 1-2
- 6 *Ibid*, 3
- 7 *Ibid*, 4
- 8 *Ibid*, 11
- 9 *Ibid* 11
- 10 Jenkins, W. J. *Annual Report of the Departments of Agriculture*, Sind, 1911-14, 7 8
- 11 Chambers Fred, "The Winds of Karachi," *Mem Ind Met Dept* 1, 4
- 12 Raendas, L. A., *Scientific notes* 1932 4, No. 41 *Ind Met Dept* 115-117
- 13 Burton, R. I., *Sind*, London, 1851, 377
- 14 Balfour, H. F., *Ind Met Mem* 52, Pt 1 258
- 15 Simpson, G. C. *Quart Jour Roy Met Soc* July 1921 47, 1921, 158 ff
- 16 Williamson, A. V., and Clerk, K. G. T., "The Variability of the Annual Rainfall of India," *Quart Jour Roy Met Soc* 57, Balfour, H. F., *Ind Met Mem* 3 Pt 1
- 17 Clerk, K. G. T., "Varietudes of the Summer Rainfall of the Indo-Gangetic Plain," *Geography* No 98, December 1932 288-291, *Quart Jour Roy Met Soc* 57, 42-43
- 18 Abercromby R., *International Science Series* London 1907, 319
- 19 Burton, R. F., *Sind Recusited* London, 1877, 104
- 20 Stein, Sir Aurel, "Archaeological Tour in Gedrosia," *Mem Arch Soc Ind*, No 43, 11
- 21 Huntington Ellsworth, *The Pulse of Asia*, 1905, 236 ff
- 22 Marshall, Sir John, *Mohen-jo-daro and the Indus Civilisation*, London, 1931, 2-5
- 23 *Ibid* 3
- 24 Majumdar, N. G., "Exploration in Sind," *Mem Arch Soc Ind*, 1927, No 48
- 25 Woodburn, Gen., "Former Climate of Sind," *Bombay Times*, 1856, 61, Pt 1, *Trans Bom Geog Soc*, M/152, 89, Appendix, p 7
- 26 Raverty, "Milieu of Sind and Its Tributaries," *Jour Asia Soc Beng*, 1892, 61 Pt 281, 312, 417
- 27 Mackay, E., *The Indus Civilization* London, 1935, 10
- 28 Walker, Sir Gilbert T., "On the Meteorological Evidence for the supposed change of climates in India," *Mem Ind Met Dept*, 21, 1





MOUNT R. PELONCHÍN,  
CHILEAN CHILE



ANNUAL PRECIPITATION (mm)



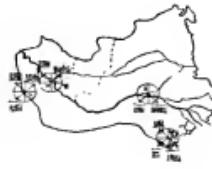
PRECIP. AND ALTITUDE SUR. 0, AND PT. P. I. P. I. F.

MONTHLY RAINFALL IN SNOW



(Data from Jose Hernandez 1970)

SURFACE WINDS OVER SNOW



ANNUAL PRECIPITATION  
(Chilean National Water & Power)



ANNUAL WIND & PRESSURE

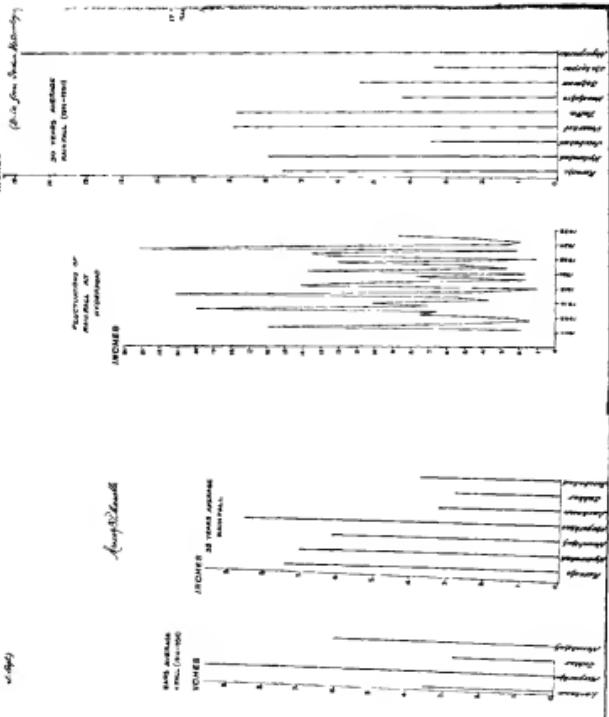


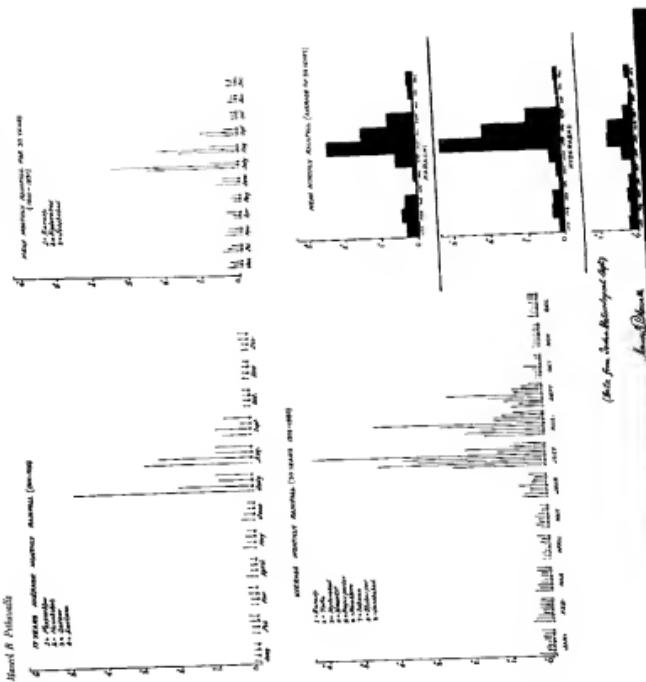
CONTINENTAL TRADE WINDS & OCEANIC  
GROWING LINES DURING MIGRATION



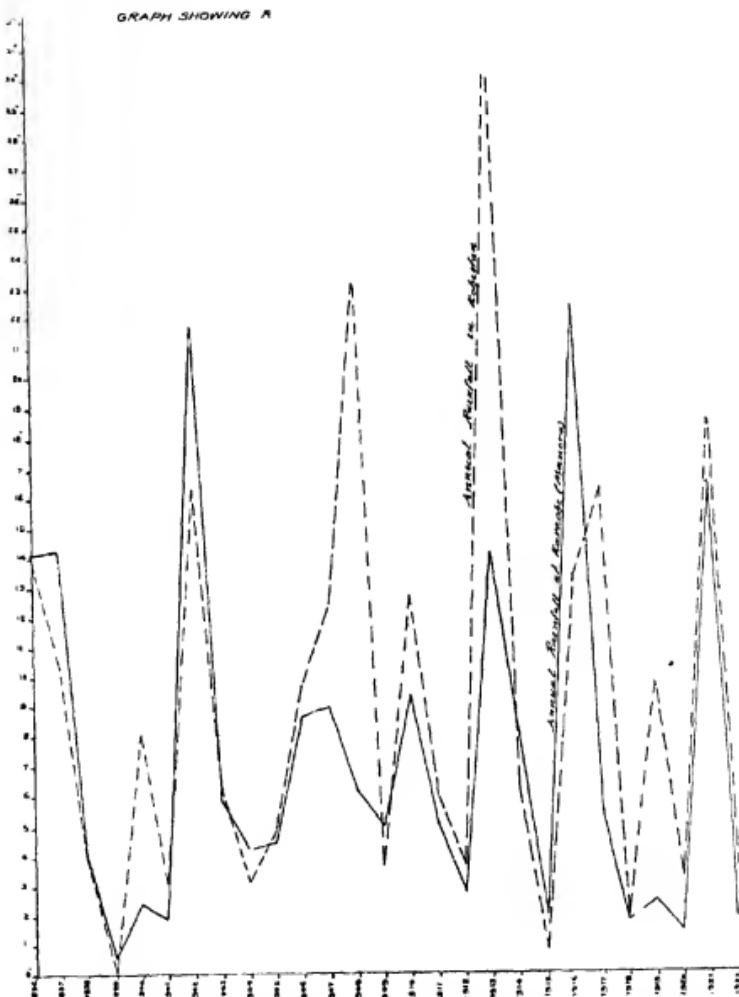
Map of Chile













## EXPLANATION OF PLATES, SKETCH-MAPS, GRAPHS, ETC

- I (a) Sketch-maps of Sind showing—
  - (1) Climatic divisions
  - (2) July temperature (Mean)
  - (3) Rainfall distribution
  - (4) Surface winds
- (b) Sketch-maps of India showing—
  - (1) July rainfall (Blanford)
  - (2) July temperature (Blanford)
  - (3) Variability of summer rainfall (Clerk)
  - (4) July wind and pressure (Blanford)
  - (5) Stream lines during monsoon (Blanford)
- II Graphs showing—
  - (1) Mean maximum temperature at Karachi, Hyderabad and Jacobabad
  - (2) Mean minimum temperature at Karachi, Hyderabad and Jacobabad
  - (3) Thermograms at Karachi
  - (4) Annual rainfall at Karachi (Manora) and Khadki and Growth of Population of Karachi
- III Graphs showing—
  - (1) Fluctuations of rainfall at Hyderabad
  - (2) Average annual rainfall at Karachi and other stations
- IV Graphs showing—
  - (1) Average monthly rainfall at Karachi and other stations
  - (2) Mean monthly rainfall at Karachi, Hyderabad and Jacobabad
- V Graph showing annual rainfall at Karachi (Manora) and Kohistan

## ECHINOIDS FROM THE BAGH BEDS.

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Received May 28, 1937

(Communicated by Prof. L. Rama Rao, M.A., F.G.S.)

### Introduction

The Cretaceous age of the Bagh Beds was first recognised by Col. Keatinge<sup>12</sup> in 1850 from the fossils he collected from the neighbourhood of Chirakhan (lat.  $22^{\circ} 22' 30''$ , long.  $75^{\circ} 7' 30''$ ). Of the fossils from these beds, only the echinoids and the ammonites have so far received proper attention. Duncan<sup>13,14</sup> on the basis of his study of the echinoids from these beds considered them to be of Cenomanian age. This conclusion was accepted by W. T. Blanford,<sup>7</sup> R. D. Oldham<sup>17</sup> and D. N. Wadia<sup>17</sup>. By others, however, these beds have been variously assigned an age from Albian to Senonian.<sup>8,10,18,44,45,51,52</sup>

While working on the echinoids from the Bagh Beds collected by the late Prof. K. K. Mathur and by the present writer, a few new forms have been discovered. These, along with such of the forms already known as required some additional remarks on the basis of the new collections, are described below.

Of the echinoids described by Fourtau<sup>19</sup> the name *Hemaster oldhami* being pre-occupied for a Maestrichtian species from Baluchistan described by Noethling,<sup>44</sup> the species is here renamed *H. fourtau*. The other two spatangid species were assigned by Fourtau to the genus *Opissaster*, Pomel, because of their ambulacral petals sinuous and unequal at the poriferous zones.<sup>20</sup> This genus is, however, regarded by Duncan<sup>15</sup> and R. T. Jackson<sup>21</sup> as an invalid one. The genus was evidently established by Pomel<sup>17,22,23</sup> to include *Hemaster* species having *Schizaster*-like physiognomy but devoid of the latero-sub-anal fasciole. Pomel himself, later on included in this genus, species with two, three or four pores, high inter-ambulacral plates and petals flexuous or non-flexuous and excavated or not. The confusion thus created led Lambert<sup>24,25</sup> to revise the genus as "having two genital pores, flexuous ambulacral petals and ranging from Eocene to Recent."

The Indian specimens under consideration certainly do not belong to *Opissaster*. They have flexuous ambulacrals and an apical disc with four perforate genital plates, the madreporite on the right anterior genital plate

extends centrally but does not separate the posterior genital plates. They thus belong to the genus *Hemistuster*.

*Description of the Species*

*Salenia mathuri,\* sp. nov.*

Pl VI, Figs 3a-3d

Dimensions —	Diameter	Height	Height/Diameter
	11.0 mm	6.8 mm	6 18/10

*Description* — Test is much depressed, ambitually tumid, very feebly convex superiorly and rounded inferiorly. Small peristome is about  $\frac{1}{3}$  of the diameter of the test, branchial incisions are well marked. Apical disc is large and polygonal being about 66.1 per cent of the diameter of the test. Periphery is raised with sutures of genital and ocular plates strongly incised. Genital plates are as broad as high, well angulated adorally, ornamented with radial grooves and ridges and unequally divided, with the outer part markedly smaller. Large elliptical genital pores are imperceptibly rimmed, adorally eccentric and placed in feeble depressions. Madreporite is on the right anterior genital plate and roughly trapezoidal in shape running towards the suture with ocular III. Sur-anal plate is larger than the genital ones but similarly ornamented. Ocular plates are radially grooved, twice as broad as high and feebly adorally convex, with adorally eccentric ocular pores. At the periphery, ocular plates are broader than the genital ones. The ocular plate I is absent. Periproct is broadly elliptical and deeply excavates the sur-anal plate, reaching the pores on the genital plates 1 and 2.

Ambulacral areas are wide and slightly flexuous. Pore-pairs are oblique with pores separated by granules. Adapically pore-pairs are uniserial over three-fourths of the ambulacral length, from halfway between ambitus and peristome they become gradually biserial attaining distinct triserial disposition on the first three adoral plates. This transition is accompanied by an appreciable widening of the ambulacral area, swelling of the inter-poriferous tubercles and widening of the miliary zone. On approaching the peristome the inter-poriferous zone is narrowed slightly with corresponding decrease in the size of the last three tubercles, while the triserial arrangement maintains the width of the ambulacral areas. The inter-poriferous zone carries two rows of small tubercles and microscopic warts disposed zig-zag in the middle area.

Inter-ambulacral areas are three times as wide as the ambulacral ones and carry two series of high plates, four or five per series. The primary

\* This species is named after the late Prof K. K. Mathur.

tubercles are crenulate, imperforate and mammeloned, the scrobicular rings are of the "Quadrata" type of Arnaud\*. The median zone with military granules is slightly narrower than the inter-poriferous zone. Each plate in the mid-zone abuts against 8 to 10 ambulacral plates at the ad-ambulacral suture.

*Comparison*—This species differs from the associated *S. keatsi*\*, Fourtau<sup>23,24,25</sup> chiefly by its higher but discoid form, apical disc and peristome smaller, lower and well-angulated genital plates, and conspicuously sudden increase in the width of ambulacral areas and military zone, and by the size of the inter-poriferous tubercles.

*S. presensis*, Desor<sup>18,19,20,21,22,23,24,25</sup> from Aptian of Algeria and Western Europe and *S. scutuligera*, Gray<sup>1,2,3,4,13,15,16,18,19,20,22</sup> from Cenomanian of Charente, Tuus and Algeria differ from the present species by their relative dimensions and nature of the corona and apical disc.

*S. cylindrica*, Arnaud\* from France differs from the species described here by its taller test, wider peristome and a smaller apical disc, otherwise these two forms are very closely similar.

*S. mammillata*, Cottean<sup>14,15</sup> from the Aptian of Aube has its apical disc sculptured very similar to that of *S. mathuri*, but differs in relative dimensions and nature of the inter-ambulacral plates.

*S. maxima*, Arnaud<sup>16,17</sup> from Senonian of Charente has its apical disc and ambulacral areas very similar to those of the present species, but they can be distinguished one from the other by their relative proportions, lower inter-ambulacral plates and wider inter-ambulacral areas.

*N.B.*—All the type specimens are preserved in the Department of Geology, Benares Hindu University.

*Diplopodis (Tetragramma) aff. micropyga*, Fourtau

Pl VI, Figs 1a-1b

Dimensions —	Diameter	Height	Height/Diameter
	27.2 mm	10.0 mm	3 7/10

*Description*—Test is discoid with small peristome in a conspicuous depression. Apical disc is not preserved but will be about one-thirds of the diameter of the test.

Ambulacral areas are narrow with poriferous zones slightly flexuous, pairs of rounded pores are oblique in arcs of four, being uniserial at the ambitus and diplopodous at half the distance from the peristome and the apical disc towards the ambitus. Plates consist of three primaries and one adoral demi-plate, the primary next to the demi-plate being the largest.

Two series of 16 or 17 primary perforate crenulate, scrobiculated tubercles cover the inter-poriferous zone, milky granules being absent

Wide inter-ambulacral areas carry six series of primary perforate crenulate tubercles with scrobicular rings. Disorderly granules in the median sutural area die out dorsally above the ambitus. The tubercles are almost equal to those of the inter-poriferous zones. Only the middle two series with 15 or 16 tubercles reach the apical disc, the lateral series dying out successively above the mid-zone. Internal ranges of tubercles show that this specimen belongs to Lambert's group of *Tetragramma zumoffeni*, Cotteau <sup>22,24,27</sup>.

*Comparison*—*E. almerai* (Lambert)<sup>24,27</sup> from Aptian of Barcelona differs from the present species by its more depressed, polygonal test and weaker doubling of the pore-pairs.

*Pseudodiadema rossyi* (Cotteau)<sup>19</sup> from Cenomanian of Sarthe has its test higher, primary tubercles more differentiated and ambulacral areas wider.

A very close affinity is found between the present specimen and *D. micropyga*, Bourtau<sup>20</sup> from the lower Cenomanian of Egypt, the bad preservation of the specimen described here however leaves their identity in doubt.

#### *Echinobrissus angulifer*, Gauthier

*Description*—The specimens available for study are not well preserved. The test is longer than broad, anteriorly tumid and feebly tapering, and more rounded than truncated behind. Periproctal groove reaches upto half the distance between the apex and the slightly indented posterior border. Summit of the test coincides with the apical disc at three-fifths length from the front. Peristome is in a depression at two-fifths length from the front. Ambulacrals are lanceolate with conjugate, elongate pores and the inter-poriferous zones are narrower than the poriferous ones.

*Comparison*—These specimens offer the closest resemblance to the variety, *E. angulifer*, Gauthier<sup>18</sup> from upper Cenomanian of Algeria.

#### *Echinobrissus* sp. indet

These specimens are also poorly preserved. They differ from those described above by their having a test almost squarish in outline with a very obtusely rounded anterior border. Distally the ambulacrals are slightly deflected outward.

These specimens resemble more closely to the variety of *E. angulifer*, Gauthier<sup>18</sup> from the Albian and middle Cenomanian of Algeria.

It is significant that these two echinobrissi should occur in the Bagh Beds at the same horizon, their Algerian allies being found in successive horizons, i.e., in Albian to middle Cenomanian and upper Cenomanian. Though with better material to work upon, these two forms may have to be merged into one species, in their present state of preservation they are best described separately.

*Hemiasaster fourtaui*, † nom. nov.

1887 *Hemiasaster cenomanensis*, Duncan non Cotteau—Rec. G. S. I., 20, p. 91.

1918 *Hemiasaster oldhami*, Fourtau non Noetling—Rec. G. S. I., 49, p. 46, pl. 2, figs. 2-3.

This species is recorded here under a new name, its old name being pre-occupied for a Me-trichtian species from Baluchistan. It differs from the Baluchistan species mainly by its test being widest behind the middle, narrower and strongly excavated anteriorly, superior face sloping more gradually to the front than behind, apical disc more eccentric behind, paired petals less divergent and peripetalous fasciole more regular.

*Hemiasaster holo-ambitus*, sp. nov.

Pl. VI, Figs. 4a-4c.

1887 *Hemiasaster similis*, Duncan non d'Orbigny nec Cotteau nec Oldham—Rec. G. S. I., 20, p. 92.

1918 *Opissaster* sp. indet.—Rec. G. S. I., 49, p. 51, pl. 2, fig. 4.

Test is oval, almost vertically truncated behind and rounded in front. Apical disc is at one-thirds length from behind, rather compact with four perforate genital plates, the madreporite on the right anterior genital plate extending centrally but not separating the posterior genital plates.

Odd ambulacrae is in shallow, narrow sulcus which stops well above the ambitus. Rounded pores are separated by granules, the flexuous paired petals are in shallow grooves, with pores arranged in "Chevrons". Posterior petals are about three-fifths of the anterior ones. The biggest of the specimens has 16 pore-pairs in the posterior petals and 28 in the anterior ones.

Periproct is towards the top of the posterior face. Peristome is at one-thirds length from the front. Peripetalous fasciole crosses the odd ambulacrae well above the ambitus.

† This species is named after R. Fourtau who had previously described this species under a different name.

This species combines the characters of *Hemaster* *vexus* stricto and of *Integaster*, Lambert and Thierry <sup>40</sup>

**Comparison**—Due to bad preservation of the specimens available for study, Fourtau had wrongly allied this species with *H. vignesi*, Cotteau from the east of Jordon <sup>41</sup>. But it has much lower test, more developed ambulacrals and apical disc more eccentric behind

The present species differs from *H. stella* (Morton) <sup>4, 9-12, 31, 33, 54</sup> from upper Cretaceous of U.S.A. by its test medianly widest and anteriorly more tufted, upper face sloping forward more gradually, anterior groove narrower and shorter, and petals more flexuous

Narrower form, shallower grooves and flatter inferior face distinguish this species from *H. asterias*, Forbes <sup>14, 31, 34</sup> from the Albian of Folkstone and Yonne

From *H. rutoi*, Lambert <sup>42</sup> from Senonian of Belgium, this species differs by its petals more flexuous, test ambitinally oval and inferiorly more convex

*Hemaster mesles*, Peron and Gauthier

1878 *Hemaster mesles*, Peron and Gauthier—*Ech foss Algeria*, fasc 4, p 10, pl 2, figs 5-8

1889 *Hemaster mesles*, Peron and Gauthier—*Ech foss Tunisie*, p 12

1914 *Hemaster mesles*, Peron and Gauthier—*Cat Invert foss Egypt Pal Ser*, No 2, p 81

1932 *Hemaster mesles*, Peron and Gauthier—*Mem Soc Geo France*, No 16, n.s. VII, fasc 4, p. 12

The specimens from Bagh Beds differ from the type of Peron and Gauthier from the Cenomanian of Algeria by their very slightly broader test and a little less conspicuous anterior indentation. Such differences, however, are not unexpected in species having such a wide geographical range

*Hemaster cf. thomass*, Peron and Gauthier

Pl VI, Fig 2

Dimensions—Length	30.6 mm	22.5 mm	20.0 mm	17.0 mm
Breadth	30.6 ..	21.4 ..	21.0 ..	16.4 ..
Height .	19.8 ..	14.0 ..	15.0 ..	11.0 ..

**Description**—Test is trigonal, widest at one-fourths length from the deeply indented front, rapidly narrowing behind, with superior face more or less regularly raised

Pores in the odd ambulacrae are oval, placed in "Chevrons" and separated by granules. An internal row of serobiculated tubercles rises up along the poriferous zones for half the length of the groove, above which they die out gradually. Two rows of granules follow the horizontal sutures of the ambulacral plates. In the paired petals, the pore-pairs are oblique at the proximal ends, straight in the middle and in "Chevrons" at the distal ends. Dorsally to the pore-pair a line of granules passes along the horizontal sutures. Posterior petals are three-fourths of the anterior ones. The biggest specimen has 36 pore-pairs in the posterior petals and 41 in the anterior ones. Inter-poriferous zones are narrower than the poriferous ones.

Periproct is towards the top of the posterior steeply truncated face. Peristome is at one-fourths length from the front. Peripetalous fasciole crosses the odd ambulacrae just above the ambitus and shows no inflexions.

Apical disc is sub-central, rather broad with four perforate genital plates. Madreporite is on the right anterior genital plate and extends posteriorly so as to separate the posterior genital plates. Ocular III penetrates between the genital plates 2 and 3.

*Comparison*—This species has a very distinctive form and need be compared only with *H. thomasi*, Peron and Gauthier<sup>18,19</sup> from the Senonian of Algeria. The Algerian species, however, attains twice the size of the Indian form, its infensor face is more inflated, superior face more raised, test a little broader and the posterior petals a little longer.

#### *Discussion and Conclusions*

The accompanying table summarises the vertical distribution and the affinity-relations of the echinoids from the Bagh Beds with those from Europe, North Africa and Western Asia. It will be seen that some of the species show close affinities with co-Cretaceous forms and appear to be ancestral to some meso-Cretaceous species *Placenticeras minor*, Vredenburg is closely related to *P. uhligi*, Choffat and *P. saadense*, Peron and Thomas. The appearance of the genus *Hemiasper* in the Albian coupled with these relations of the echinoids and ammonites led Fourtau to regard upper Albian (upper Gault of the British Stratigraphers) rather than lower Cenomanian as the more probable age for the Bagh Beds<sup>20,21</sup>. Such a view was not unjustified, considering that the material with which Fourtau had to deal consisted only of the few type specimens in the collection of the Geological Survey of India.

With the extensive collection at the disposal of the present writer it has been possible to carry out a detailed study of these echinoids. Besides the eight species previously known to be represented in these beds, six new

and important forms are now added to the list, thus giving a basis much more definite, than was possible hitherto, for the correlation of these beds with those outside India.

Among such valuable forms is *Diplopodopsis cf. microphyga*, Fourtau showing lower Cenomanian affinities<sup>20</sup>. The genus *Diplopodopsis*, McCoy, not known to survive the Cenomanian (lower Chalk)<sup>21,22</sup> fixes Cenomanian as the upper age limit for the echinoid fauna of the Bagh Beds. Four out of five species of *Hemaster* recorded from these beds of the Narbada valley belong to the sub-genera *Proraster*, *Mecaster* and *Integraster* all of which make their first appearance in the Cenomanian<sup>23</sup>, obviously, this gives the lower age limit for the beds under consideration.

*Hemaster fourtau*, nom. nov., by far the commonest of the echinoid species from these beds, has its nearest ally *H. lyneesi*, Cotteau in the Cenomanian of Palestine. *H. meslei*, Peron and Gauthier, though not very abundantly represented in the Bagh Beds, occurs in the Cenomanian of Algeria, Tunis and Egypt<sup>24,25,26</sup>. *H. sub-similis* (Fourtau)<sup>21,22</sup> and *H. cf. thomasi*, Peron and Gauthier<sup>24,25</sup> have their near relations in the Senonian of Persia and Algeria respectively.

Vredenburg<sup>27</sup> regarded his *Placenticeras minor* as being very closely related to *P. tamolicum*, Stoliczka from the Trichinopoly stage (lower Senonian) of the South Indian Cretaceous. It is also closely related to *P. whiagi*, Choffat<sup>24,28</sup> and *P. saadense*, Peron and Thomas<sup>24,25</sup> of which the latter is a Cenomanian form, the former ranging from Gault to lower Cenomanian.

Though the genus *Hemaster* makes its appearance in the Albian, its species from the Bagh Beds, as is shown above, cannot certainly be dated older than the Cenomanian. Thus taking into consideration the affinities of these echinoid species in general, the more appropriate age for the Bagh Beds, so far as the present study shows, will be lower Cenomanian rather than upper Albian.

P. N. Bose<sup>8</sup> was inclined to regard the different subdivisions of the Bagh Beds as representing distinct stages approximately equivalent to those of the Cretaceous Series of South India. The vertical distribution of the echinoid species as given in the accompanying table is, however, quite contrary to this idea. The different constituents of this series of the Narbada valley must be considered, as Duncan has mentioned,<sup>24</sup> to belong to a single geological age, the differences being due more to variations in the lithological facies than to geological time.

Table showing the Vertical Distribution and Affinity-Relations of the Echinoids from Bight Beds

No	Species from Bight Beds	Related species with stratigraphical position	Nodular Limestone	Lower Coralline Limestone	Decca	Chubak Marl	Upper <sup>2</sup> Grotto Limestone
1	<i>Doxocidaris seminudus</i> , Duane	<i>Zonulites</i> , Gaither	Apitan of Algeria				
2	<i>Selena fonscavai</i> , Fournas	<i>S. seminudus</i> , Coteau	Apitan of France				
3	<i>S. matthei</i> , sp. nov.	<i>S. cylindrica</i> , Arnaud	Apitan Alban-of France				
4	<i>Cyphocidaris seminudus</i> , Fournas	<i>C. seminudus</i> , Berremans of France and Switzerland					
5	<i>Orikopis isidus</i> , Duane	<i>O. repiliatus</i> , Virey, Perrmann Alban of France, Portugal and Switzerland					
6	<i>Diplopoda (Terebratula) off macro</i>	<i>D. macrospira</i> , Fournas, Lower Cenomanian of Egypt					
7	<i>Schuchardtia lydia</i> , Fournas	<i>E. eddianus</i> Gaither, Apitan Alban of Algeria and Tunis					
8	<i>E. angustus</i> , Gaither	<i>E. elongatus</i> , Gaither var. <i>opercularis</i> of Algeria					
9	<i>E. sp. valid</i>	<i>E. cylindrica</i> , Gaither, var. <i>Albanus-middle Cenomanian</i> of Algeria					
10	<i>Hemicidaris fonscavai</i> , nom. nov.	<i>H. luteus</i> , Coteau Cenomanian of Palestine					
11	<i>H. (Ptyctocidaris) subsemisphaerica</i> (Fournas)	<i>H. Morosa</i> , Coteau and Gaither, Senonian of Paris					
12	<i>H. holomorphoides</i> , sp. nov.	<i>H. excentrica</i> Forbes, Alban of Tunis and Fakhro and <i>H. recta</i> , Lambert, Senonian of Belgium					
13	<i>H. (Meocidaris) meocidaris</i> , Fournas & Gaither	<i>H. regularis</i> , Peron and Gauthier, Cenomanian of Algeria, Tunis and Egypt					
14	<i>H. (Meocidaris) tenuis</i> , Fournas and Gaither	<i>H. tenuis</i> , Peron and Gauthier, Senonian of Algeria					

## Acknowledgments

In conclusion, I have to express my indebtedness to the Council of the Benares Hindu University for the grant of a Research Scholarship during which period part of the present work was carried out. It is a pleasant memory to recall the kind encouragement that I received at the hands of the late Prof. K. K. Mathur. My grateful thanks are also due to Dr. Raj Nath, Head of the Department of Geology, for his kind interest in the present work and to the Director, Geological Survey of India, for permission to work in their Museum and Library.

## BIBLIOGRAPHY

1 Agassiz, L. *Monographie des Echinodermes Saliniers* 1838  
 2 ——— *Echinides fossiles de la Suisse*, 1839, 1  
 3 ——— *Ibid.*, 1840, 2

4 Agassiz and Desor *Catalogue Raisonné des Echinodermes*, 1847

5 Arnaud, H. *Mém. Soc. Geo. France*, 1877, 2, 10 fasc. 4

6 ——— *Act. Soc. Linn. Bordeaux*, 1897, 6, 2

7 Blanford, W. T. *Alm. Geo. Sur. India*, 1869, 4, pt. 3

8 Bose, P. N. *Ibid.*, 1884, 21, pt. 1

9 Clark, W. B. *John Hopkins University Circular*, 1891, 10

10 ——— *Ibid.*, 1893, 12

11 ——— *United States Geological Survey Bull.*, 1897, 97

12 Clark, W. B., and Twitchell, M. W. *Monograph of the United States Geological Survey* 1915, 54

13 Coquand *Bull. Soc. Geo. France*, 1859, 2, 16, 945-1023

14 Cotteau, G. *Echinides fossiles de l'Yonné*, 1857-78 2

15 ——— *Paléontologie Française, terr. crit.*, 1857-69, 7

16 ——— *Echinides fossiles de la Pyrénées*, 1865, 2

17 Cotteau, G., and Gauthier, V. *Mémoir Scientifique en Perse*, 1895, 3, pt. 2

18 Cotteau, Peron and Gauthier *Echinides fossiles de l'Algérie*, 1870-83, fasc. 3-8

19 Cotteau and Trigger *Echinides fossiles de la Département de la Sarthe* 1855-69

20 Cotter, G. de P. *Rec. Geo. Sur. India*, 1926, 59, pt. 4

21 Desor *Synopsis des Echinides fossiles et vivants*, 1854

22 Dumars *Statistique Géologique et Minéralogique du Département du Gard*, 1876, 2

23 Duncan Martin, P. *Quart. Journ. Geo. Soc. London*, 1865, 21

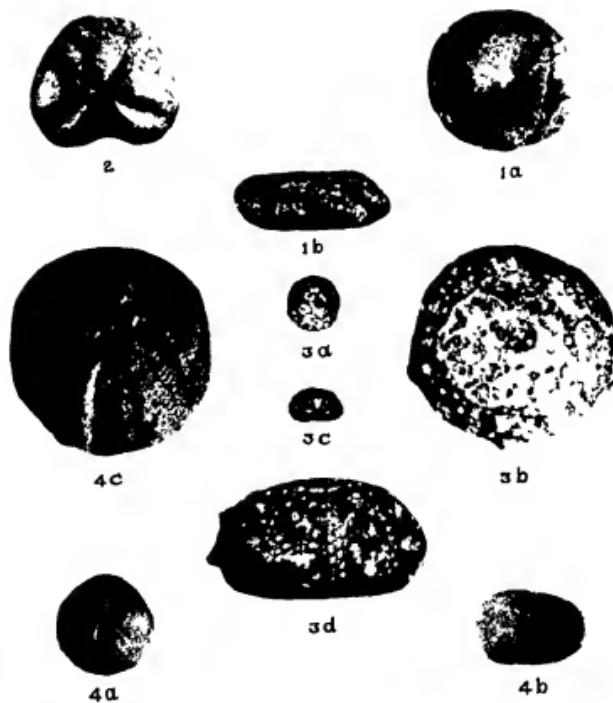
24 ——— *Rec. Geo. Sur. India*, 1887, 20, pt. 1

25 ——— *Journ. Linn. Soc. London (Zoo.)*, 1889, 23, Nos. 141-44

26 Forbes, E. *Morris's Catalogue of British Fossils*, 2nd Ed., 1854

27 Fourtau, R  
 28 —————  
 29 —————  
 30 —————  
 31 Jackson, R. T  
 32 Johnson  
 33 Keatinge, Col  
 34 Lambert, J  
 35 —————  
 36 —————  
 37 —————  
 38 —————  
 39 —————  
 40 Lambert, J., and  
     Thierry, P  
 41 Lartet  
 42 Leymerie and Cottreau  
 43 Loriot, P. de  
 44 Muckerjee, P. N  
 45 —————  
 46 Noeling, F  
 47 Oldham, R. D  
 48 Orhongy, A. de  
 49 Pomel  
 50 Ravn, J. P. J  
 51 Rode, K. P., and  
     Chiplonker, G. W  
 52 Spath L. F  
 53 Thomas and Gauthier  
 54 Thomas and Peron  
 55 Vredenburg, E  
 56 Wade, B  
 57 Wadia, D. N

*Mem. Inst. Egypt.,* 1899, 3, fasc. 7.  
*Catalogue des Invertébrés fossiles de l'Egypte, terr. crét*  
*Echinodermes, Geo Sur Egypt, Pal. Ser.,* 1914, No 2  
*Rec. Geo. Sur. India,* 1918, 49, pt 1  
*Catalogue des Invertébrés fossiles de l'Egypte, terr. crét*  
*Echinodermes (suppl.), Geo Sur Egypt, Pal. Ser.,*  
 1921, No. 5  
*'Echinodermata,' in Text-Book of Palaeontology, 2nd Ed.,*  
 1927, 1, 2nd Ed. reprinted, by Estmann, C. R.—Zittel  
*Proc. Acad. Nat. Sci. Philadelphia,* 1905, 57  
*Journ. As. Soc. Bengal,* 1858, 27, 116  
*Mem. Soc. Geo. France,* 1902, 9, fasc. 3, mem. 24  
*Ibid.*, 1906, 14, fasc. 2-3, mem. 24  
*Mem. Musée Roy. Nat. Hist. Belgique,* 1910, 4  
*Mem. Ciencias Nat. Barcelona, geo. ser.*, 1927, I, fasc. 1  
*Mem. Soc. Geo. France,* 1931, 16, (ns 7, pt 2)  
*Ibid.*, 1932, 16, (ns 7, pt 4)  
*Essais Raisonné du Nomenclature des Echinodermes,* 1909-25,  
 fasc. 1-9  
*Explor. Soc. de la Mer Morte,* 1877  
*Bull. Soc. Geo. France,* 1856, 2, 13, 319-54  
*Echinologie Helvétique—Méthodique pour la Paléontologie*  
*de la Suisse* 1873, 2  
*Rec. Geo. Sur. India,* 1933, 67, 71-73  
*Ibid.*, 1935, 69, 81  
*Pal. Ind.*, 1897, 14, 1, pt 3  
*Manual of Geology of India,* 1893, 2nd Ed.  
*Paleontologie française terr. crét,* 1853-59, 6  
*"Classification Méthodique et genres des Echinides fossiles*  
*et récents," published in *Paleontologie ou Description*  
*des Animaux fossiles de l'Algérie, Echinodermes,* 1885,  
 fasc. 2  
*Mem. Acad. Roy. Denmark,* 1928, 9, 1  
*Curr. Sci.*, 1935, 4, 322  
*Pal. Ind.*, 1930, ns., 15, pt 5, 64  
*Echinides fossiles de Tunisie,* 1889  
*Mollusques fossiles de Tunisie,* 1889  
*Rec. Geo. Sur. India,* 1907, 36, pt 2  
*United States Geological Survey, Professional Papers,*  
 1926, 137  
*Geology of India* (MacMillan & Co.), 1926, revised ed.*





EXPLANATION OF PLATE VI

FIG. 1—*Diplopodia (Tetragramma) aff. micropyga*, Fourtau.

(a) inferior view, natural size, (b) side view, natural size (B.H.U. No. E/4)

FIG. 2—*Hemaster cf. thomasi*, Peron and Gauthier, upper view, natural size, (B.H.U. No. E/12)

FIG. 3—*Solenia mathuri*, sp. nov., (a) superior view natural size, (b) same approximately 35 times natural size, (c) side view, natural size, (d) same approximately 35 times natural size (B.H.U. No. E/3)

FIG. 4—*Hemaster holoambitus*, sp. nov., (a) superior view, natural size, (b) side view, natural size, (c) apical view, approximately 35 times natural size (B.H.U. No. E/10)



**Papers Communicated for the Nineteenth Scientific Meet**  
**to be held on Saturday, the 4th September 1937, at 5-15 P.M., in the**  
**Central Building of the Indian Institute of Science, Bangalore**

**SECTION A**

<i>No</i>	<i>Title of the Paper</i>	<i>Author or Authors</i>
1	Einige diophantische probleme und Zahlentheoretische resultate	By Prof Dr Alfred Moessner (Com by Sir C V Raman)
2	On the emission and absorption band spectra of selenium	By Dr R K Asundi and Mr Y P Parti
3	Generalised action functions in Born's electro-dynamics	By Prof B N Madhava Rao
4	Generalisation of a theorem of Davenport on the addition of residue classes	By Dr S S Pillai (Com by Dr S Chowla)
5	The Condensation of aldehydes with in inionic acid in the presence of organic bases—Part IX. The condensation of $\beta$ -hydroxynaphthaldehyde (2-hydroxy- 1-naphthaldehyde)	By Prof K C Pandya and Mr Toquiq Ahmad Vahidy
6	The Replication of an experiment I Identical Samples from a binomial population	By Dr S R Savur

**SECTION B**

1	Cytogenetic aspects for transferring the virus localization ability of <i>Nicotiana</i> species at the background of <i>Nicotiana</i> <i>Tabacum</i>	By Prof Dr Dendho Kostoff (Com by Prof C R Narayan Rao)
2	Fungi of Allahabad, India, Part III ..	By Messrs J H Mitter and R N Tandon (Com by Dr Shri Ranjan)



ON PROTOCYATHEA RAJMAHALENSE SP. NOV.,  
A CYATHEACEOUS TREE-FERN, WITH NOTES ON  
THE GEOLOGICAL DISTRIBUTION OF THE  
CYATHEACEAE.

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Department of Botany, University of Lucknow

Received May 24, 1937

(Communicated by Prof B. Sahni, sc.b., f r s )

1 *Introduction.*

THE specimens described below were collected in December 1934 in the company of my friend, Mr. R N Mehrotra, M Sc , from certain plant-bearing beds at Sakrigalighat, situated about one mile north of Sakrigali railway station in Behar (see map in Sahni and Rao<sup>1</sup>) The collection from this locality was supplemented the following winter, when I had the privilege of accompanying Prof. Sahni to this particular locality. A third visit was made in December 1936. The specimens described in the present communication appeared to be of sufficient interest to deserve separate treatment. The remaining specimens from Sakrigalighat will be described in a subsequent paper.

The plant-bearing beds are exposed about half-a-mile west of the ferry on the northern slope of a small hill abutting on the Ganges. Surface collections were made previously by others. Mr W N Edwards was the first to locate the fossiliferous beds at Sakrigalighat<sup>2</sup>, but as far as I know none of the plant remains previously described from this locality were collected *in situ*.

The plant-bearing beds are in two main zones (Pl VII, Figs 1-4). The lower zone (A), composed mainly of highly brittle shales, immediately underlies the upper hard silicified shales (B). The lower zone (A), over which the river flows during the major part of the year, is further differentiated into several bands which are, however, not clearly seen in the accompanying photographs (Pl VII, Figs 1-4). The upper zone (B), about six inches to one foot in thickness, can be traced almost horizontally along the base of the cliff.

<sup>1</sup> Sahni and Rao (1931), p. 184.

<sup>2</sup> See Sahni and Rao (1931), p. 185, footnote.

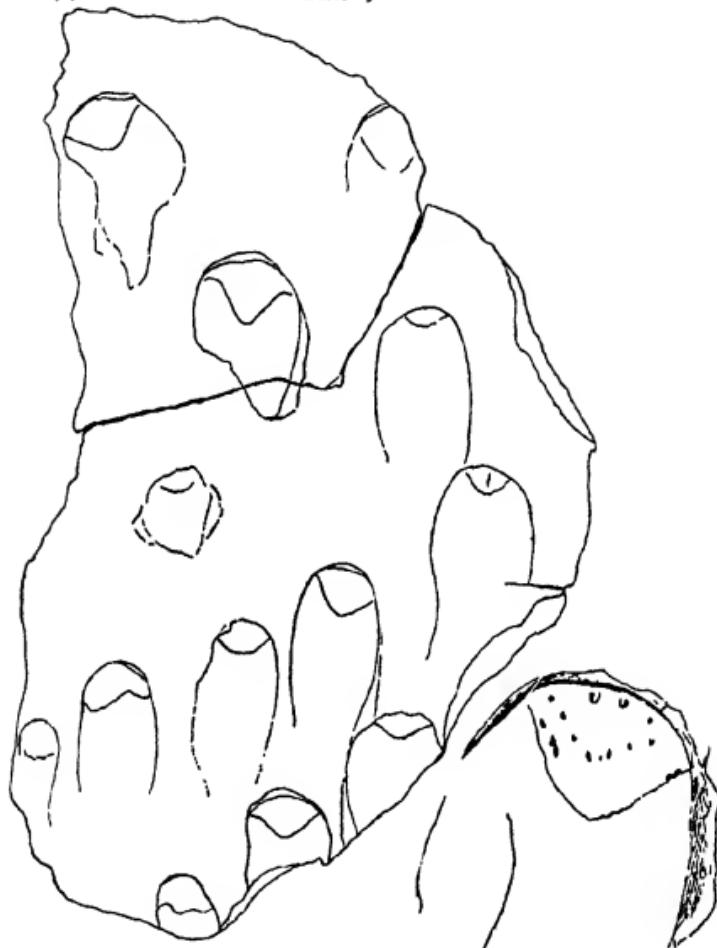


Fig.1

Text Fig. 2

TEXT-FIG. 1.—*Protocyathea rajmahalense* sp. nov. Rough sketch of the type-specimen showing the arrangement of the leaf-cushions and scars  $\times$  Natural size.

TEXT-FIG. 2.—*Protocyathea rajmahalense* sp. nov. One of the leaf-scars showing the arrangement of leaf-traces  $\times$  3

The fossils described in the present communication were found in the upper band of hard silicified shales (B), the lower zone composed of brittle shales (A), though rich in other plant remains, has not so far yielded any fossils similar to those described below.

Cyatheaceous stems were hitherto described mainly from the Cretaceous rocks. Quite recently two Upper Jurassic species from Korea were described by Ogura.<sup>4</sup> The discovery of these fossils in the Rajmahal series, which is at present considered to be probably of Middle Jurassic age,<sup>5</sup> takes us further back in the geological scale, assuming, of course, that the Sakrigali beds are not higher than the rest of the Rajmahal series. A careful geological study of this and other plant-bearing localities in the Rajmahal Hills from the stratigraphical point of view should help to clear up much doubt regarding the exact horizons to which the various members of this classical flora belong.

## 2 Description

### *Protocyathea* Feistmantel 1877 <sup>6,7</sup>

The genus *Protocyathea* created by Feistmantel is a convenient designation for all tree ferns of Cyatheaceous affinity, that are preserved either in the form of casts or impressions, and whose real affinity to any particular living genus cannot be determined owing to the absence of preserved internal structures. The vascular bundles within the leaf-scars are generally preserved as a number of warts arranged in the form of arcs. There is little doubt that, as at present understood, this comprehensive genus is highly artificial.

The type specimen of the genus, now preserved in the Museum of the Geological Survey of India (Pl. X), is a stem cast showing large spirally arranged leaf-scars with traces of a few vascular bundles within them, it was collected by H. F. Blanford from rocks regarded as Cretaceous in the Trichinopoly District of South India.<sup>8</sup>

<sup>4</sup> Ogura (1927), pp. 352-364. Ogura (1927), pp. 364-368.

<sup>5</sup> Sahni (1932), pp. 14, 15.

<sup>6</sup> The genus *Protocyathea* Fst., and the family *Protocyatheaceae* created by Bower (1926, pp. 282-292) to receive the two comparatively primitive genera *Lophosoria* and *Metasya* have no direct relationship whatever with each other.

<sup>7</sup> Feistmantel (1877), p. 136.

<sup>8</sup> See below page 82.

*Protocyathaea raymahanense* sp. nov

E 188, E 189; E 190 (unfigured), E 191 \*

(Plate VIII, Figs 5-7, Plate IX, Figs. 8-11, Text-Figs 1, 2)

Type specimen. E 188 (Plate VIII, Fig 5; Text-Fig 1).

**Diagnosis.** Stem impressions with large spirally arranged leaf-cushions or bases, each bearing at its upper end the rhomboid scar of a fallen leaf. Cushions compactly arranged on the older parts of the stem, about 5.0 to 5.5 cm by 1.7 to 2 cm, surface wrinkled. Scars roughly rhomboidal with their longer axes horizontal, about 0.9 to 1.0 cm. by 1.4 cm., vascular bundles few (about fourteen?), in two curved rows, with two large median bundles at the top in the upper row (see Text-Fig. 2). In the younger parts of the stem the cushions are greatly reduced, the petiolar scars of adjacent leaves being almost contiguous, vascular bundles not preserved.

Four specimens are known of which one (E 190) is in a very bad state of preservation and is not figured. They represent impressions of the younger as well as the older regions of the stem. In the latter, the surface of the stem is seen covered over by spirally arranged persistent leaf-bases or cushions. But due to incomplete preservation it is difficult to determine the phyllotaxy. At the top of each leaf-cushion or leaf-base is seen the scar of a fallen leaf, with small marks, no doubt of vascular bundles (Pl VIII, Fig. 5; Pl IX, Fig. 11, Text-Figs 1, 2). Below the scar the leaf-cushions exhibit a wrinkled appearance which is probably due to the scars of ramenta, though it may as well be due to a general shrinkage of the surface of the leaf-base. The leaf-cushions or bases are mostly elongated along the length of the stem (Pl VIII, Fig. 5, Pl IX, Fig. 8, Text-Fig 1). This is the condition seen in specimens which probably represent the older parts of the stem. In other specimens the leaf-scars have their long axes horizontal (Pl VIII, Fig 6; Pl IX, Fig. 8, top half), and the scars are densely crowded, apparently with hardly any trace of the leaf-cushions between them (Pl VIII, Fig. 6, b). This condition seems to represent the younger parts of the stem. A similar compact arrangement of scars is seen in some of the living Cyatheaceae\*. In Fig 6, on Pl. VIII, both these conditions are seen in one and the same specimen.

The arrangement of the relatively few vascular bundles within the scar is not complicated, it seems, however, that they are not all of them preserved, hence a clear description of their arrangement is impossible.

The four specimens are described separately below.

\* Numbers in this form indicate the specimens from Sakrigalighat (locality E, in Sahni and Rao, 1931).

\* Ogura (1927 a), Figs 35-37, 70.



Text Fig. 3



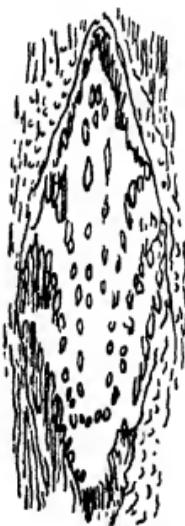
Text Fig. 4



Text Fig. 5



Text Fig. 6



Text Fig. 7

TEXT-FIG 3.—*Protocyathus cyathoides* (Unger) One of the scars showing the arrangement of the leaf-trace bundles (After Unger 1867, Taf 1, Fig 3)

TEXT-FIG. 4.—*Protocyathus trichopollenensis* Feistmantel One of the scars showing the arrangement of the leaf-trace bundles. (After Feistmantel, 1877, Pl 1, Fig 1)

TEXT-FIG. 5.—*Protocyathus cretacea* (Stens) One of the leaf-scars showing the arrangement of the leaf-trace bundles. (After Hossus und van der Marck, 1890, Taf 43, Fig 86)

TEXT-FIG 6.—*Protocyathus Tokunagai* Ogura One of the scars showing the arrangement of the leaf-trace bundles  $\times 2/3$  (After Ogura, 1931, Text-Fig. 1)

TEXT-FIG 7.—*Cyathocaulis nakatongensis* Ogura One of the scars showing the arrangement of the leaf-trace bundles  $\times 4/5$  (After Ogura, 1927, Text-Fig 1)

#### Specimen 1 (Holotype)

E. 188 (Plate VIII, Fig 5; Plate IX, Figs 9-11, Text-Figs 1, 2)

This specimen shows the older part of the stem, with thirteen persistent leaf-cushions spirally arranged (Pl VIII, Fig 5, Text-Fig 1) Each cushion measures about 5 to 5.5 cm by 1.4 to 1.7 cm, and at the top of each can be seen the scar of a fallen leaf (Pl IX, Fig 11) The leaf-scars measure about 0.9 to 1.0 cm by 1.4 cm, and show a few small marks of vascular bundles, which unfortunately are only partly preserved (Pl IX, Fig 11, Text-Fig. 2) The best preserved and completest scar is shown in Pl IX, Fig 11 As far as can be made out, about fourteen separate and wart-like vascular bundles are seen more or less regularly arranged as follows about eight bundles are arranged in an arc in the centre of the scar, and an upper row of three or four bundles runs close to the upper margin, the two median bundles in this upper row are larger than the rest (Pl IX, Fig 11, Text-Fig 2) below, and on one side of the central row of eight bundles, is seen a group of three bundles which perhaps formed part of the lowermost arc of vascular traces that are only incompletely preserved on one side Impressions of certain elongated cells (sclerenchyme or ? tracheids) are preserved in certain parts of the stem (Pl VIII, Fig 5, Pl IX, Fig 10) A few marks of adventitious roots are also seen between the leaf-cushions, but the preservation is too poor to show the details

#### Specimen 2.

E. 191 (Plate VIII, Figs 6, 7)

In this specimen the leaf-scars are densely crowded, apparently with hardly any trace of the leaf-cushions between them (Pl VIII, Fig. 6b) Lower

down, however, are seen one or two older scars with well-developed leaf-cushions somewhat similar in shape to those seen in the previous specimen (Pl. VIII, Fig. 6 a) The "wrinkling" on the surface of these leaf-cushions is also similar. It is quite possible that the specimen belongs to a younger part of the stem, where the leaf-cushions have not yet developed (cf. Ogura, 1927 a, Figs. 35-37, 70) Vascular bundles are, however, not preserved. A few impressions, probably of adventitious roots, are present. One such root is seen clearly in the side view of the specimen in Plate VIII, Fig. 7 r. The deep groove (marked *l* in this figure) which originates from one of the smaller rhomboidal scars may be the impression of an attached leaf-stalk (Pl. VIII, Fig. 7 l).

*Specimen 3*

E 189 (Plate IX, Fig. 8)

This specimen is interesting in that it shows a gradation between the upper closely appressed scars which hardly possess any trace of cushions, and the lower scars which display well-developed leaf-cushions between them (Pl. IX, Fig. 8). As pointed out above, a similar differentiation in the arrangement of the leaf scars in the younger and older parts of the stem is often clearly displayed in the living Cyatheaceæ (cf. Ogura, 1927 a, Figs. 35-37, 70).

*Specimen 4*

E 190 (Unfigured)

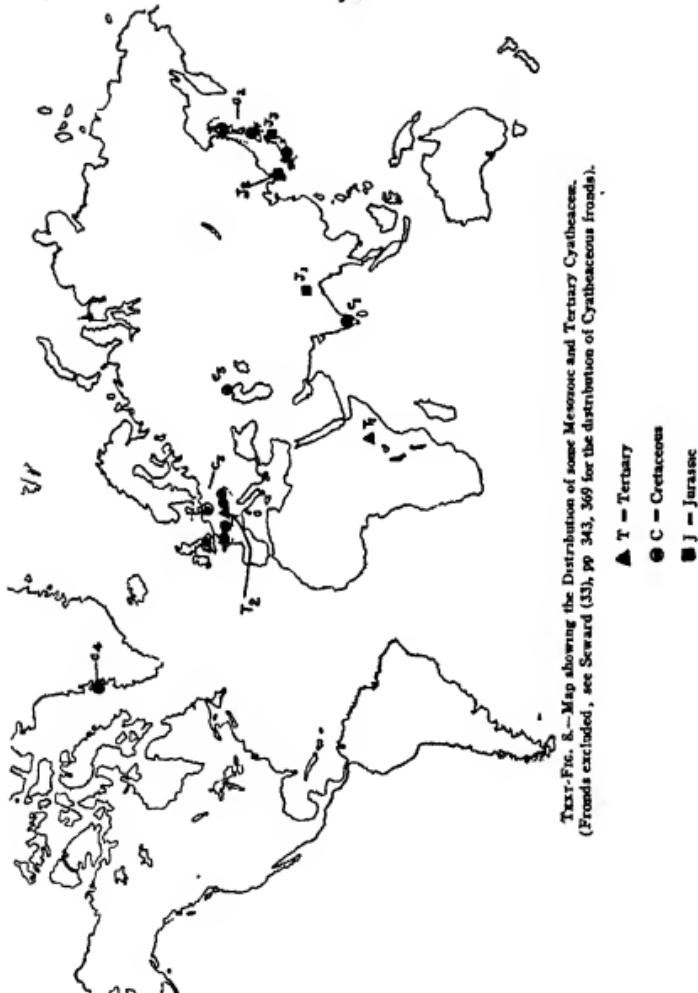
A badly preserved curved stem which shows indistinct leaf-cushions.

I have ventured to keep the above four specimens within the single species *Protocyathea rajmahalense* sp. nov. There can be very little doubt that Specimens 1 and 3 belong to the same species. Specimen 2 at first sight appears to be somewhat different from the rest, because it shows leaf-scars which possess practically no trace of cushions. But Specimen 3 displays a gradual transition from the lower scars with well-developed leaf-cushions to the upper leaf-scars with hardly any cushions. As a similar differentiation in the form of the leaf-scars of the younger and older parts of the stem is also seen in the living Cyatheaceæ, I have little hesitation in considering the above specimens as belonging to the identical species.

*3. Discussion*

(a) *Systematic position of the Rajmahal species.*

From the above description it seems fairly clear that our fossils belong most probably to tree-ferns of Cyatheaceous affinity. The large size of the leaf-cushions, the gradation between the scars of the lower and higher regions, and the arrangement of the vascular bundles, all point out—as far as mere impressions of vegetative parts can be a guide to affinity—that they are



TEXT-FIG. 8.—Map showing the Distribution of some Mesozoic and Tertiary Cyathacean.  
(Fronds excluded, see Seward (1931, pp. 343, 369) for the distribution of Cyathaceous fronds).

probably related to the modern family Cyatheaceæ. The characters of the fossils agree most nearly with those of the genus *Protocyathea* Feist.

In his diagnosis of *Protocyathea* Feistmantel<sup>14</sup> mentioned the following characters as distinctive of the genus—" *Filix arborescens, caule teretis; cicatricibus ramorum (foliorum) spiraliter dispositis, nunc maximus nunc mediocribus, structura earum cicatricibus Cyathearum venustum proxima* " And our species can best be accommodated in Feistmantel's genus, within which should also be included all stem impressions or casts of Cyatheaceous affinity which show separate vascular bundles in the leaf-scar.

As far as I have been able to judge from the published records, our fossils are not identical with any species of *Protocyathea* previously described. They are therefore referred to a new species *Protocyathea rajmahalense*.

(b) *A review of the genus Protocyathea Feist*

A detailed comparison of the Rajmahal specimens with the described species of *Protocyathea* is best carried out after a general review of the genus. Four species of *Protocyathea* have hitherto been described, and the different species are distinguished primarily by the number and arrangement of the vascular bundles within the leaf-scar, and to some extent by the shape of the leaf-cushions. The chief characters of the four species are briefly dealt with below. For a detailed study of the individual species, see also Table I.

*Protocyathea cyatheoides* (Unger) Feistmantel  
(Text-Fig. 3, Table I)

1867 *Caulopteris cyatheoides* Unger<sup>15</sup>

1877 *Protocyathea Ungerii* Feistmantel<sup>16</sup>

1900 *Alsophilina cyatheoides* Potonié<sup>17</sup>

1927 *Protopteris cyatheoides* Hirmer<sup>18</sup>

From the Lower Cretaceous (Neocomian) of Ischl in Austria. The chief characters of this species are—Leaf-scars large and spirally disposed, tapering towards the ends, leaf-trace composed of 40 to 60 bundles, somewhat irregularly arranged, as shown in Text-Fig. 3.

*Protocyathea cretacea* (Stenzel) Ogura  
(Text-Fig. 5, Table I)

1880 *Protopteris punctata* Hosius und van der Marck<sup>19</sup>  
(only figured, not described)

<sup>14</sup> Feistmantel (77), p. 136.

<sup>15</sup> Unger (67), p. 643, Pl. I, Figs. 1-4; Renault (83), p. 72, Stenzel (97), p. 16.

<sup>16</sup> Feistmantel (77), p. 136, Posthumus (31), p. 137.

<sup>17</sup> Potonié (00), p. 39.

<sup>18</sup> Hirmer (27), p. 641.

<sup>19</sup> Hosius und van der Marck (80), Pl. 43, Fig. 186.

1897 *Caulopteris cretacea* Stenzel <sup>14</sup>

1927 *Protopteris cretacea* Hirmer <sup>15</sup>

1931 *Protocyathaea cretacea* Ogura <sup>16</sup>

From the Upper Cretaceous (Senonian) of Westphalia in Germany The chief characters of this species are —Leaf-scars somewhat small, spirally arranged, leaf-trace composed of about twenty-six separate bundles, regularly arranged, as shown in Text-Fig 5

*Protocyathaea Tokunaga* Ogura

(Text-Fig 6, Table I)

1931 *Protocyathaea Tokunaga* Ogura <sup>17</sup>

From the Upper Cretaceous (Senonian) of Japan The species is characterised by about thirteen vertical rows of spirally disposed large leaf-scars each of which shows about a hundred separate and regularly arranged bundles as shown in Text-Fig 6 The arrangement recalls *Cyathocaulis naklongensis* Ogura<sup>18</sup> (Text-Fig 7), a species which has been placed under a separate genus as the internal anatomy is known

*Protocyathaea trichinopolensis* Feistmantel

(Plate X, Text-Fig 4, Table I)

1877 *Protocyathaea trichinopolensis* Feistmantel <sup>19</sup>

From the Upper Cretaceous (Cenomanian) of Trichinopoly in South India Prof L Rama Rao of Bangalore in a letter to me expresses the opinion, (which I may be allowed to quote), that the fossil probably " belongs to the Utatur group which forms the oldest sub-division of the Trichinopoly Cretaceous " The leaf-cushions are spirally disposed Feistmantel<sup>20</sup> describes the formation of a convex disc in the upper portion of the scar This disc is not well seen in the original specimen which, thanks to the kindness of the Director of the Geological Survey of India, I have been able to examine in Calcutta Apparently the leaf-scar and leaf-cushion are not clearly differentiated Each scar shows about twenty separate bundles of which nine or ten are seen arranged in a lower arc, a few bundles at the top of the scar which are seen in the type specimen, however, are indistinctly drawn in Feistmantel's figure, here reproduced as Text-Fig 4, they are somewhat

<sup>14</sup> Stenzel (97), p. 17.

<sup>15</sup> Hirmer (27), p. 641

<sup>16</sup> Ogura (31), p. 58.

<sup>17</sup> Ogura (31), p. 58, Text-Fig 1, Pl. IV

<sup>18</sup> Ogura (27), p. 352, Text-Fig 1

<sup>19</sup> Feistmantel (77), p. 136, Pl. I, Figs. 1, 2.

<sup>20</sup> Feistmantel (77), p. 136.



TABLE I.  
SIXTY-ONE HUNDRED NINETEEN FRESH SHEETS OF QUADRATIC FINE SHEET (the General Photocell Co.)



irregularly arranged in the original specimen. Certain oblong grooves are present in the lower half of the leaf-cushion (Pl. X). Feistmantel<sup>22</sup> applies the term 'stigmata' to these structures. To judge from their appearance in the original specimen they are structures of a different nature from vascular bundles. These structures may be compared with pneumatophores as figured by Bower in *Alsophila*<sup>23</sup>.

(c) Comparison with the other species of *Protocyathea*

From a brief consideration of the different species of *Protocyathea* at present included within the form genus, it is clear that the Rajmahal species shows the closest resemblance with the South Indian species *P. trichinopoliensis* Fst.<sup>24</sup> To some extent the shape and size and the arrangement of the leaf-scars in the older parts of the stem, is similar to that in *P. trichinopoliensis* Fst. The general plan of distribution of the few vascular bundles in *P. trichinopoliensis* Fst is less complicated than in the other species of *Protocyathea* (cf. Text-Fig. 4 with Text-Figs. 3, 5-7). On the other hand, the irregular arrangement of the vascular bundles above the lower arc of bundles in *P. trichinopoliensis* Fst (Text-Fig. 4) is quite different from that in the Rajmahal species, where the uppermost bundles are seen disposed quite regularly, running close to the adaxial margin (Text-Fig. 2). *P. trichinopoliensis* Fst, moreover, differs from the Rajmahal species in the absence of a clearly differentiated leaf-scar on the upper part of the cushion and of corrugations on the leaf-cushions. In the absence of 'stigmata' and in the presence of a smaller number of vascular bundles, our specimen shows further important differences from the Cretaceous species from South India. Therefore the species is described as new.

The other species of *Protocyathea* show little resemblance with *Protocyathea rajmahalense* sp. nov. The size and shape of the leaf-scars, and the arrangement of the numerous vascular bundles, both in *Protocyathea cyathoides* (Unger)<sup>25</sup> and *Protocyathea Tokunagae* Ogura,<sup>26</sup> are totally different (see Text-Figs. 3, 6). As regards *P. cretacea* (Stenz.),<sup>27</sup> although it comes nearer to the Indian species in the size of the scars and in the smaller number of vascular bundles, the arrangement of the bundles is different.

<sup>22</sup> Feistmantel (77), pp. 136, 137, Pl. I, Figs. 1, 2.

<sup>23</sup> Bower (23), p. 203, Fig. 193.

<sup>24</sup> Feistmantel (77), pp. 136, 137, Pl. I, Figs. 1, 2.

<sup>25</sup> Unger (67), Pl. I, Figs. 1-4.

<sup>26</sup> Ogura (31), Pl. IV, Text-Fig. 1.

<sup>27</sup> Hosius und van der Marek (80), Pl. 43, Fig. 186.

(d) *Geological and geographical distribution of the Cyatheaceæ*

*General*—For a clear understanding of the geological and geographical distribution of the Cyatheaceæ, records of fern fronds as well as stem remains have to be briefly considered. The distribution of fern fronds believed to be of Cyatheaceous affinity has already been treated by Seward and others,<sup>22</sup> and there is little necessity to go over that difficult ground again. Our attention is confined mainly to the stem remains preserved in the form of petrifactions, casts or impressions, which may be reasonably referred to the Cyatheaceæ (see Table II). Before entering into a discussion regarding their distribution, geological and geographical, the value of each genus as evidence of Cyatheaceous affinity should be considered. Posthumus<sup>23</sup> and Bancroft<sup>24</sup> have recently made similar attempts.

*Petrifications*—The most important and widely distributed stem genus referred to this group is *Protopteris*, which includes both petrifactions and casts.<sup>25</sup> The probable Cyatheaceous affinities of this genus can be recognised by its leaf-trace with inwardly curved ends and plicate margin. The two east Asiatic species, *Cibotocaulis Talewia* Ogura<sup>26</sup> and *Cibotium swansonii* Ogura,<sup>27</sup> resemble the living genus *Cibotium* in the mode of departure of the leaf-traces and in the arrangement of the vascular bundles in the petiolar base. *Caulopteris arborescens* Stenzel<sup>28</sup> and *Caulopteris Brownii* Renault<sup>29</sup> show to a certain extent stelar similarity with the modern Cyatheaceæ. But certain features in which the two above-mentioned species differ from the living Cyathaceous were already pointed out by Bancroft<sup>24</sup> and Rao.<sup>30</sup> The stem anatomy of *Cyathocaulis naktongensis* Ogura<sup>31</sup> can best be compared, according to Ogura, with that of the living species *Dicksonia antarctica* *Dendropteridium cyatheoides* Bancroft<sup>32</sup> shows an undoubtedly polycyclic

<sup>22</sup> Seward (33), pp. 343, 369; Seward (10), p. 367; Halle (13), pp. 17, 94; Hirmer (27), p. 637; Thomas (11), p. 387; Potonié and Gothan (21); Schenk in Zittel (90), pp. 92-95.

<sup>23</sup> Posthumus (31).

<sup>24</sup> Bancroft (32), p. 249.

<sup>25</sup> Seward (10), pp. 370-375; see also for further references.

<sup>26</sup> Ogura (27), pp. 364-368; Text-Figs. 10, 11, Pl. III, Figs. 13-15; Pl. VIII, Figs. 43-49.

<sup>27</sup> Ogura (33), p. 748; Text-Figs. 1, 2, Pl. II, Figs. 1-4.

<sup>28</sup> Stenzel (97), p. 10; Pls. I, II, III, Figs. 16-19.

<sup>29</sup> Renault (83), p. 73; Pl. VIII, Fig. 10; Rao (34), pp. 221-225; Pl. XXXIII, Figs. 3-6.

<sup>30</sup> Bancroft (32), p. 349.

<sup>31</sup> Rao (34), p. 221.

<sup>32</sup> Ogura (27), p. 351; Text-Figs. 1-9; Pl. II, Figs. 1-6; Pl. III, Figs. 7-12; Pls. IV-VI.

<sup>33</sup> Bancroft (32), p. 241; Text-Figs. 1, 2, Pls. IX, X.



TABLE II.—  
Tally of the Number of Individuals and Geographical Distribution of Some Mammals and Testaceous Crustaceans (Friends excluded) Found in the Distribution of *Ypsilon* in the Atlantic and Indian Oceans.



structure of Cyatheaceous type. The genus *Cyathorachis Fujiana* Ogura<sup>41</sup> is a well preserved piece of rachis showing numerous bundles arranged as in some modern Cyatheaceæ *Rhizodendron oppolense* Göpp.<sup>42</sup> on the other hand, is a species of doubtful affinity which has been included under the Cyatheaceæ by certain authors. It shows fibrous bundles in the cortex and pith with sclerenchymatous tissue outside the main stele. The leaf-trace has four or more strands.

*Impressions*—Among the casts and impressions of Cyatheaceous affinity *Oncopteryx*<sup>43</sup> and *Protocyathea*<sup>44</sup> are the two important genera. *Protopteryx*, as mentioned above, is also known as casts. The internal anatomy of the species included in the first two genera is at present unknown, and till their internal structure is known they cannot be removed from the present position of uncertain affinity to a more stable footing. But their affinities are no doubt more with the Cyatheaceæ than with any other group of ferns. The genus *Oncopteryx* shows, besides a ring of separate wart-like bundles, two <-shaped or C-shaped bundles at the top of the scar. A similar arrangement of the vascular bundles in the leaf-base is also noticed in some of the living Dicksoniæ. The affinities of *Protocyathea* with the living Cyatheaceæ have already been considered elsewhere in detail.

From the above brief discussion it would appear that as far as their anatomy is concerned, the Cyatheaceæ have descended from their Mesozoic ancestors with but little modification.

Table II is intended to illustrate the distribution in space and time of such fossil stems as may be reasonably regarded, on data at present available, as members of the Cyatheaceæ.

The geographical distribution of the living Cyatheaceæ has been ably dealt with by Diels<sup>45</sup>. Their distribution in a broad belt throughout the tropics and sub-tropics of the Old and the New Worlds forms a striking contrast to their distribution in the past.

According to Seward<sup>46</sup> "We have as yet no satisfactory evidence of the existence of the Cyatheaceæ in Palæozoic flora."

<sup>41</sup> Ogura (27), p. 368. Text-Figs. 12, 13, Pl. VIII, Figs. 50-51.

<sup>42</sup> Göppert (65), p. 397. Stenzel (86), p. 5, Pl. I, Figs. 1, 3, 5-12, Pl. II, Pl. III, Figs. 20-29, Rao (34), pp. 225, 226.

<sup>43</sup> Krejci (53); Fenstmantel (72); Velenovsky (88); Potonié (90); Frič und Bayer (01); Seward (10); Pelourde (11); (14); Engelhardt (81); Stenzel (97); Hirmer (27); Ogura (31); Velenovsky and Viníkiař (29).

<sup>44</sup> See Table I for full references.

<sup>45</sup> Diels (02), pp. 117, 122, 124-138.

<sup>46</sup> Seward (10), p. 366.

*Rhaetic*.—It is not till the Rhaetic period is reached that we find anything like definite evidence of this family of ferns, and in these rocks, too, they have so far been found only in the form of leaf impressions<sup>47</sup>. The only Rhaetic record so far known is from Tonkin.

*Jurassic*.—During the Jurassic, however, the Cyatheaceae had a world-wide distribution. We have several undoubted records of Cyatheaceous ferns (both stem remains and fronds). Of petrified stems, the earliest reliable evidence so far available was from the Upper Jurassic, namely, *Cibotocaulis Tatewae* Ogura<sup>48</sup> from Korea, and *Cyathocaulis naklongensis* Ogura<sup>49</sup> from Japan. The discovery of *Protocyathea rajmahalense* sp. nov. from the Rajmahal series, which is at present believed to be of Middle Jurassic age,<sup>50</sup> therefore takes us further back in the geological scale (see Table II). In the Jurassic rocks fern fronds are more commonly found than stem remains. *Oncopteris hymenophylloides* (Brong.) and *Eboracis lobifolia* (Phil.)<sup>51</sup> were well represented in the vegetation of this period. Another fern possibly belonging to the same family was *Stachypteryx*<sup>52</sup>.

*Cretaceous*.—The Cretaceous records are mainly confined to the stem remains, the largest number being found in the Upper Cretaceous (see Table II). The fronds which were abundantly represented in the Jurassic are singularly rare in these rocks. But there is enough evidence to show (from the stem remains so far known), that the Cyatheaceae were more or less well represented during this period as well (see Table II), though probably not to such a great extent as in the Jurassic. *Protopteris punctata* Stern described by Heer<sup>53</sup> from the Cretaceous of Greenland, establishes the extreme northerly distribution of the group *Protocyathea trichinopoliensis* Fst from the Cretaceous rocks of South India, also shows the wide range of the family during that period.

*Tertiary*.—There is great scarcity of Cyatheaceous records of any type after the Cretaceous. Except for two species of stems, *Oncopteris Laubers* (Engelb.) probably from the early Tertiary of Bohemia,<sup>54</sup> and *Dendropteris cyatheoides* Banc from the late Tertiary of E. Africa,<sup>55</sup> there is no evidence of a reliable nature.

<sup>47</sup> Zeiller (03), p. 36, Pl. IV, Fig. 1, Halle (13), p. 94, Seward (33), pp. 343, 350

<sup>48</sup> Ogura (27), p. 364

Ogura (27), p. 351

Sahm (32), pp. 14, 15.

See Halle (13), for full reference, pp. 19-21 *Oncopteris hymenophylloides*, pp. 16-17 *Eboracis lobifolia*, Hirmer (27), pp. 637-639

Thomas (12), Seward (33), p. 350.

Heer (82), Pl. XLVII

Engelhardt (81), p. 284, Taf. 1, Figs. 1-4

Bancroft (32), p. 241

**Conclusion**—Thus, if we consider the records of stem remains and frond impressions collectively as evidence for the existence of Cyatheaceous ferns, it can safely be said that the group was abundantly represented in the vegetation of the Jurassic and Cretaceous periods, and enjoyed an almost cosmopolitan existence. By the end of the Cretaceous they had already started disappearing from the northern regions, and during the period that followed, in company with other groups of ferns such as the Gleicheniaceae and the Marattiaceae, they became confined to their present restricted distribution within the tropics and south temperate zone.

(e) *Geological age of the Sakrigalighat beds*

A definite opinion on the geological age of the beds cannot be ventured at this stage. It will, however, be discussed after the other specimens from this locality are examined and described. But it may be mentioned that *Protocyathea* is a genus previously recorded only from the Cretaceous rocks.

**4 Summary**

1 *Protocyathea rajmahalense* sp. nov. is described from certain beds at Sakrigalighat, in Behar.

2 The earliest recorded species of Cyatheaceous stems are from the Upper Jurassic of Korea. The discovery of the Indian species takes us further back in the geological scale, because the Rajmahal series is probably not so young as the Upper Jurassic, unless of course, the Sakrigalighat beds are higher than the rest of the Rajmahal series, a question which deserves the attention of geologists.

3 The affinities of the Rajmahal species with the other species of *Protocyathea* are discussed. The closest resemblance is with *Protocyathea trichopolensis* Fst from the Cretaceous of South India. Due to certain important differences from the latter, the Rajmahal form is described as a new species.

4 The distribution of the Cyatheaceae, both living and fossil, is briefly discussed.

**5 Acknowledgment**

My grateful thanks are due to Professor B. Sahni, F.R.S., for his invaluable and most generous help and criticisms throughout the course of this work. For his personal inspiration, which has been a constant source of encouragement, and for all that my association with him for the last few years has meant for me, which is much more than I can mention here, I wish to express my deepest gratitude.

To the Director, Geological Survey of India, I am indebted for permission to examine the type specimen of *Protocyathus trichinopolensis* at Calcutta, and to reproduce a photograph of it (Pl. X).

I am also indebted to Prof L. Rama Rao of the Department of Geology, Central College, Bangalore, for valuable information regarding the probable stage of the Trichinopoly Cretaceous to which *P. trichinopolensis* Pat. belongs.

My thanks are also due to the authorities of the Lucknow University for the award of a Research Fellowship during the course of this work.

#### BIBLIOGRAPHY

Bancroft, H "A fossil Cyatheoid stem from Mount Elgon, East Africa," *New Phytologist*, 1932, 31

Bower, F O "The Ferns (Filicales)," 1923, 1 Cambridge Botanical Handbooks.

— "The Ferns (Filicales)," 1926, 2 Cambridge Botanical Handbooks.

Diels, L "Cyatheace," in Engler and Prantl, *Die Naturlichen Pflanzenfamilien*, Leipzig, 1902, Teil 1, Abt. 4

Engelhardt, H "Ueber die fossilen Pflanzen des Süßwassersandsteins von Grassee," *Acta der Kaiserlichen Deutschen Akademie der Naturforscher*, 1881, Band 43, 4.

Feistmantel, O "Notes on the fossil floras of India, 9. On a tree-fern stem from the Cretaceous rocks near Trichinopoly in Southern India," *Rec. Geol. Surv. Ind.*, 1877, 10

\*Fric, A., and Bayer, E "Studien um Gebiete der böhmischen Kreideformation," *Arch für die Naturwissenschaften und Landesforschung von Böhmen*, 1901, Band 11, 2

Göppert, H R "Ueber das Vorkommen von Baumfarren in der fossilen Flora, ins besondere in der Kreideformation," *Neues Jahrbuch für Min. Geol. Paläont.*, 1865

Gothan, W "Rhacodendron opolense Göppert (nach der anatomischen Structur) Alsophila spec (nach der Oberflächen-skulptur des Stammes), in H Potonié," *Abb. und. Beschreibung Pflanzen*, 1904, Lieff. 2.

\*Heer, O "Die fossile Flora Grönlands I," *Flora Fennica Arctica* 1882, 6

Hirmer, M "Handbuch der Paläobotanik, I," München und Berlin, 1927

Hosius, A., and van der Mark, W. "Die Flora der Westfälischen Kreideformation," *Palaeontographica*, 1880, 26

Ogura, Y "On the structure and affinities of some fossil tree-ferns from Japan," *Journ. Fac. Sci., Imp. Univ. Tokyo*, 1927, Sec. 3, 1, Pt. 3.

— "Comparative anatomy of Japanese Cyatheaceae," *Journ. Fac. Sci. Imp. Univ. Tokyo*, 1928, Bot., 1, Pt. 2.

\* Not seen by me.

*Cyatheaceous Tree-fern* *Protocyathea rajmahalense* sp. nov. 89

Ogura, Y. "On a fossil tree-fern stem from the Upper Cretaceous of Iwaki," *Jap Journ Geol Geogr*, 1931, 9, No 1

Posthumus, O. "Catalogue of the fossil remains described as fern stems and petioles, Java," 1931

Potonić, H. "Ueber die fossilen Filicales im Allgemeinen und die Reste derselben zweifelhafter Verwandtschaft," in Engler und Prantl, *Die Natürlichen Pflanzenfamilien*, 1900, Teil 1, Abt 4

Potonić, H., und Gothan, W. "Lehrbuch der Palaeobotanik," Berlin, 1921

Rau, A. R. "Notes on the anatomy of some silicified ferns from the Cretaceous of Germany," *Proc Ind Acad Sci*, 1934, 1, No 5

Renault, B. "Cours de Botanique Fossile," 1883, 3, Paris

Sahn, B. "On a petrified *Hillmannia* *W. Sewardiana* sp. nov. from the Rajmahal Hills," *Pal Ind* 5 S 1932, 20 Mem 3

Sahn, B., and Rao, A. R. "On some Jurassic plants from the Rajmahal Hills," *Journ Proc Asiatic Soc Bengal*, N.S., 1931, 27, No 2

Saporta, G. de. "Notice sur les Végétaux fossiles de la Craie inférieure des environs de Havre," *Mém Soc Géol Normandie*, 1877, 6

Schenk, A. "In Zittel's Handbuch der Palaeontologie" 1890 Abt 11 München und Leipzig

Schümper, W. Ph. "Traité de Paléontologie Végétale," Paris 1869 Atlas 1870-72

Seward, A. C. "Fossil Plants," 1910, 2, Cambridge

Stenzel, K. G. "Plant life through the ages," 1933 2nd edn, Cambridge

"*Rhaeodendron opolense* Gopp," *Jahrest Schles Gesell Naturl Cultur*, 1886, *Ergebnisheft*, 63

"Verkieselte Farne von Kamenz in Sachsen," *Mitt Miner Geol Museum, Dresden*, 1897, Heft 13

Stopes, M. C. "Catalogue of the Mesozoic plants in the British Museum," The Cretaceous Flora, 1913, Pt 1, London

Thomas, H. H. "On the spores of some Jurassic ferns," *Proc Camb Phil Soc* 1911, 16 Pt 4

"On *Stachypterus Hallei* a new Jurassic fern," *Proc Camb Phil Soc* 1912, 17, Pt 4

Unger, F. "Kreidepflanzen aus Oesterreich," *Sitz Katz Akad Wiss, Wien, Math-Phys Klasse*, 1867, LV

Velenovský, J. "Die Farne der bohmischen Kreideformation," *Abh Kon böhm Ges Wiss* 1888, Folge 7, Band 2

Velenovský, J., und Vinklář, L. "Flora Cretacea Bohemie," *Nove Bistrage Bohem Kreideflora*, 1929, Teil 3

Zenler, R. "Flore fossile des Gîtes de Charbon du Tonkin," 1903, Paris.

## EXPLANATION OF PLATES

All figures are untouched photographs. With the exception of the original of Plate X, all the figured specimens come from Sakrigalighat, and are preserved in the Department of Botany, Lucknow University.

## PLATE VII

FIG. 1—Photograph showing the band of hard silicified shale (Zone B) which runs along the base of the cliff, Sakrigalighat. All the specimens of *Protoxylea rajmahalense* here described were collected from this stratum.

FIG. 2—A view of the same zone (Zone B) in the month of October when the river is in floods. The soft brittle strata (Zone A) are submerged.

FIG. 3—A view of the soft strata (Zone A) from the north-west. The bed of hard silicified shales (Zone B) can be seen in the background overlying the soft shales.

FIG. 4—Photograph showing the soft shales (Zone A) dipping north into the river. A view from the south-east, when the water has receded. The man is seen standing at the spot which yielded the major part of the collections from this zone (to be described in a subsequent paper). The hard band is seen higher up at B, also dipping north.

## PLATE VIII

FIG. 5.—*Protoxylea rajmahalense* sp. nov., showing the leaf-cushions and scars  $\times 5/6$ . Specimen 1 (E 188) a, b, two of the leaf-cushions magnified in Pl. IX, Figs 11 and 9 respectively.

FIG. 6.—*Protoxylea rajmahalense* sp. nov., showing two types of leaf-scars a, scars with well-developed leaf-cushions, b scars with more or less undeveloped leaf-cushions. Natural size. Specimen 2 (E 191).

FIG. 7.—*Protoxylea rajmahalense* sp. nov. A side view of Specimen 2 (E 191), figured in Plate VIII, Fig. 6, showing on the left an adventitious root (r), and on the right a leaf-stalk (?) Natural size.

## PLATE IX

FIG. 8.—*Protoxylea rajmahalense* sp. nov. Specimen 3 (E 189). Slightly reduced.

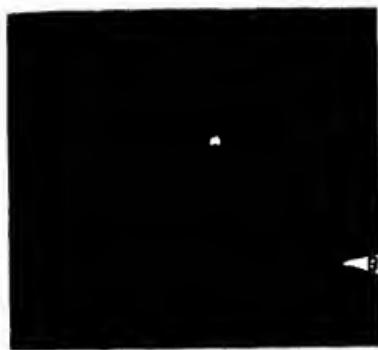
FIG. 9.—*Protoxylea rajmahalense* sp. nov. One of the leaf-scars marked (b) in Pl. VIII, Fig. 5, enlarged, showing imprints of scales or hairs  $\times 2/6$ .

FIG. 10.—*Protoxylea rajmahalense* sp. nov., showing imprints of sclerenchyma or tracheids  $\times 4$ .

FIG. 11.—*Protoxylea rajmahalense* sp. nov. One of the leaf-cushions marked (a) in Pl. VIII, Fig. 5, enlarged, showing the leaf-trace bundles and imprints of scales or hairs below the leaf-scar  $\times 3$ .

## PLATE X

FIG. 12.—*Protoxylea trichinopolitana* Fst. The type specimen (from the Cretaceous of S. India) preserved in the museum of the Geological Survey of India. From a photograph supplied by the Geological Survey of India. Natural size.



1



2



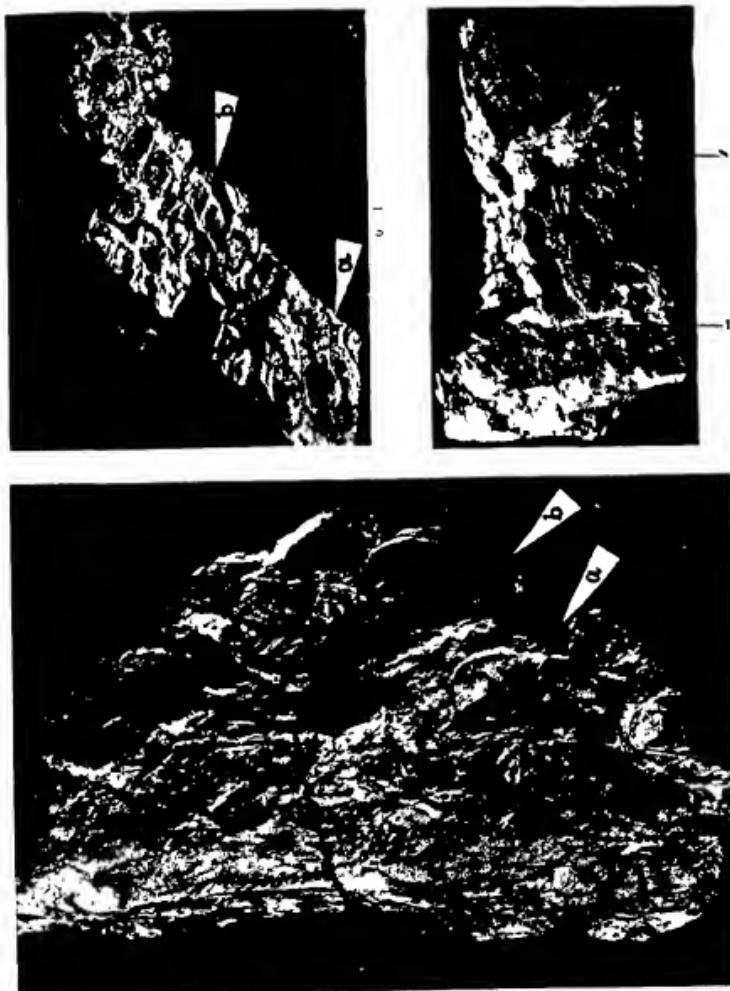
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4

4









10 - 3

11 - 3







## STUDIES IN THE PHYSICAL AND CHEMICAL PROPERTIES OF SOME SUGARCANE SOILS.

BY A. I. SUNDAR RAO, B.Sc. (HONS.)

(Allahabad)

Received June 8, 1937

(Communicated by Dr A. L. Narayan, D.Sc., F.I.P.)

ALTHOUGH it is not easy to find the cause which accounts for the fact, it is well known that the growth of sugarcane in many places becomes poorer year after year. Though the cultivation operations improve the physical condition of the soil and increase the availability of plant nutrients, there is a limit however to the benefit which can thus be obtained. In the middle of the last century, Reynoso<sup>1</sup> working on certain sugarcane soils, finds that the most robust canes are grown in calcareous soils and that these also afford juices rich in sugar. From that time many investigators have worked on this problem. Notable among these being Harrison<sup>2</sup> working on British Guinea soils, Crawley<sup>3</sup> on Cuban soils and Burgess and Kelley<sup>4</sup> on Hawaiian soils. But none of these workers seem to have tried to co-ordinate and find out the cause of this deterioration of the soils and of the presence of poorly grown canes. Isaburo Wado and Sunao Ato<sup>5</sup> tried to co-ordinate the results of analysis of some of the robust and poorly grown canes and some of the "fertile" and "infertile" soils supplied to them by the Eusuke Sugar Manufacturing Co., but their results do not seem to be conclusive.

The chemical changes that proceed in a well-cultivated soil are essentially of the nature of oxidation. Hence it appears quite possible that if the changes can be accelerated by certain chemical treatments, better plant growth and greater increase in yield may be expected. During recent years there has been an increasing evidence to show that many of the elements which have been hitherto regarded as "unessential", exercise marked influence on the plant growth. Subramanyan and collaborators<sup>6</sup> have shown in a series of papers that the organic matter either belonging to the soil or that which is added as manure undergoes decomposition yielding mineral nutrients in available form. They have also shown that the decomposition which is rather slow under natural conditions can be considerably hastened by the addition of mild oxidising agents resulting in the larger release of plant nutrients and larger yield of crop.

According to Leibig<sup>7</sup> the productivity of a soil is not however so much governed by the combined effect of all controlling factors as by the influence

of one decisive feature. These trace elements may effect the availability of soil nutrient reserves or when applied with ordinary chemical fertilisers may increase their effect—even rendering them of benefit where they normally fail. Among the various workers who have contributed to this line of research mention must be made of Bertrand and co-workers,<sup>8</sup> Warrington,<sup>9</sup> Somer and Hass<sup>10</sup> and Reid who have shown that minute quantities of F, I, Zn, Al, Mn and B are essential to the normal growth of plant. The great importance of traces of Mn for the plant has been demonstrated by McHargue, McLean, Kelley and Gerrestsen<sup>11</sup> Kelley while studying the Hawaiian soils, observed the presence of large quantities of Mn and Ti while Gerrestsen believes that Mn intensifies photosynthesis by accelerating the oxidation processes connected with the photochemical reactions in the leaf, shortage of Mn resulting in a retarded carbon dioxide assimilation. The occurrence of Mo and V in nature has been well studied by Muelen,<sup>12</sup> Dingwall,<sup>13</sup> Horner<sup>14</sup> and others. These investigators have observed that applications of Mo will increase at times the growth of *azotobacter* cultures two- or three-fold.

Recently, Dhar and his collaborators<sup>15</sup> investigating the application of molasses to the soil and the consequent photomineralisation, observe that in tropical soils the fixation of atmospheric nitrogen by the addition of energy-rich compounds is photochemical and catalytic, and that compounds of Mn, Zn and traces of Ti greatly facilitate the oxidation reactions occurring in the soil.

In connection with his recent experiments on the utilisation of molasses as fertiliser, the author had occasion to investigate the physical and chemical characteristics of some typical soils from cane-growing areas. It is proposed here to give the results of the investigation which mainly deal with fertile and infertile samples from the same locality. After years of cultivation and manurial treatment it was found that in the same locality some soils were unproductive for sugarcane cultivation while the others responded well to manurial treatments, as indicated by the return in the quantity as well as the quality of the cane. On this basis the soils (0 1' depth) were kindly supplied to the author by Mr S. S. Patrudu, Superintendent of the Agricultural Research Station, Anakapalli, as representing "fertile" and "infertile" fields in adjacent blocks.

*Physical Studies*—The soil-water relationships are largely dependent upon the textural composition of the soil. The mechanical composition as a method of expressing the texture of soils has been generally recognised. But in the ordinary methods the dispersion effected seems to be inadequate to separate the soil colloids from the mineral particles.

Samples from a fertile and infertile zone are analysed for a number of physical properties, including a study of the spectra emitted under suitable conditions by these soils. The samples (from the fertile and infertile regions obtained from a sugarcane field near Vizagapatam) contained the following main fractions —

Clay (0.002 mm & below)	15.12 %
Silt (0.02 to 0.002 mm)	6.15 %
F Sand (0.2 to 0.02 mm)	44.12 %
C Sand (2.0 mm to 0.2 mm)	33.10 %

These soils occupy a large proportion of the cultivated and uncultivated areas of the tract. The absorptive capacities of the soils were next studied in squat form weighing bottles. About 10 gm of the sample is taken in the weighing bottle and exposed to sulphuric acid-water mixtures in vacuum desiccators,<sup>14</sup> which were kept in a thermostat at 30° C. Constancy of weight was attained after a period of forty-eight hours. But they are usually exposed for not less than three days and the moisture content determined in a hot-air oven at 100° C. The results obtained for duplicate samples from the fertile and infertile regions are given below for the relative humidities at which the experiment was done.

TABLE I

Sample	Rel. Hum. 20%	40%	60%	80%	100%
Fertile 1	2.43	1.01	5.73	7.01	9.98
	2.31	4.00	5.80	7.05	10.12
Infertile 1	0.52	1.63	2.21	3.01	5.13
	0.56	1.68	2.27	3.00	5.18

(For the hygroscopic moisture at 100% R. H. water was used in the desiccator instead of sulphuric acid.)

It is interesting to note that though the mechanical composition of the soils was practically the same for the two fields yet the powers of absorption of water varied considerably and it is here that we can seek for an explanation for the difference in fertility of the two fields.

The absorption of water vapour by the soils when exposed to sulphuric acid of 33 per cent strength at 28° 5 C giving a relative humidity of

98 per cent was studied over a period of 50 days in order to see whether these differences in the absorptive capacities persist. The results of this experiment are given in Table II below.

TABLE II  
*Moisture Content at Different Periods*

Time in days	0	5	10	15	20	25	30	35	40	45	50
Sample 1 . .	4.2	9.12	10.5	11.4	11.8	12.0	12.1	12.1	12.2	12.2	12.2
Sample 2 . .	1.4	4.2	5.3	5.8	6.2	6.4	6.5	6.6	6.6	6.6	6.6

Sample 1 is from a fertile field and the sample 2 is from the infertile area. Duplicates agreed well, hence a mean value only is given above.

The absorption is very rapid at first and then the rate diminishes. The rate of absorption follows an exponential relationship with time of the type

$$R = k (1 - ae^{-kt})$$

The equation when applied to the absorption of moisture by the soil samples given above becomes

$$R = 12.2 (1 - 0.65 \times 10^{-0.004t}) \text{ for sample 1 and}$$

$$R = 6.7 (1 - 0.79 \times 10^{-0.007t}) \text{ for sample 2, where}$$

R is the moisture content in time *t* days

It is clear from the above that the higher absorptive capacities displayed by the samples from the fertile field might be due to the presence of higher amount of colloidal in these samples.<sup>17</sup>

A determination of the colloidal matter present in the soil samples was made using the method due to G. J. Bouyoucos<sup>18</sup> known commonly as the hydrometer method. The duplicate samples from the two fields gave the following values at the laboratory temperature

Fertile . (Sample 1) 21.12% colloids

(Sample 2) 20.03% "

Infertile . (Sample 1) 12.21% "

(Sample 2) 10.13% "

The above values confirm the observations made earlier that the differences in the absorptive capacities might be due to the different amounts of colloids present in the samples.

The Chemical Analysis of the above samples was done by the methods of the A.O.A.C. It is surprising to note that with the exception of Lime the major constituents did not vary in the two blocks.

	Fertile block	Infertile block
Insolubles	74.12	83.17
Iron and Alumina	6.73	7.42
Lime as CaO	12.10	6.52
MgO	1.52	1.56
Na <sub>2</sub> O	0.21	0.27
K <sub>2</sub> O	0.64	0.55
P <sub>2</sub> O <sub>5</sub>	0.09	0.07

Organic carbon and total nitrogen<sup>19</sup> in the samples was found to be as follows

	Fertile	Infertile
Organic Carbon	1.20	1.05
Total Nitrogen	0.0475	0.0398

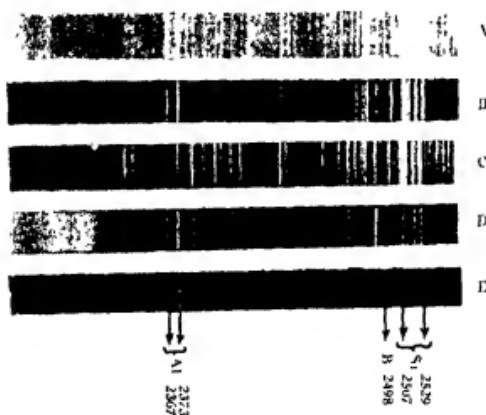
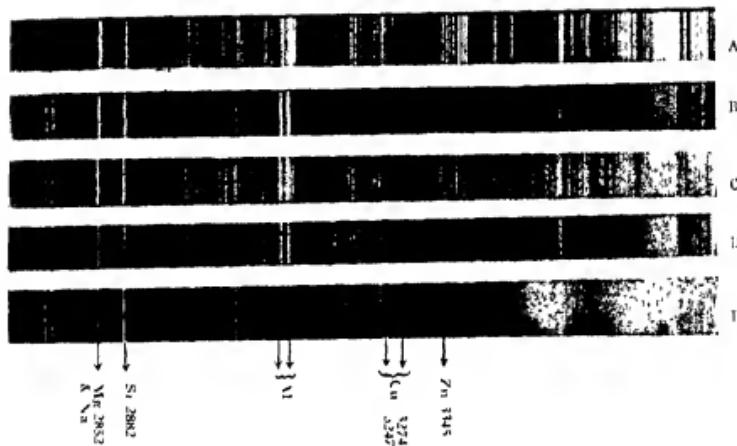
The above values do not conclusively show any great differences in the major constituents of the soils including the total nitrogen and organic carbon, for the fertile and infertile blocks, at least as much as to be able to account for the observed differences in the absorption towards water. The experiments conducted by the author show beyond doubt that these differences in the powers of absorption are due to the differences in the colloidal content. In this connection attention may be drawn to the views of F. J. Alwaye,<sup>20</sup> P. L. Gile<sup>21</sup> and their co-workers. "The colloidal material of the soils", according to Emile Troug<sup>22</sup>, "is usually largely mineral and rarely is 10 per cent or more of it organic".

For reasons stated above a search for the mineral constituents including the trace elements was made. The work reported here is of a preliminary nature, as far as the section on the spectrographic analysis is concerned, which was mainly undertaken with a view to standardise the technique employed in the analysis of soils. Concentrations of the trace elements are usually so minute that their detection and estimation would require

profound study of methods followed by laborious research in large quantities of the material

Spectrographic investigations of trace elements avoid these difficulties and the process of obtaining an ultimate mineral analysis of a soil becomes very simple. The spectrograph can present successive stages of a profile on a single plate revealing at a glance variations in the quantity of an element. A comparison of the intensities of the lines in the various spectra leads to fairly reliable results. The author has therefore photographed the arc spectra of the soil samples and this preliminary investigation is mainly intended to finding all the metallic elements contained in the soils without allowing even the rare ones to escape detection. For this purpose, soils previously prepared according to the official method and reduced to fine powder were used in a carbon arc, using for the purpose hollowed out carbon electrodes filled with the samples and a direct current of five amperes. A slightly different form of carbon arc which was found to be specially serviceable in obtaining the spectra of metals having low boiling points was used. In this the upper carbon is surrounded with a water-box through which there is kept a constant flow of cold water which prevented the temperature of the electrode from becoming too high. The spectra were photographed with a Hilger quartz spectrograph and standard spectra were impressed on each plate using for the purpose samples made for pure metals and chlorides of metals. The carbon electrodes used in these experiments were of the H S brand purity supplied by Adam Hilger. In all cases they were mounted vertically and were brought together with a gap of about 2 mm and the arc was struck by drawing a third carbon electrode of the same kind across the gap and the current was maintained constant for all the exposures.

The spectra are reproduced in Plate XI in which 'A' is the spectrum obtained with the mixture containing Na, K, Ca, Mg, Cu, Fe, Si, Al, and Ti, Mn, Zn, V in suitable ratio. 'B' and 'C' are spectra of fertile soils and 'D' and 'E' for the infertile samples for the same locality. The minerals which could be indubitably detected and identified from these are Na, K, Cu, Mg, Ca, Al, Si, and Fe and the trace elements Zn, Ti, Mn and B while the presence of Be is suspected. By comparing the spectra of these soil samples with those of a series of suitable ratio powders of known composition attempts have been made to determine the proportion of the minor constituents. By this method, the Mn content of the majority of the fertile soils was found to range from 0.04 to 0.15 while the value of Zn ranged from 0.03 to 0.06. While the problem is evidently more complicated than would at first sight appear, there seems to be nonetheless a connection between Zn, Mn and Ti content and fertility.





Valuable information might be obtained by the spectroscopic examination of soils and the ashes, roots and stems and leaves of plants that grow in these soils supplemented by the chemical estimation of the trace elements. Further experiments in this direction are in progress and the author proposes to deal with these in another communication.

#### Summary

A detailed study of the physical and chemical properties of some fertile and infertile soils from sugarcane-growing areas has been carried out. The study involves the determination of the colloid content, chemical composition and spectroscopic examination for the minor constituents. Although there has been no significant difference in soil composition, attention is drawn to the role of trace elements Zn and Ti in plant nutrition. Are spectra of these soil samples have been photographed and by comparing these spectra with those of a series of suitable ratio powders of known composition attempts are made to determine the proportion of the trace elements.

#### REFERENCES

- 1 Noel Deer, *Cane Sugar* 1921, p 68
- 2 *Ibid.*, 1921, p 70
- 3 *Estonian Central Agronomica, Cuba Boletin* 28
- 4 *HSPA Exp Sta Agric Series Bull* 45
- 5 *Soc Papers Phys and Chem Research Inst Tokyo* 1923 2 113-24
- 6 *Proc Ind Acad Sci, Sec B*, August 1934, 1  
*Nature* 1933, 132, 1001  
*Curr Sci.* 1933, 2, 161  
*Curr Sci.*, 1934, 2, 384
- 7 Noel Deer, *Cane Sugar*, 1921, p 67
- 8 *Compt Rend* 1897, 124, 1932, 1335, *ibid* 1911 152, 225 1905, 141, 1255.  
1921, 173, 333 1118, 1922, 174, 491
- 9 *Ann Bot* 1923, 37, 629 *ibid* 1926 40, 27
- 10 *Bot Gaz* 1927, 83, 77
- 11 *Trans 3rd Int Congress of Soil Science*, 1 189-91
- 12 *Rec Trans Chin* 1932, 51, 549
- 13 *Canad Jour Research*, 1934, 11, 32
- 14 *Jour Agric Res* 1934, 48, 981
- 15 *Proc Acad Nat Acad Sci*, Dec 1935, p 30, Jan 1937, p 42  
*Jour Ind Chem Soc* 1934, 11, 727
- 16 *Jour Ind and Eng Chem*, 1921, 13, 326
- 17 *Trans 3rd Int Congress of Soil Science* 1935, 2, 40
- 18 *Sci* 1927, 66, 16
- 19 Robinson, McLean and Williams, *J Agric Soc*, 1929, 19, 315
- 20 F J Alwave and Mac Dole, *Jour Indi Res*, 1917 9, 27
- 21 *J American Soc Agr*, 1925, 17, 275
- 22 Emil Troug, "Colloidal Chemistry of Soils," *Colloid Symposium*, 111, 228

ON THE NATURE AND IDENTIFICATION OF  
SOME ROUNDISH BODIES FOUND EITHER FREE  
OR AS ENDOGLOBULAR PARASITES IN THE  
BLOOD OF *CALOTES VERSICOLOR* DAUD.  
SUBSPECIES *MAJOR* BLYTH.

BY PROF COL I FROILANO DE MELLO

AND

CAETANO CORRÊA DE MEYRELLER

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Received July 13, 1937

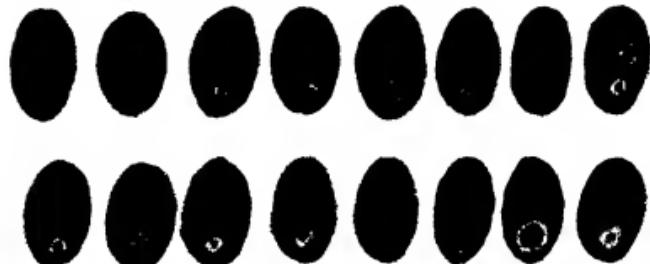
AMONG numerous specimens of *Calotes versicolor* subsp *major*, one of them showed us some minute, roundish, curious parasites which will be described in this note. In living condition, in hanging drop preparations, they appear as circular bodies, with refringent greenish protoplasm, surrounded by a rather strong membrane and provided to a certain extent with a kind of movement which seems brownian. The red globule has sometimes in its interior 2 or 3 of these bodies, but does not show any alteration. Similar bodies are also found free and some of them, either free or endoglobular, have a small bud attached to the mother-cell, as it happens in yeasts.

Stained by Leishmann's solution or by May-Grunwald-Giemsa, these bodies appear under various morphological aspects, figured in our coloured plate and which can be described in the following way:

- 1 Small round chromatic dot, of anaplasmodoid nature (Pl XII, Fig 2)
- 2 Small roundish body surrounded by a more or less strong membrane, taking a chromatic stain, and possessing a central nuclear granule, more or less developed (Pl XII, Figs 1, 3, 4)
  - (a) the membrane may have two or three chromatic granules, attached to it, moreover the central nucleus (Pl XII, Fig 8),
  - (b) the body may be devoid of any granule, resembling a mere vacuole, surrounded by the membrane (Pl XII, Figs 11, 12)
  - (c) the central granule may be located on the periphery (Pl XII, Fig 10) and show a kind of budding (Pl XII, Figs 15, 16)
- 3 In all figures described under alinea 2 the interior of the corpuscle or its protoplasm was colourless. We have now figures where the protoplasm is stained blue or grey blue, often surrounded by a white circular halo

(Pl. XII, Fig. 13) without any granule at all (Pl. XII, Fig. 6), with a central chromatic granule (Fig. 13) or the nuclear mass more or less peripheric (Figs 6, 7). It is not rare to find globules as figured in our Fig. 9, where it seems that the parasite is almost to penetrate in the red cell. The general form is circular, but often, specially when there is an appearance of budding, the form may be elliptic, elongated.

Some blood smears were stained by iron-haematoxylin of Heidenhain after wet fixation by Schaudinn's sublimate and Bouin. The general structure is the same as that found in Romanowsky's stain either anaplasmod, or of a corpuscle with a more or less conspicuous membrane, surrounded or not by a halo and possessing in its interior, vacuolated or with a slight trace of protoplasm, a chromatic granule more or less developed. This chromatic granule which acts as nucleus may sometimes show the appearance of a ring.



What becomes more evident by iron-haematoxylin staining is the process of division of this organism. The nucleus constituted by a small minute granule is enlarged and takes a ring form. Further on it becomes compact and gives origin to a small bud which makes a kind of protrusion to the exterior. One part of the membrane surrounds the nuclear bud and in subsequent stages the cytohesis occurs.



Schema of the Division.

What is the nature of these bodies? Have they been found and described by other authors?

Consulting the literature at our disposal we see in Dutton, Todd and Tobey their Fig. 55 of Plate XXV labelled as "the unidentified parasite of snake-round form" and Figs 106, 108, 110, 111, 114 and perhaps 104 of the Plate XXIX, all these labelled as *Cytamæba*, which seem to us of the same nature as our actual parasite. Dutton and co-workers classified them as *Cytamæba*.

This name was employed by Dahlewsky (1890) as syn. of *Plasmodium*. Labb  (1894) gave the name *Cytamæba bacterifera* for some peculiar bodies in the red cells of *Rana esculenta* which have been named *Bacillus krusei* by Laveran in 1899. Having come across with the so-called *B. krusei* some years ago, the senior author can assure that their nodulo-filamentar structure has nothing in common with the structure which we have described in our parasite.

Wenyon, in his *Protozoology* (pp. 1050-51), assembles under the same title of *Cytamæba*, in the work of Dutton and alia, both the corpuscles which we have indicated above as others, associated with some rods of crystalline nature (Dutton and alia, Plate XXIV, Figs. 95-98), which, described by these authors as "an unidentified parasite", were included by Fran a in his genus *Toddia*, sp. *Toddia bufonis* (1910).

More recently Schwetz in Belgian Congo has described the *Bacillus krusei*<sup>2</sup> as "organismes intracellulaires, arrondis, tant t granuleux tant t presque homog nes et tant t ayant l'aspect d'un amas de minuscules bacilles".

They were found in *Rana albilabris*, *oxyrhynchus* and *occipitalis* and in *Bufo regularis*. The author did not find the amoeboid movements seen by other authors. He states that these parasites have been described by numerous authors under the names *Bacillus krusei* Laveran, *Cytamæba bacterifera* Labb  and *Toddia bufonis* Fran a.

Reviewing these parasites of frogs we believe

- (a) that the so-called *Bacillus krusei* Lav. of frogs is easily recognizable by its nodulo-filamentous contents and that *Cytamæba bacterifera* is a syn. of *B. krusei*,
- (b) that those parasites (?) which were associated with some crystals and were described by Dutton, Todd and Tobey as "an unidentified parasite", belong to so-called *Toddia* Fran a whose nature and systematic position remains yet doubtful,
- (c) that the so-called *Cytamæba* of Dutton, Todd and Tobey are of the same nature as the parasites we are studying now.

In 1915 we find a paper by Annie Porter<sup>3</sup> where some organisms figured by the author in her Text-Figs. 8, 9 and 10, belonging to the blood of cold-blooded vertebrates such as snakes and frogs, seem to us to belong to the same kind of organisms. Annie Porter identified them to *Anaplasma*, but we are very doubtful on such identification, as at that time much controversy reigned over the nature of *Anaplasma*, some authors as Dias and Araga<sup>84</sup> denying their protozoan nature. It seems that Porter included among those bodies various kinds of organisms and perhaps the mistake on the recognition of their differentiation came just because she considered that the structure was the same "whatever was the host from which they were obtained—man, cattle, mice, birds, reptiles, amphibia or fish." Our opinion is based on the fact that at the side of "small, uniform, usually spheric masses, that stained intensely with chromatic stain, that is, they were basophilic" were observed other corpuscles with a somewhat less densely staining portion of cytoplasm, surrounding the chromatine bodies. Indeed the author claims that "no marked exterior limit to such an area or halo could be found and the staining appeared to be merely an idiosyncrasy of that portion of the host cell." Whatever be the nature of those bodies—some of them, the so-called uniformly basophilic, perhaps *Anaplasma*, perhaps relicts of cell nuclei, or *anaplasmod*, the others with surrounding cytoplasm probably of the same rank as the bodies we are describing now—what is interesting to note in the description of Porter is the process of multiplication "near one point of the periphery of the nucleus of the red cell, a small bud appears. This tiny projection increases in size and gradually becomes somewhat spherical. The bud thus formed is extended and passes into the cytoplasm as a small spherical body." She described also a multiple budding, "the result being a small rosette of *Anaplasma* in the cytoplasm of the host cell."

It will not be devoid of interest to state the confusion yet existing on the nature and identification of mammalian *Anaplasma*-like bodies and the classification of Carpano<sup>8</sup> should here be recorded—

1<sup>st</sup> group—Karyolytic masses caused by nuclear delysis due to the introduction of chemical anemia producing substances or to toxins of microbes or metazoa—*Pseudoanaplasma*

2<sup>nd</sup> group—Stages on the evolution cycle of some *Piroplasma* (*Nuttalia*, *Theileria*, *Gonderia*, etc.)—*Anaplasmod*

3<sup>rd</sup> group—Microparasites constituting transitional forms between anaplasms and piroplasms and showing in experimental inoculations rare piroplasmoid forms—*Par-anaplasma* (sp. *Anaplasma centrale*)

4th group — Microparasites which may be transmitted in a pure stage—*Anaplasma* (sp *A. marginale*)

In 1924, Franchim<sup>4</sup> described in the blood of a bird some interesting bodies, which may give rise to some confusion with the parasites we are describing. In fresh blood they appeared as "de petits corps qui avaient plutôt l'aspect de vacuoles". In stained smears "les parasites les plus jeunes ont l'aspect d'une petite tâche colorée faiblement en bleu, ensuite cette tâche augmente de volume, se disposant en forme de virgule, mais plus souvent en forme de demi-cercle ou de cercle complet. Les parasites sont parfois isolés, mais plus fréquemment réunis à plusieurs dans le même globule rouge et à ce stade le protoplasme est uniforme. A un stade plus avancé, les corps en question augmentent de volume, ils prennent une forme ovalaire ou plus souvent arrondie en forme de bague. Leur protoplasme est plus foncé et il n'est pas rare de voir à son intérieur un noyau compact ou composé de grains de chromatine. Le contour du parasite est très apparent, il devient épais et chez le parasite adulte son épaisseur augmente encore. On pourrait croire qu'il s'agit de formes enkystées. Le noyau du parasite est toujours unique, il n'y a pas de nucléole et jamais nous n'avons pu voir des formes de multiplication. Le protoplasme ne contient pas de pigment. Il ne s'agit ni d'un Protoplasme ni d'un Anaplasme. Il ressemble à certains hématozoaires décrits antérieurement par Balfour".

Yes, such bodies were first described by Balfour<sup>5</sup> and his Plate VI is extremely clear. They were believed by Balfour to constitute the *after phase* stage of fowl spirochaetosis. This hypothesis is again referred to in the Fourth Report<sup>6</sup>. In 1929, Curasson and Andrijsky,<sup>7</sup> through some inoculation experiments, definitely state that the "Balfour Bodies" do not belong to the cycle of Spirochaetes and Brumpt, discussing this paper, says "Au cours d'une mission récente en Egypte j'ai eu l'occasion de discuter la nature des corps de Balfour avec le professeur M. Carpano. Cet Auteur a démontré la nature parasitaire de ces corps en les inoculant à des animaux sensibles, il a donné à ces corps qu'il considère comme des piroplasmidés le nom d'*Aegyptianella pullorum* . . . Les très intéressants germes découverts par Balfour qui au début de ses recherches, les considérait comme voisins des représentants de la famille des Piroplasmidés, doivent d'après Carpano et moi être classés dans cette famille, mon opinion est que ces êtres présentent surtout des affinités avec des parasites du genre *Anaplasma*".

These "after phase bodies", whose nature and structure is very different from that of the parasite which constitutes the object of this paper, have also been found in fowls by Bouet (1909) in French Soudan, by Dschounkowsky and Luhs in Transcaucasia, by Yowet (1910) in Cape Town, Donatien and Lestouquard (1890) in Alger and by Yakimoff in Caucasus. They are now classified as *Aegyptianella pullorum* Carpano (1929) (syn *Aegyptianella granulosa* Brumpt).

\* \* \*

A parasite entirely similar to ours has been described by Cerruti.<sup>10</sup> The coloured plate which illustrates his paper does entirely agree with our figures excepting his pyriform, cucumber-like organism depicted in Fig. 4. His description is also very suggestive in fresh blood "i corpuscoli si presentavano all interno dell' emazie come picoli corpi rotondi molto refringenti la luce, quasi immobili o dotati di movimenti di vibrazione piuttosto vivaci e di lenti movimenti di traslazione, per cui potevano spostarsi lentamente nell' interno della emazie". Stained by May-Grünwald-Giemsa "i granuli si coloravano in azzurro scuro piu o meno carico, teudente al bleu, la loro colorazione non era sempre uniforme, in fatti accentuau elementi colorati intensamente, ve ne erano altri che assumavano molto debolmente la sostanza colorante. Generalmente i granuli in questione si presentavano sotto forma rotonda, simili a cocci (tipo anaplasma) ma analogamente aquelli de *Testudo graeca* prendevano talvolta una forma ovale o nettamente bacillare (tipo *Bartonella Grahamella*), qualcuno assumeva la forma incurvata o reniforme od a rachetta. Nei preparati allestiti specialmente dalla nula, erano evidenti forme molto piu grandi, rotondeggianti o costituite da un anello periferico debilmente colorato in azzurro, limitante un vacuolo incolore e colorato molto debilmente. Internamente i corpuscoli apparivano ben limitati nel protoplasma della emazie, talvolta invece detto protoplasma formava intorno ad esse un alone vero e proprio".

Excepting for the *Bartonella* and racket-like forms every line of the description of Cerruti could be applied to our parasite.

Cerruti has classified this parasite in the genus *Grahamella* and named the sp. *G. Thalassachelys*.

The genus *Grahamella* was created by Brumpt in 1911<sup>11</sup> for the bacillary basophil parasites included in red cells of niles and first seen by Graham Smith<sup>12</sup> in 1905 "Leur aspect est bacilliforme. Certains éléments sont sphénques ou ovoides et mesurent un diamètre moyen d'un tiers de micron". Such bodies were also described and figured by Balfour<sup>13</sup> in two excellent micro-photographs where the red cells are seen filled up with rods

and very minute rods, an aspect familiar to every one having come across with *Grahamella*. In the Fourth Report, Balfour figures the appearance of these bodies stained by Romanowsky (his Plate VII, Fig 5) and if he is somewhat reticent about their nature, he agrees definitively with the opinion of Brumpt in 1911.<sup>14</sup> Brumpt continues "Ces corpuscules se colorent parfois d'une façon homogène intense par le Giemsa, le plus souvent ils ont leurs extrémités fortement colorées et le milieu est clair, c'est ce que l'on voit surtout dans les formes que je considère comme étant en voie de division (his Text-Figs I, E, G H)

Important to note is the process of division "une forme longue s'étrangle vers le milieu qui devient clair, tandis que les pôles se colorent intensément par suite de la condensation du protoplasma, l'étranglement s'accentue, la section s'effectue, il en résulte deux éléments arrondis ou ovalaires fort colorés qui s'allongent et se reproduisent de nouveau. Dans certains cas la division est inégale et constitue un bourgeonnement"

Brumpt defines the genus in the following way "Parasites arrondis ou bacilliformes, vivant dans les hématies des vertébrés, se reproduisant par division transversale et par bourgeonnement Sp typ *Grahamella talpæ*

The word *arrondis* does not apply certainly to such a large form as ours, but to the minute granules which fill the red blood-corpuscules of some mammals and giving a look to the illustrations inserted in page 1057 of Wenyon's *Protozoology*, we are doubtful whether the large anaplasmodoid bodies of his Figs. 1, 2, 13 and 31 will belong to the same type as the *Grahamella* of the other figures.

After this long discussion and a good personal knowledge of *Grahamella* such as they are found in the blood of some mammals, we come to the conclusion that our parasite and that of Cerruti cannot be included in the genus *Grahamella*.

We cannot identify our parasite to the genus *Pirrhæmocytion* (Chatton and Blanc, 1914), because whatever be the validity of the genus, the presence of the parasite in the red cell was associated with the appearance of a globular albuminous body in another part of the cell.

The genus *Cingula* (Awerinzew, 1914), on whose parasitic nature Wenyon has expressed his doubts, has some resemblance with our parasite. But this so-called parasite occurs as a small granule surrounded by a clear area and later on a vacuole appears, converting it into a ring on one side of which a nucleus could be seen. Division into two is then said to take place. Johnston described in 1917 similar bodies, in two snakes from West

Africa (*Echis carinatus* and *Causus rhombatus*) under two types one staining in homogeneous blue colour after Giemsa or Leishmann, while the other was granular and stained red<sup>11</sup> As the bodies we are describing resemble very much those found by Johnston, we will quote his own words "a small, spherical, hyaline, body, appearing in the red cells, taking a clear pale blue with Giemsa's or Leishmann's stain and occasionally showing a more darkly staining spherical centre Associated with this is usually another body of similar shape but often somewhat smaller—both bodies vary considerably in size—finely granular in appearance, taking a dull red stain."

In the parasite of Johnston it seems that the affected erythrocyte "as a rule stains somewhat poorly, it may contain vacuoles, one or more. The nucleus is often smaller and more rounded than that of a normal cell, it may be apparently degenerating, staining poorly and showing vacuolation or it may even be absent."

The author is very reticent about the nature of these bodies whether they are parasites "though it may be considered probable. They may perhaps be a new species or perhaps some stage in the development of a parasite already described, such as a hemogregarina"

Carini in 1930<sup>12</sup> described in the *Leptodactylus pentadactylus* from São Paulo, Brazil, some bodies which are very similar to our and Cerruti's parasite "à l'examen direct... petites vacuoles qui, étant plus refrigentes et incolores, se détachent dans le protoplasme de l'hématie... se colorent en bleu par le Giemsa, parfois il n'y a qu'un petit anneau périphérique qui se colore en limitant une vacuole, mais dans les préparations mieux réussies on voit un granule plus intensément coloré"

After vital staining "les corpscules se colorent en quelques minutes et apparaissent avec une grande netteté dans les globules presque incolores. Généralement ronds ou ovalaires, leurs dimensions sont variables, les plus petits sont de la grosseur d'un coccus tandis que les plus gros ont un diamètre de 1-2 microns. Dans les plus gros, on distingue une partie colorée en rose qui contient un granule bleu foncé autour des corpscules on voit parfois dans le protoplasma de l'hématie une zone circulaire plus pâle. Les hématies parasitées ne sont pas altérées"

Carini finds that they have affinities with *Anaplasma*, *Aegyptianella*, *Grahamella*, *Bartonella*, *Eperythrozoon*, but cannot be included in any of these genera and proposes to this parasite the name *Bertarellia leptodactylis*

Brumpt and Lavier have described under the name *Pirrhämacylon* a parasite of *Lacerta viridis* which seems to us very similar to ours

The authors give firstly a short description of *P. tarentolæ* apud Chatton and Blanc (a) young spheric elements, 1-4 micr., clear amoeboid outline, possessing in their interior one or many chromophyl masses "Celle-ci affectent des formes très variées; elles sont soit entières, soit bi, tri, quadrilobées, ou bien ces lobes sont séparés, formant autant de masses distinctes de tailles diverses" In some of these parasites it may be seen that "même lorsqu'ils ne sont pas au contact du noyau de l'hématie, leur masse chromatique se trouve unie à ce dernier par un tractus filiforme très colorable qui se termine soit en pointe, soit plus souvent par une capitulation très nette", (b) spheric masses, 3,5 micr., without surrounding cytoplasmic area, "leur chromatine est divisée en fins granules, rangés à la périphérie, sauf un corpuscule qui reste central" Important to note, as we have said above, is that in all parasitized erythrocytes there is "une inclusion globoïde, mesurant 1-8 micr., indépendante d'ailleurs structuralement du parasite, mais liée à sa présence, réfringente à frais, homogène et cyanophile et représentant une réaction de l'erythrocyte parasité"

Brumpt and Lavier do not find this body in the red cells of *L. viridis* and notwithstanding they classify their parasite as *P. lacerta* whose following stages are described by the authors (a) anaplasmod, homogeneous body, often surrounded by a light cytoplasmic halo, which sometimes at the contact with the cell protoplasm takes a bluish stain Forms where the cytoplasm has the appearance of fine lilac granulations around a central chromatic dot are not rare Diamètre up to 4 micr., (b) spheroidal masses 2, 4, 5 micr., outline rather irregular, showing amoeboid movements, chromatic mass single, often multilobated, or composed of 2, 3, 4 independent dots Sometimes a chromatic filament seems to start from the nuclear mass and project to the periphery, constituting in small forms the leishmanoid aspect already recorded by Chatton and Blanc, spherical masses of 5 or more micr. "avec plusieurs petits grains chromatiques massés dans un seul hémisphère. nous n'avons jamais observé de granule occupant le centre et il nous semble avoir en affaire seulement à un nombre restreint (6 en général) de granulations chromatiques, non sphériques mais légèrement allongées, de tailles souvent inégales, et présentant une disposition assez régulière"

The parasite was successfully inoculated to other lizards of the same species. The authors say "fait important, si Chatton et Blanc ont signalé, comme nous l'avons dit, la présence constante d'une inclusion globoïde dans chaque globule parasité, chez notre lézard, les erythrocytes ne montraient pas la moindre réaction ni de ce genre, ni d'aucun autre Néanmoins notre

parasite sans être identique à celui de la tarante, est certainement extrêmement voisin, aussi proposons nous de le nommer *Pirrhämocyon tarentola* n sp

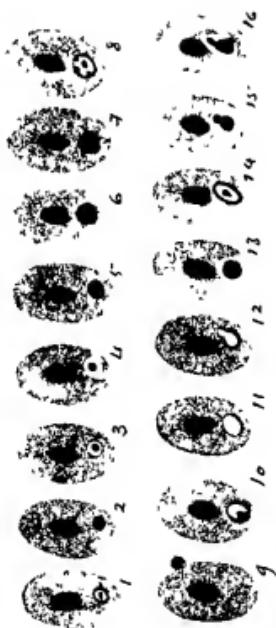
We don't agree with Brumpt and Lavier in this identification. Their parasite is not a *Pirrhämocyon* and seems of the same nature as the parasite we are describing in this paper, for which we maintain the genus *Bertarellia* Carini, (1930), classifying it as a sp n which we will name *Bertarellia calotis*

*N.B.* — Prof Carini in a private letter, emphasizing that the genus *Bertarellia* belongs to the ill-defined group of *Bartonella*, *Pirrhämocyon*, *Grahamella*, *Aegyptianella*, *Tuncella* etc., and pointing out that the parasitic nature of *Bertarellia* is probable, but not demonstrated, has promised to send us slides of Brazilian tortoise blood containing bodies similar to those described by Cerruti<sup>10</sup>. As we have also found *Bertarellia* in the blood of the Indian tortoise *Emyda granosa*, the definite characteristics of all these genera, together with the description of the tortoise *Bertarellia*, will be the subject of a further paper.

#### BIBLIOGRAPHY

- 1 Dutton J Everett Todd John L, and Tobey I. N, 'Concerning certain parasitic protozoa observed in Africa' *Ann of Trop Med and Parasit* 1907 1 No 3
- 2 Schwetz, J, 'Notes protozoologiques. Les hématozoaires des grenouilles et des crapauds de Stanleyville (Congo Belge)' *Ann de Parasit Humaine et Comparée*, 1930 8, No 2
- 3 Porter, Annie, 'On Anaplasma like bodies in the blood of vertebrates,' *Ann of Trop Med and Parasit* 1915 9 No 4
- 4 Dias, Ezequiel Caetano, and Aragão, Henrique de Beaurepaire, 'Pesquisas sobre a natureza dos anaplasmas,' (text also in German) *Memórias do Inst Oswaldo Cruz*, 1914, Tom VI, Fasc. III
- 5 Carpano, M, 'Sur la nature des anaplasmes et en particulier de l'Anaplasma centrale,' *Ann de Parasit Humaine et Comparée* 1930, 8 No 2
- 6 Franchini, G, 'Hematozoaires particuliers d'un oiseau (Hypoleis hypoleis),' *Bull Soc Path Exot*, 1929, 22, No 5
- 7 Balfour A, 'Sprochocotosis of Sudanese Owls,' *Third Report Wellcome Res Lab* 1908
- 8 Balfour, A, 'Sprochocotosis of Sudanese Owls,' *Fourth Report Wellcome Res Lab* 1911, A
- 9 Curasson, M G, et Andrijesky, P, 'Sur les corps de Balfour du sang de la poule,' *Bull Soc Path Exot*, 1929 22, No 5
- 10 Cerruti, Carl G, "Su di una Grahamella parassita di *Talassochelys caretta* (*Grahamella talassochelys* n sp)," *Archivio italiano di Scienze mediche Coloniali* 1931 12, \* Fasc 6
- 11 Brumpt, E, 'Note sur les parasites des hématies de la taupe *Grahamella talpa* n. sp.,' *Bull Soc Path Exot* 1911, 4, No 8

- 12 Graham Smith, G. S. "A new form of parasite found in the red blood corpuscles of moles," *J. of Hyg.*, 1905, 5, No 4
- 13 Balfour, A. "Changes in the erythrocytes of gerbou," *Second Report Wellcome Res. Lab.*, 1906
- 14 Balfour, A. "A note on the new genus *Grahamella* Brumpt," *Bull. Soc. Path. Exot.*, 1911, 4, No 10
- 15 Johnston, J. E. L. "On some peculiar bodies found in the blood of certain African snakes," *Journ. of Trop. Med. and Hyg.*, 1917, 20, No 21
- 16 Carini, A. "Présence de corpuscules de nature parasitaire probable (*Beriorellia leptodactyli*) dans les hématies de *Leptodactylus pentadactylus*," *Compt. Rend. Soc. Biologie*, 1930, CIII, 1312-13
- 17 Brumpt, E. and Lavier, G. "Sur un hématozoaire nouveau du lézard vert *Purhescocystis lacertæ* n. sp." *Ann. de Parasit. Humaine et Comparée*, 1935, 13, No 6





## STUDIES IN SOIL BACTERIA OF THE SUBTROPICAL REGION—PUNJAB, NORTH INDIA.

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Received March 29, 1937

(Communicated by Dr S L Ghose, M Sc, Ph D)

### 1 Introduction

A good deal of quantitative work has been done in other countries as regards the seasonal or daily variations in bacterial numbers in order to correlate such variations with soil fertility and physical factors such as temperature, moisture and rainfall, etc. In India, however, as far as it could be traced, no such work has so far been carried out and it appeared highly desirable to investigate the type of fluctuations occurring in our field soils during different seasons of the year—seasons which are so particularly marked in the Punjab. In the Punjab, seasons of the year can be divided into four distinct types. (1) Hot summer from middle of May to middle of September. The temperature during this period goes sometimes as high as 120° F. (2) Autumn months from middle of September to middle of November. (3) Cold winter extends from middle of November up till the middle of March. The temperature during the cold months goes as low as 32° F or less. (4) From the middle of March to the middle of May is the spring weather. Keeping in view the marked changes in seasons in the Punjab, the present work was undertaken to investigate the following points: (a) What is the state of the bacterial population in the Punjab soils with particular reference to its quantitative aspect? (b) What are the changes that are taking place in the bacterial population in our field soils during the different seasons of the year?

### 2 Previous Work

Hiltner and Störmer (1903) found little differences between bacterial numbers in summer and winter. Remy (1902) and Wojtkiewicz (1914) have reported highest numbers of bacteria in spring. Given and Willis (1912), on the other hand, found highest numbers in September. Brown and Halverson (1919) found two maxima in February and in June. Wilson (1930) working on variations in *Rhizobium* sp., found a drop as winter advanced. In spring the numbers increased till June. He found two maxima in October and June and one minimum in January. Snow (1926-35)

found while studying the micro-organisms of wind-blown soils, two maxima in January and July and one minimum in February. Cutler, Crump and Sandon (1922) have reported one maximum in November, a minimum in February and a second maximum at the end of June. Their curves did not correlate with seasonal changes in soil temperature and moisture. Löhnis and Sabaschnikoff (1912), Müntz and Gaudechon (1912), Wojtkiewicz (1914), Russell and Appleyard (1917), Lemmermann and Wickers (1920) have all reported greater stimulation of chemical processes in the soil either in spring or autumn or both.

As regards the relation of bacterial numbers with physical factors Waksmann (1916 & 1932), Dixon (1928), Thornton and Gray (1930), Newton (1930) found no correlations with either the soil moisture or temperature. Conn (1910), Russell (1913), Harder (1916), Lockhead (1923), Mary-Jo-Cobb (1932) have, however, pointed out that bacterial numbers and soil moisture are closely related to each other.

### 3 Experimental Work and Technique

Two plots were selected in the Government College Botanical Gardens for experimental work. One of these plots was manured with farmyard manure for a number of years and was thus named as manured plot. In the other plot no manure was put and was called as an unmanured plot. This prepared us to see seasonal changes in both the kinds of soils. The methods used for sampling, suspension, disintegration, dilution, plating, incubation and enumeration, etc., are those suggested in appendix, Russell (1932).

### 4 Results

Date	Plot	Moisture %	Average Colony count of five plates	Standard error
14th December 1935	Manured	17.88	88.8	± 2.9
	Unmanured	15.00	51.0	± 1.58
25th December 1935	Manured	20.48	54.8	± 2.80
	Unmanured	18.00	29.6	± 2.37
11th January 1936	Manured	18.56	87.2	± 4.15
	Unmanured	16.38	74.0	± 3.60
35th January 1936	Manured	18.10	51.6	± 2.65
	Unmanured	15.66	29.2	± 2.00

## Results—(Contd.)

Date	Plot	Moisture %	Average Colony count of five plates	Standard error
8th February 1936	Manured Unmanured	21.96 21.548	82.4 60.6	± 2.84 ± 2.46
23rd February 1936	Manured Unmanured	19.46 18.12	64.2 64.4	± 5.00 ± 4.30
7th March 1936	Manured Unmanured	16.68 15.80	58.2 41.2	± 2.00 ± 3.30
21st March 1936	Manured Unmanured	21.28 20.50	85.1 95.2	± 4.00 ± 5.70
6th April 1936	Manured Unmanured	23.04 21.14	127.0 111.6	± 8.00 ± 7.01
18th April 1936	Manured Unmanured	15.80 15.38	126.8 94.75	± 5.10 ± 3.80
4th May 1936	Manured Unmanured	18.92 16.04	222.20 199.40	± 12.70 ± 5.00
5th May 1936	Manured Unmanured	13.81 12.64	198.20 159.00	± 7.00 ± 4.10
30th May 1936	Manured Unmanured	8.20 5.00	151.40 117.00	± 4.90 ± 5.60
15th June 1936	Manured Unmanured	16.52 15.06	133.80 128.00	± 2.40 ± 3.50
27th June 1936	Manured Unmanured	17.40 14.16	153.80 131.60	± 4.50 ± 7.00
11th July 1936	Manured Unmanured	19.52 17.12	119.20 102.00	± 6.9 ± 6.3
26th July 1936	Manured Unmanured	13.24 11.16	104.20 96.20	± 6.3 ± 8.2
8th August 1936	Manured Unmanured	10.32 8.40	69.40 58.00	± 2.7 ± 4.1
26th August 1936	Manured Unmanured	22.00 17.76	93.60 71.80	± 2.5 ± 4.2

## Results—(Contd.)

Date	Plot	Moisture %	Average Colony count of five plates	Standard error
5th September 1936	Manured	15.50	81.20	± 3.2
	Unmanured	14.60	72.60	± 2.9
19th September 1936	Manured	17.64	105.80	± 5.4
	Unmanured	16.62	82.50	± 3.0
3rd October 1936	Manured	18.80	115.00	± 7.3
	Unmanured	17.54	100.68	± 4.1
17th October 1936	Manured	17.36	216.60	± 11.1
	Unmanured	16.92	186.30	± 4.3
1st November 1936	Manured	22.21	87.60	± 4.36
	Unmanured	20.20	97.00	± 6.50
14th November 1936	Manured	22.72	79.80	± 2.70
	Unmanured	19.04	83.30	± 2.10
28th November 1936	Manured	16.20	115.00	± 2.17
	Unmanured	15.80	102.30	± 4.60

## 5. Discussion

It has been pointed out that the main object of this work was to see the effect of various seasons and other physical factors upon the bacterial numbers in our field soils. It is desirable that they should be discussed separately since these factors play such an important part and their intensities are so very well marked.

*Seasonal effect*—The graph, Fig. 1, of the bacterial numbers of the manured and unmanured soils, show that there are two maxima, one sometimes in October and the other towards the end of April or beginning of May, and there are two minima in the end of January and in the first week of August. Our results have hence shown that there is a marked periodicity of the bacterial population in the soil—a rise of numbers in spring and autumn, and a distinct fall in summer and winter. These results are in general concordance with those of Russell and Appleyard (1917), Brown and Halverson (1919), Cutler and his associates (1922), Wilson (1930) and others, while on the other hand, Hiltner and Störmer (1908) found little differences between bacterial numbers in summer and winter. It may

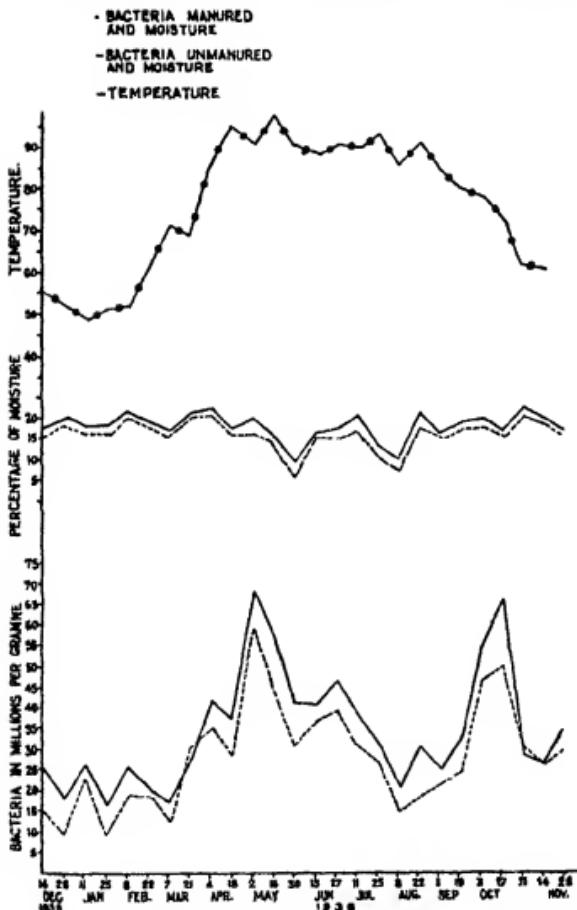


FIG. 1. Fortnightly Averages of Bacteria, Moisture and Temperature  
Manured and Unmanured Plots

further be pointed out that our maxima and minima occur at different times of the year and this one might have been expected since our conditions are different from those of all the other workers

*Influence of soil moisture*—When curves, Fig. 1, for average moisture contents and average numbers are compared in greater detail, it becomes impossible to draw any generalization. Thus in winter the numbers have been low but the moisture contents were fairly high, while in summer the moisture had naturally decreased and there was as well a decrease in numbers, but there does not seem to be an apparent correlation since the two decreases are not simultaneous or nearly so. In the beginning of June, for example, the moisture contents are at its minimum but the numbers are fairly high and the reverse is the case towards the middle of August—numbers very low but moisture contents fairly high. In general it can be stated that there is no definite relation of the bacterial numbers with the moisture present in the soil. Our results hence accord with the results of Remy (1902) and Wojtkiewicz (1914), Given and Willis (1911-12), Waksman (1916), Brown and Halverson (1919), Cutler, Crump and Sandon (1922), Thornton and Gray (1930), Newton (1930).

Hiltner and Stormer (1903), Conn (1910), Russell (1913), Harder (1916), Lockhead (1924), have found a correlation of the numbers with the moisture contents

*Influence of rainfall*—It has been suggested by Russell and some other workers, that a heavy shower of rain by increasing the aeration of the soil population may have beneficial effects upon the micro-organisms. In our case, however, rainfall cannot be said to have marked effect, on the changes in numbers since they occurred even when there was no rainfall. Thus the greater part of autumn and spring rise in numbers occurred when no rain fell for a considerable time preceding these months.

*Influence of temperature*—That the temperature has effect on the bacterial numbers is apparent from the graph, Fig 1. With the rise of temperature from winter months the bacterial numbers also show a corresponding rise which seem to continue with certain optimum temperature limits. Then with persistent high temperature the numbers show a decrease. Again with the lowering of the temperature and probably with the partial removal of the bad effects of high temperature to the bacterial numbers, there is a tendency of recovery. It may be stated, however, that the effect of temperature on the bacterial population is accumulative in nature. A persistent low temperature decreases the numbers as in winter, then a rise in temperature has a wholesome effect as in spring. A continuous high

temperature has a tendency of keeping the numbers down as in summer, and lastly the numbers again rise up with slight lowering of the temperature as in autumn.

Hiltner and Stormer (1903), Cutler, Crump and Sandon (1922), Dixon (1928) did not find any correlation between the bacterial numbers and the temperature, but the finding of Given and Wilhs (1911-12), Brown and Halverson (1919) definitely support this.

*Influence of chemical changes in the soil*—No special chemicals were tried but from the two kinds of plots it is definite that one which contained more organic matter has more organisms both in numbers and types.

In the end it may be pointed out that the maximum bacterial activity in our parts is manifested from the middle of April to the middle of May in spring, and from the middle of September to the middle of October in autumn. One may as well reasonably assume that the availability of other plant nutrients such as nitrates, ammonium-salts, potassium, phosphates, etc., may also be at their maximum somewhere in the months noted above. On reviewing the crop sowing operations in the Punjab, it is noted that wheat is grown sometimes in October and is harvested in April or May. Sowing of cotton is done in May or so along with some other leguminous crops, and harvesting is effected in September or October. These are the two most important crops in the Punjab, of course, so many other types of crops are grown as well—sugarcane, vegetables, etc. From the above farm operations it is quite apparent that the months of maximum activity—April-May and September-October—are not utilised as they should be, in the process of the growth of crops. During the above-noted months of great biological activity, the fields are almost without any crops, and it may be suggested that farm operations may be regulated in such a way that the high available plant food materials in spring and autumn in the above-noted months should be utilised by the growing crops in some form or another.

*Qualitative*—A few words may be said about the floristic characteristics with regard to the soil bacteria. No attempt was made to make a detailed study of the different types of bacteria occurring in our soils. But during the course of the work attention was kept on a spreading type of a *Bacterium*, since this form very easily overran some of the plates. It was noticed that this form was hardly visible on the plates in winter months but with the rise in temperature in March and April it could very easily be seen. Again in very hot summer months no trace of the type could be found and it again became apparent in the beginning of autumn months.

It is quite probable that this spreading type may have some relation with temperature or with some other external factors, and it is not unlikely that some other less conspicuous forms in the soil may be appearing and disappearing with different seasons of the year

#### 6 Summary and Conclusions

I. In this paper the seasonal variations for bacterial numbers have been described in manured and unmanured field soils in the Punjab

II. Two maxima in the middle of October and beginning of May, and two minima in the end of January and in the first week of August were obtained

III. The variations of bacterial numbers in the soil cannot be explained as entirely to be due to any of the physical factors individually, but may be the result of the accumulative effect of all the factors of which the temperature seems to be the most important

IV. It may be stated that in our soils the number of bacteria as occurring on the plate method is higher than those reported by the European and American workers

V. Different types of bacteria seem to be appearing and disappearing at different seasons of the year as indicated by a spreading type of *Bacterium*, but an exclusive study of this problem is desired before any definite statement could be made

The author wishes to express his thanks to Mr H Chand who helped during the course of the work in doing plating sometimes

#### LITERATURE CITED

1. Brown, P. E., and Helverson, W. V. *Iowa Agr. Exp. Sta. Res. Bull.*, 1919, 56, 251-58
2. Conn, H. J. *Centbl Bact.*, 1910, 2, 28
3. Cutler, D. W., Crump, L. M., and Sandon, H. *Phil Trans Roy Soc, Series B*, 1922, 211, 317-50
4. Dixon, D. *Aust Jour Exp Bio. & Med. Sci.*, 1928, 5, 223-32.
5. Given and Willis. *Penn Agr. Exp. Sta. Ann. Report*, 1911-12, 441-54
6. Harder, E. C. *Bot. Gaz.*, 1916, 61, 507-17
7. Hiltner and Stormer. *Arb K. Gesundheitsamt, Biol. Abt.*, 1903, Heft 5, 445
8. Lemmermann, O., and Wickers, J. L. *Centr. Bakt.*, 1920, 50, Abt II, 33
9. Lockhead, A. C. *Trans Roy Soc Canada*, 1923, 18 (V), 382-423
10. Lohnis, F., and Sabaschnikoff, A. *Centr. Bakt.*, 1908, 20, Abt II, 322
11. Mary-Jo-Cobb. *Sod Sci.*, 1932, 33, 325.

12 Muntz, A., and  
Gaudichon, H. *Comptes Rend Akad Sci.*, 1912, 154, 163

13 Newton, J. D. *Soil Agr. (Ottawa)*, 1930, 10, 361-66

14 Remy, T. *Centr Bakt. B. Abt II*, 761

15 Russell, E. J. *Jour Agr Sci.*, 1913, 5

16 Russell, E. J., and  
Appleyard *Ibid.*, 7.

17 Snow, M. Laetitia *Soil Sci.*, 1926, 21, 143-65, 1927, 24, 34-39, 1927, 24  
341-55, 1935, 40, 181-89

18. Thornton, H. G., and  
Gray, P. H. H. *Proc Roy Soc, Seris B*, 1930, 106, 399

19. Wakemann, S. A. *Soil Sci.*, 1916, 1, 363-80

20. *Principles of Soil Microbiology*, 1932

21. Wilson, G. K. *Soil Sci.*, 1930, No 5, 289

22 Wojtkiewicz, A. *Centr Bakt.*, 1914, 42, Abt 11, 254



# SOME STUDIES ON TEMPERATURE OF THE COTTON PLANT IN THE PUNJAB.

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Received June 10, 1937

(Communicated by Mr B. N. Sastri, M.Sc., A.I.C., A.I.E.E.)

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## I Introduction

IT is well known that temperature as an environmental factor has a marked influence on the various life processes of a plant, *i.e.*, respiration, transpiration, root absorption, etc. According to Vant Hoff's rule (1896) "The rate of chemical reaction doubles for a rise in temperature of  $10^{\circ}\text{C}$ ." Its effect on economic crops is particularly a matter for instructive and useful study in the Punjab, where the air temperature rises sometimes to  $118^{\circ}\text{F}$ . in summer and falls below zero in winter. American cotton, to which the investigation reported in the paper relates, is cultivated almost exclusively in the canal colonies, where the plant has to adapt itself to extremes as well as to fluctuations of temperature and humidity during its growing period, which extends from April to December. The measurement of temperature of the cotton plant was undertaken to ascertain the extent to which the plant adjusts itself to atmospheric heat under such trying

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circumstances. There have been successes and failures of American cottons in this Province and it seems likely that unfavourable years were chiefly the results of adverse climate conditions and some soil factors particularly moisture contents.

References to earlier work on plant temperatures as far as the writers have been able to obtain are the following —

Clun (1926) while taking the temperature of leaves of *Fuchsia speciosa*, *Phaseolus vulgaris*, *Brassica oleracea* and *Syringa vulgaris* observed that, in general, plants in dry soil, and vaselined leaves were 2° to 4° C warmer than the controls. Balls (1912) working on cotton leaves in Egypt reports that the temperature of old leaves rarely fell below air temperature, but frequently rose above the air temperature by 3° C to 10° C. Eaton and Belden (1929) in U.S.A. report that the leaf temperature of cotton plants are correlated with transpiration, varietal differences and yields. The yields of the upland varieties were distinctly higher in 1923 than in the hotter years 1922 and 1924. Miller and Saunders (1923) while studying the temperature-transpiration relation of the leaves of corn, sorghum, cowpeas, soybeans and watermelons growing under field conditions, found that the temperature of leaves never remained constant. They also found that the temperature of the wilted leaves of corn, sorghum, etc., remained 1.85°, 1.55°, 2.8° and 4.65° C higher than the temperature of turgid leaves. Ezekiel and Taubenhaus (1932) found that the leaves of cotton plants, wilted from *Phymatotrichum* root-rot were as much as 0.5° F warmer than the air. Seeley (1917) in Michigan found the temperature of strawberry leaves on clear days to be 8.4° C above air temperature. Trought (1931) states "The temperature of the leaves follows closely the temperature of the surrounding air. For cotton, unpublished figures obtained by R. S. Jai Chand Luthra confirm this fact."

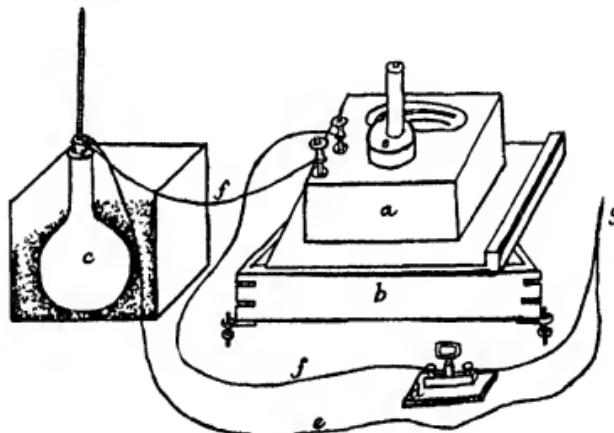
#### II Method of work and apparatus employed

Determinations of temperature have been made on 4F cotton plant—a Punjab acclimatised upland American cotton (*G. hirsutum*). The work was started in 1928 and has been in progress since then. For this purpose, leaves of the same size, and of comparable age 30–35 days old and those hanging at right angles to sun on the secondary branches and at the height of about 2 feet were always selected on a single plant. Temperatures of the upper and lower surfaces of the leaf and the internal tissue of the midrib and the petiole were measured. As the temperature of the leaf was found to fluctuate rapidly, six determinations were taken each on a different leaf and the average value was recorded. The standard error for these sets of six figures ranged from .1 to .6 and is statistically sound.

Variations of temperature are associated with a number of factors. The following are some of the factors studied —

- (A) The surrounding air
- (B) Soil moisture
- (C) Moisture content of leaves
- (D) Age of leaves
- (E) Wilting of leaves

*Apparatus* — The apparatus (Fig. 0) used for the determination of temperature was devised on the principle of thermo-electricity by modification of the one used by some American workers. It consists of two thermo-junctions, a galvanometer and a spring key. The thermo-junctions were



REFERENCES

FIG. 0. *a* Galvanometer, *b* Levelling Tripod stand, *c* Thermos flask, *d* Key, *e* Constantan wire, *f* Copper wire, *g* Free junction

constructed by soldering the ends of a constantan wire No. 32 to one end of each of two copper wires No. 36. The absorption coefficient of the wires was nil. The free end of one copper wire was connected with key and that of the other to a galvanometer, thus forming a complete circuit. The wires were insulated. One of the thermo-junctions was inserted into the neck of a stoppered Dewar's flask along with a thermometer. Hot water was put into the flask in winter and cold water in summer. The bulb of the

thermometer was dipped into the water. The Dewar's flask was placed in a box filled with sawdust. The temperature of the water in the Dewar's flask was constant. The second thermo-junction was held in a clamp in such a way that it would be conveniently placed upon the leaf surface. The clamp consisted of a pair of brass tongs to the ends of which pieces of cork were glued. For determining the temperature of the inner tissue the free thermo-junction was inserted into the petiole or midrib. The temperature of the atmosphere was taken by holding the free junction in the air and placing it in a position where air played freely on it at a height of 2 feet from ground level. Necessary precautions were taken to screen the thermo-junctions from direct sun rays. A pivot galvanometer No L 27156 manufactured by the Cambridge Instrument Company, Ltd., England, sensitive enough to indicate at least 0.0025 micro-amperes was placed in a circuit along with a spring key. Since the temperature of the thermo-junction in Dewar's flask was known and constant, the difference in temperatures between the two junctions could be shown by a swing of the indicator of the galvanometer. The galvanometer was placed on a levelling tripod, which could be adjusted to get a level surface for the galvanometer. The whole apparatus was kept in a cabinet to protect it from the sun (Plate XIII).

*III (A) Relationship between the leaf temperature and the surrounding air*

Work was started in 1928 and has been continued upto 1934. The data of 1931 are reported herein. The work was repeated during the years 1932, 1933 and 1934, and the results obtained when compared with the data of 1931 showed minor differences, which are accounted for by differences in relative humidity, soil moisture and variation in leaf moisture content resulting from different amounts of rain received during the growth period of the plant. However, the general trend of results is the same and there is a corroboration of the results of 1931. Seed of a pure strain of 4F cotton was sown in May each year in a typical field of the botanical area at Lyallpur. Representative plants of the crop were marked for temperature determinations. For the first three years and for June and July of 1931 leaf temperatures were taken thrice daily at 7 A.M., 1 P.M. and 7 P.M. respectively. The data collected during three years' work was studied and it was considered that three observations during the day at an interval of six hours did not furnish sufficient data to study fully the range of variability of leaf temperatures from that of the air. Consequently from August 1931 onwards the work was extended and determinations were made every two hours throughout day and night during August and September and for

the day only during October and November. The results obtained during different months are as follows —

*June and July 1931* — Leaf temperatures were taken thrice daily and the results obtained are given in Table I. In the morning and evening plants show air temperature. During the middle of the day the temperature of the plant is lower by about two degrees and the variability is found to be greater than at the other hours.

TABLE I

*Monthly mean departures of leaf temperatures (columns 4-6) from air temperatures (column 3) during June and July 1931*

*"Irrigated crop"*

Month	Time	Air tempera- ture in degrees C	Upper surface	Lower surface	Inner tissue	
					Petiole	Midrib
1	2	3	4	5	6	
June	7 A.M.	28.7	- 2	- 5	- 1	- 2
	1 P.M.	37.5	- 2.0	- 3.0	- 2.3	- 3.1
	7 P.M.	32.0	+ 0.2	+ 0.2	+ 0.0	- 1
July	7 A.M.	28.4	+ 0.5	+ 0.4	+ 0.4	- 0.3
	1 P.M.	40.0	- 1.8	- 2.8	- 1.7	- 2.9
	7 P.M.	34.6	+ 0.3	- 1	- 4	- 2

*Note* — It is realized that full information can only be got from daily variations, but as it is not feasible to record such cumbersome data, monthly means are given. To supplement this information, data of highest and lowest departure are also noted in the remarks column where necessary.

*July 1932* — In July 1932 readings were taken every three hours during the day, and the data are given in Table II, Fig. 1. It will be seen that leaves have almost the same temperature as the air at 6 A.M., but then they gradually begin to be warmer and at 9 A.M. there is a rise of about 1° to 2.5° C above the air temperature. Afterwards there is a slow fall and the leaves remain cooler than the air by about 3° to 4° C till evening, when they again acquire the temperature of the air.

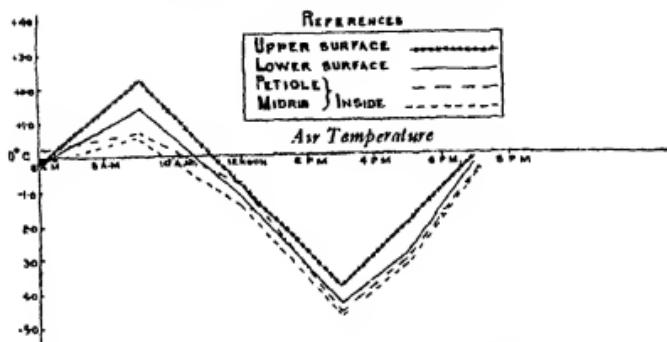


FIG. 1 Monthly mean departures of leaf temperatures of irrigated 4F Cotton Plants from air during July at different periods of the day.

TABLE II

*Monthly mean departures of leaf temperatures (columns 3-5)  
from air (column 2) during July 1932 (Irrigated crop)*

*"Bud formation" Stage*

Time	Air tempera- ture in degrees C°	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
6 A.M.	29.1	— 2	— 1	+ 1	— 2
9 A.M.	33.8	+ 2.2	+ 1.3	+ 0.5	+ 4
12 Noon	38.6	— 9	— 1.2	— 0.9	— 1.3
3 P.M.	40.5	+ 3.0	+ 4.3	+ 6.6	+ 4.8
5 P.M.	38.2	+ 1.0	+ 2.0	+ 3.0	+ 2.1
7 P.M.	33.7	— 1	— 0.2	— 0.5	— 0.5

## II

Comparison of mean departures of leaf temperatures  
from air on July 15th, 1932

## " Irrigated crop "

Hours of observa- tions	Mean Temperature of air in °C	Mean Temperature of Upper Surface in °C	Mean Temperature of Lower Surface in °C	Inner leaves		Remarks
				Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
6 A.M.	30.0 ± 20 Difference from air temperature	29.6 ± 15 — 4 ± 25 Insignificant	29.7 ± 11 — 3 ± 22 Insignificant	29.8 ± 10 — 2 ± 22 Insignificant	29.7 ± 10 3 ± 22 Insignificant	
9 A.M.	34.4 ± 20 Difference from air temperature	37.0 ± 10 12.0 ± 15 Significant	36.1 ± 10 11.7 ± 15 Significant	35.2 ± 10 — 4 ± 20 Significant	35.3 ± 10 + 7 ± 20 Just significant	In the morning and even- ing leaves show air tem- pera- ture but they are warmer at 9 A.M. and signifi- cantly cooler than the air in the afternoon
12 Noon	39.0 ± 20 Difference from air temperature	37.8 ± 15 — 1.2 ± 25 Significant	37.5 ± 18 — 1.5 ± 28 Significant	37.7 ± 15 — 1.3 ± 25 Significant	37.4 ± 15 — 1.0 ± 25 Significant	
3 P.M.	41.2 ± 45 Difference from air temperature	37.0 ± 40 — 4.2 ± 60 Significant	36.8 ± 30 — 4.4 ± 54 Significant	36.7 ± 40 — 4.5 ± 6 Significant	36.8 ± 35 — 4.4 ± 57 Significant	
5 P.M.	38.0 ± 40 Difference from air temperature	36.2 ± 30 — 2.4 ± 50 Significant	36.0 ± 15 — 1.8 ± 45 Significant	35.2 ± 40 — 3.4 ± 50 Significant	35.1 ± 40 — 3.5 ± 55 Significant	
7 P.M.	34.4 ± 30 Difference from air temperature	34.0 ± 20 — 4 ± 36 Insignificant	33.9 ± 16 5 ± 13 Insignificant	33.8 ± 15 — 6 ± 33 Insignificant	33.7 ± 12 — 7 ± 32 Insignificant	

August 1931.—The data obtained are summarised in Table III and graphically represented in Fig 1(a). For convenience and brevity, cotton temperatures represented in the tables and figures are given as departures from the air temperature which is shown as a straight line in the figures and is taken as zero. It is found that the temperature of the leaves is never constant and keeps on fluctuating with that of the air. In the morning at 6 A.M. the leaf temperature almost coincides with the air. The lower surface and the inside of the midrib are, however, cooler by about

1° C. Afterwards it begins to rise gradually and at 8 A.M. it is about 5° to 6° C above the air in the case of upper and lower surfaces and 3° to 4° C

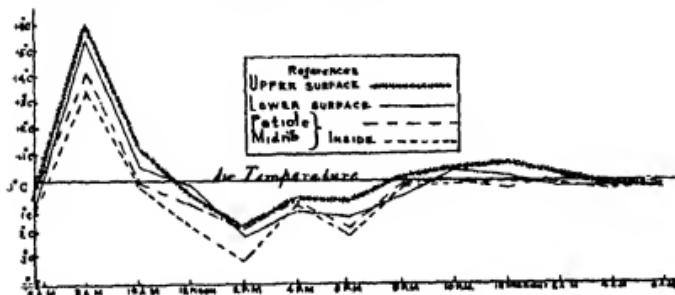


FIG. 1(a) Monthly mean departures of leaf temperatures of irrigated 4F Cotton Plants from air during August at different periods of the day

TABLE III  
*Monthly mean departures of leaf temperatures (columns 3-5)  
 from air (column 2) during August 1931 (Irrigated crop)*  
*"Bud formation—Stray flowers" Stage*

Time	Air tem. perature in degrees C	Upper surface	Lower surface	Inner tissue		Remarks
				Petiole	Midrib	
1	2	3	4	5		6
6 A.M.	27.71	- .21	- 1.11	- .38	- 1.39	
8 A.M.	28.90	+ 6.07	+ 5.37	+ 4.10	+ 3.59	
10 A.M.	30.56	+ 1.24	+ .60	- .13	- .20	
12 Noon	38.7	- 0	- 2.22	- .92	- 1.8	
2 P.M.	35.22	- 1.79	- 2.19	- 1.88	- 3.11	
4 P.M.	34.05	- .65	- 1.15	- .70	- .95	
6 P.M.	33.73	- .76	- 1.47	- 1.81	- 2.07	
8 P.M.	30.78	+ 1.10	- .06	-.12	-.19	
10 P.M.	29.55	+ .40	+ .38	-.09	-.49	
12 Mid night	29.7	+ .62	+ 0.01	- .38	- .09	On 12th and 14th August inner tissue showed low temperature (-2° C to -3° C) at 8 a.m.
2 A.M.	28.2	+ .20	- .30	+ .10	- .20	
4 A.M.	27.6	- 1	- .24	- .20	- .40	
6 A.M.	27.0	- .38	- .30	- .08	- .05	These days were cloudy.

TABLE III  
*Comparison of mean departures of leaf temperatures  
 from air on August 4, 1931*  
*"Irrigated crop"*

Hours of observa- tion	Mean Temperature of air in ° C	Mean Temperature of Upper Surface in ° C	Mean Temperature of Lower Surface in ° C	Inner tissue		Remarks
				Mean Temperature of Petiole in ° C	Mean Temperature of Midrib in ° C	
6 A.M.	27.6 ± 21 Difference from air temperature	27.4 ± 31 - 2 ± 37 Insignificant	26.5 ± 30 - 1.10 ± 36 Insignificant	27.4 ± 28 - 2 ± 34 Insignificant	26.9 ± 29 - 7 ± 36 Insignificant	Leaves have al- most air tempera- ture
8 A.M.	28.5 ± 28 Difference from air temperature	34.7 ± 1.66 - 6.2 ± 1.68 Significant	32.7 ± 1.10 - 4.2 ± 1.13 Significant	32.0 ± 61 + 3.5 ± 67 Significant	31.7 ± 80 + 1.2 ± 66 Significant	Leaves warmer than air by about 3°C to 6°C
10 A.M.	35.9 ± 21 Difference from air temperature	37.0 ± 65 + 1.10 ± 68 Insignificant	36.5 ± 33 + 6 ± 39 Insignificant	35.3 ± 34 - 0.9 ± 39 Insignificant	35.4 ± 36 - 0.9 ± 43 Insignificant	Leaves show air tempera- ture
12 Noon	39.1 ± 38 Difference from air temperature	38.9 ± 37 - 2 ± 53 Insignificant	38.6 ± 40 - 5 ± 55 Insignificant	37.9 ± 20 - 1.2 ± 42 Insignificant	36.8 ± 30 - 2.3 ± 54 Significant	Do
2 P.M.	35.9 ± 14 Difference from air temperature	34.2 ± 40 - 1.8 ± 40 Significant	33.4 ± 37 - 2.5 ± 39 Significant	33.3 ± 27 - 2.6 ± 30 Significant	32.9 ± 44 - 3.0 ± 40 Significant	In the afternoon leaves are cooler by about 1°C, to 3°C than the air
4 P.M.	34.2 ± 18 Difference from air temperature	33.4 ± 10 - 8 ± 2 Significant	32.6 ± 36 - 1.8 ± 40 Significant	33.2 ± 15 - 1.0 ± 22 Significant	32.9 ± 10 - 1.3 ± 20 Significant	
6 P.M.	33.1 ± 10 Difference from air temperature	32.3 ± 20 - 8 ± 22 Significant	31.2 ± 19 - 1.9 ± 21 Significant	30.9 ± 21 - 1.2 ± 23 Significant	30.2 ± 20 - 1.9 ± 22 Significant	
8 P.M.	29.6 ± 64 Difference from air temperature	29.5 ± 7 - 1.1 ± 64 Insignificant	29.0 ± 61 - 0.0 ± 88 Insignificant	29.3 ± 48 - 1.7 ± 80 Insignificant	29.0 ± 6 - 6 ± 87 Insignificant	In the evening the leaves acquire air tem- perature again

higher in the tissues of petiole and midrib. From 8 A.M. onwards, a striking fall of temperature is observable and at 10 A.M. leaves acquire the air

temperature except the upper surface which is a little warmer. From 12 noon to 8 P.M. the leaves remain cooler by about 2 to 3 degrees than the air. Late in the evening the temperature of leaves begins to rise and corresponds with the air temperature. Throughout the night there was practically no difference between the air temperature and the temperature of the leaves. At midnight a slight rise above the air temperature had occurred in the case of upper surface.

September 1931.—During this month as given in Table IV and Fig. 2 the leaf temperatures are more or less similar to those of August except that the maximum temperature is attained at 10 A.M. instead of 8 A.M. in August. The leaves have air temperature in the morning at 6 A.M. There is a slow rise till 10 A.M., when the leaf temperature is about 3 degrees

TABLE IV

*Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during September 1931 (Irrigated crop)*

"Maximum flowering" Stage

Time	Air temperature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	-
6 A.M.	23.0	+ .07	+ .21	- .10	- .03
8 A.M.	27.59	+ 2.07	+ 1.20	+ 1.30	+ .78
10 A.M.	33.84	+ 3.57	+ 3.04	+ 1.90	+ 1.25
12 Noon	37.99	+ 2.37	+ 1.45	+ 1.79	+ .85
2 P.M.	38.66	- 1.27	- 1.50	- .26	- 1.54
4 P.M.	37.0	- 1.35	- 1.37	- .70	- 1.27
6 P.M.	29.49	- .48	- .71	.68	- .08
8 P.M.	27.7	- .22	- .45	+ .93	+ .65
10 P.M.	26.78	+ .07	+ .55	+ 1.35	+ 1.51
12 Mid night	25.48	+ .04	+ .18	+ 1.00	+ 1.33
2 A.M.	23.84	+ .37	- .22	+ .67	+ .65
4 A.M.	23.98	+ .40	- .45	+ .30	+ .49
6 A.M.	24.15	- .83	- .99	+ .28	+ .14

TABLE IV  
*Comparison of mean departures of leaf temperatures  
 from air on September 11, 1931  
 "Irrigated crop"*

Hours of observa- tion	Mean Tempera- ture of air in °C	Mean Tempera- ture of Upper Surface in °C	Mean Tempera- ture of Lower Surface in °C	Inner leaves		Remarks
				Mean Tempera- ture of Pitho- le in °C	Mean Tempera- ture of Midrib in °C	
6 A.M.	25.2 ± 31 Difference from air temperature	25.4 ± 25 + 2 ± 30 Insignificant	25.6 ± 37 + 4 ± 34 Insignificant	25.0 ± 29 - 2 ± 42 Insignificant	25.1 ± 31 - 1 ± 43 Insignificant	Leaves show air tempera- ture -
8 A.M.	29.0 ± 15 Difference from air temperature	31.8 ± 19 + 2.8 ± 24 Significant	30.6 ± 14 + 1.6 ± 20 Significant	30.8 ± 09 + 1.8 ± 14 Significant	30.2 ± 04 + 1.2 ± 15 Significant	Leaves definitely warmer than air by about 1°C to 4°C
10 A.M.	35.1 ± 26 Difference from air temperature	39.4 ± 43 + 4.3 ± 5 Significant	38.8 ± 51 + 3.7 ± 57 Significant	37.9 ± 29 + 2.8 ± 39 Significant	37.2 ± 20 + 2.1 ± 32 Significant	but the midrib acquires air tempera- ture at 12 Noon
12 Noon	38.2 ± 31 Difference from air temperature	42.0 ± 46 + 2.8 ± 55 Significant	40.9 ± 21 + 1.7 ± 37 Significant	41.0 ± 10 + 1.8 ± 34 Significant	40.1 ± 30 + 9 ± 41 Insignificant	
2 P.M.	40.0 ± 53 Difference from air temperature	37.1 ± 27 - 2.0 ± 60 Significant	36.5 ± 57 - 1.5 ± 87 Significant	38.0 ± 19 - 2.0 ± 62 Significant	37.8 ± 26 - 2.2 ± 65 Significant	Leaves cooler than air in the afternoon
4 P.M.	35.6 ± 36 Difference from air temperature	33.4 ± 16 - 2.2 ± 38 Significant	33.0 ± 12 - 2.6 ± 36 Significant	34.0 ± 09 - 1.1 ± 34 Significant	34.0 ± 10 - 1.6 ± 36 Significant	by about 1°C to 3°C
6 P.M.	31.2 ± 42 Difference from air temperature	30.4 ± 36 - 8 ± 55 Insignificant	32.2 ± 27 - 1.0 ± 48 Insignificant	30.5 ± 29 - 7 ± 50 Insignificant	30.1 ± 38 - 1.1 ± 57 Insignificant	Leaves show air tempera- ture in the even- ing
8 P.M.	28.9 ± 25 Difference from air temperature	28.6 ± 12 + 7 ± 27 Insignificant	29.4 ± 10 + 5 ± 27 Insignificant	29.5 ± 13 + 0 ± 28 Insignificant	29.3 ± 11 + 4 ± 27 Insignificant	

higher than the air temperature. Afterwards there is a steady fall and the leaves are cooler than the air upto 4 P.M. by about 1 to 1.5 degrees. It rises again and during night, the leaves remain slightly warmer than the air especially the inner tissue. At dawn the temperature of the leaves begins to be equal to that of the air temperature.

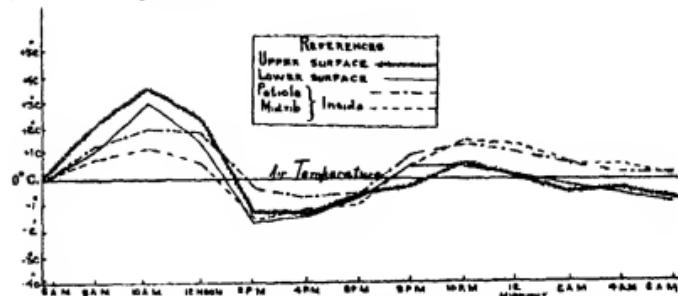


FIG. 2 Monthly mean departures of leaf temperatures of irrigated 4F Cotton Plants from air during September at different periods of the day.

October 1931.—It was observed that during the whole of October, the leaves did not behave uniformly with regard to adjustment with the air temperature. Therefore the data obtained for the month is examined in two portions. During the first fortnight, the variation of the leaf temperature is nearly the same as in August and September. At 6 A.M. there is a difference of about 1° C from the air temperature (Table V and Fig. 3).

TABLE V  
Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during October 1931 (1st half) (Irrigated crop)  
"Boll Development—Stray boll picking" Stage

Time	Air tem perature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
6 A.M.	23.94	+ .81	+ 1.11	+ .96	+ 1.06
8 A.M.	30.20	+ 4.7	+ 4.4	+ 2.4	+ 3.0
10 A.M.	33.96	+ 5.25	+ 3.78	+ 2.56	+ 2.31
12 Noon	36.8	+ 3.4	+ 3.0	+ 2.65	+ 3.0
2 P.M.	37.4	+ 1.2	+ 1.21	- 15	- .09
4 P.M.	32.0	+ .0	- 4	- 54	- 54
6 P.M.	27.4	+ .9	- .61	- 42	+ .51
8 P.M.	22.5	+ .67	+ .21	+ .61	+ .58

TABLE V  
*Comparison of mean departures of leaf temperatures  
 from air on October 8, 1931*  
*"Irrigated crop"*

Hours of observa- tion	Mean Temperature of air in °C	Mean Temperature of Upper Surface in °C	Mean Temperature of Lower Surface in °C	Inner tissues		Remarks
				Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
6 A.M.	25.1 ± 27 Difference from air temperature	26.4 ± 13 + 1.3 ± 20 Significant	26.2 ± 10 + 1.1 ± 13 Significant	26.4 ± 17 + 1.3 ± 32 Significant	26.9 ± 11 + 1.8 ± 29 Significant	
8 A.M.	29.4 ± 43 Difference from air temperature	34.4 ± 46 + 5.0 ± 66 Significant	33.7 ± 38 + 3.7 ± 39 Significant	32.8 ± 40 + 3.2 ± 69 Significant	32.8 ± 25 + 3.4 ± 50 Significant	From 6 A.M. to 2 P.M. the leaves are warmer than the air and differences are statis- tically signifi- cant but in the evening from 4 P.M. to 8 P.M. leaves show air tempera- ture
10 A.M.	34.5 ± 38 Difference from air temperature	40.2 ± 41 + 5.7 ± 56 Significant	39.1 ± 27 + 4.6 ± 46 Significant	37.2 ± 31 + 2.7 ± 37 Significant	37.5 ± 40 + 3.0 ± 56 Significant	
12 Noon	37.2 ± 40 Difference from air temperature	40.8 ± 21 + 3.6 ± 45 Significant	40.2 ± 17 + 3.0 ± 43 Significant	39.9 ± 12 + 2.7 ± 41 Significant	40.1 ± 18 + 2.9 ± 43 Significant	
2 P.M.	38.0 ± 31 Difference from air temperature	39.9 ± 18 + 1.9 ± 36 Significant	39.7 ± 12 + 1.7 ± 43 Significant	38.5 ± 18 + 5 ± 36 Insignificant	38.2 ± 16 + 2 ± 34 Insignificant	
4 P.M.	33.5 ± 36 Difference from air temperature	33.8 ± 24 + 3.3 ± 43 Insignificant	33.7 ± 23 + 2 ± 42 Insignificant	32.7 ± 18 - 8 ± 40 Insignificant	32.8 ± 16 - 7 ± 39 Insignificant	
6 P.M.	26.0 ± 28 Difference from air temperature	26.2 ± 16 + 2 ± 32 Insignificant	25.5 ± 18 - 8 ± 33 Insignificant	25.4 ± 19 - 6 ± 38 Insignificant	25.7 ± 20 - 3 ± 34 Insignificant	
8 P.M.	20.9 ± 25 Difference from air temperature	21.6 ± 12 + 7 ± 27 Insignificant	21.4 ± 10 + 6 ± 27 Insignificant	21.5 ± 13 + 6 ± 28 Insignificant	21.3 ± 11 + 4 ± 27 Insignificant	

The temperature then rises till 8 A.M. or 10 A.M. but at a different rate. From 10 A.M. onwards there is a fall which continues till the afternoon. At 4 P.M. leaves show air temperature and tend to remain so till the evening; the upper surface, however, shows slightly higher temperatures at 6 and 8 P.M. In October, night readings were abandoned on account of cold.

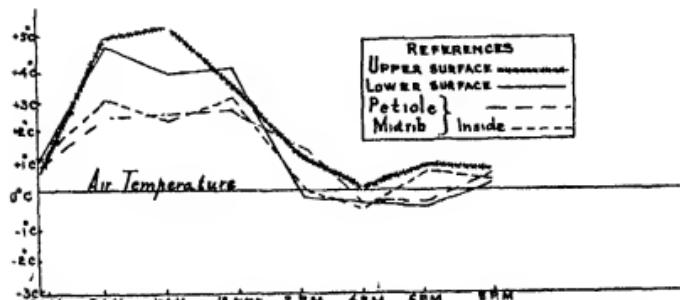


FIG. 3 Monthly mean departures of leaf temperatures of irrigated 4F Cotton Plants during October (I part) at different periods of the day

October 1931 (2nd half) — The most striking feature of the behaviour of leaves in this part of the month as shown in Table VI, Fig. 4, is that at no time the temperature of leaves fell below the air temperature. At 7 A.M. the upper surface and lower surface start with almost air temperature, but the temperature of petiole and midrib is higher by 1°C. The departure

TABLE VI

Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during October 1931 (2nd half) (Irrigated crop)  
"Stray boll picking" Stage

Time	Air temperature in degrees C	Upper surface	Lower surface	Inner tissue		Remarks
				Petiole	Midrib	
7 A.M.	12.5	+ 30	+ 51	+ 90	+ 12	
9 A.M.	21.5	+ 4.27	+ 3.03	+ 2.7	+ 3.42	
11 A.M.	28.4	+ 5.93	+ 3.92	+ 2.76	+ 1.73	
1 P.M.	31.55	+ 6.46	+ 5.00	+ .35	+ .33	
3 P.M.	28.60	+ 7.39	+ 6.8	+ 1.38	+ 1.84	
5 P.M.	22.60	+ 2.16	+ 1.8	+ 1.0	+ .85	
7 P.M.	15.81	+ .98	+ 1.02	+ .82	+ .97	

TABLE VI

*Comparison of mean departures of leaf temperatures  
from air on October 23, 1931*  
"Irrigated crop"

Hours of observa- tion	Mean Temperature of air in °C	Mean Temperature of Upper Surface in °C	Mean Temperature of Lower Surface in °C	Inner tissue		Remarks
				Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
7 A.M.	12.0 ± 20 Difference from air temperature	12.6 ± 18 + 6 ± 27 Insignificant	12.7 ± 20 + 7 ± 28 Insignificant	13.0 ± 08 + 1.0 ± 21 Significant	13.5 ± 05 + 1.5 ± 20 Significant	The inner tissue is warmer than the upper and lower surface show air tempera- ture in the morn- ing
9 A.M.	19.5 ± 36 Difference from air temperature	24.7 ± 31 + 5.2 ± 47 Significant	23.4 ± 26 + 3.0 ± 44 Significant	22.7 ± 18 + 3.2 ± 40 Significant	23.1 ± 19 + 3.6 ± 40 Significant	upper and lower surface show air tempera- ture in the morn- ing
11 A.M.	24.7 ± 26 Difference from air temperature	27.9 ± 30 + 3.2 ± 39 Significant	27.8 ± 26 + 3.1 ± 35 Significant	26.4 ± 20 + 1.7 ± 17 Significant	26.2 ± 20 + 1.5 ± 32 Significant	inner tissue is warmer than the upper and lower surface show air tempera- ture in the morn- ing
1 P.M.	25.0 ± 40 Difference from air temperature	31.1 ± 50 + 5.2 ± 64 Significant	30.2 ± 30 + 4.3 ± 50 Significant	28.0 ± 15 + 2.1 ± 42 Significant	27.9 ± 12 + 2.0 ± 41 Significant	The leaves remain warmer through- out the day up to 7 P.M.
3 P.M.	26.5 ± 26 Difference from air temperature	29.0 ± 19 + 2.5 ± 32 Significant	29.1 ± 15 + 2.6 ± 30 Significant	28.7 ± 11 + 2.2 ± 28 Significant	28.5 ± 14 + 2.0 ± 29 Significant	than the air except that of inner tissue which at 7 P.M.
5 P.M.	20.1 ± 30 Difference from air temperature	22.9 ± 15 + 2.8 ± 33 Significant	22.5 ± 13 + 2.4 ± 32 Significant	22.4 ± 11 + 2.3 ± 32 Significant	22.6 ± 18 + 2.5 ± 36 Significant	shows air tempera- ture
7 P.M.	14.2 ± 40 Difference from air temperature	16.0 ± 40 + 1.8 ± 57 Significant	15.9 ± 30 + 1.7 ± 6 Significant	15.0 ± 16 + 8 ± 42 Insignificant	15.1 ± 12 + 0 ± 41 Insignificant	

from the air temperature in the case of upper and lower surfaces goes on increasing till 3 P.M. when it stands at +7.39° and +6.80° C respectively. After 3 P.M. a fall sets in and at 7 P.M. the temperature approaches that of the air. The temperature of petiole and midrib increased upto 9 A.M. and then fell to that of the air at 1 P.M. It rose again at 3 P.M. At 7 P.M. it was about 1° C above the air temperature.

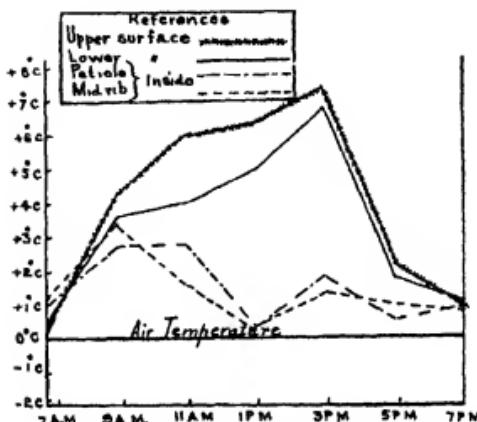


Fig. 4 Monthly mean departures of leaf temperatures of irrigated 4P Cotton Plants from air during October (II part) at different periods of the day.

November 1931—As represented in Table VII, Fig. 5, the data indicate that the leaves in the morning at 7 A.M. have the same temperature as the

TABLE VII

Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during November 1931 (Irrigated crop)  
"Peak of boll picking" Stage

Time	Air temperature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
7 A.M.	9.0	+ 1	+ 2	+ 0	+ 1
9 A.M.	15.43	+ 4.37	+ 4.11	+ 1.89	+ 2.01
11 A.M.	26.94	+ 3.75	+ 3.68	+ 1.35	+ 1.15
1 P.M.	26.73	+ 5.22	+ 4.63	+ 1.94	+ 1.77
3 P.M.	26.91	+ 2.85	+ 2.58	+ 1.9	+ 1.01
5 P.M.	18.96	+ 3.43	+ 2.86	+ 1.98	+ 2.23
7 P.M.	13.54	+ 1.70	+ 1.39	+ .74	+ .80

TABLE VII

Comparison of mean departures of leaf temperatures  
from air on November 15, 1931

"Irrigated crop"

Hours of observa- tion	Mean Tempera- ture of air in °C	Mean Tempera- ture of Upper Surface in °C	Mean Tempera- ture of Lower Surface in °C	Inner tissue		Remarks
				Mean Tempera- ture of Petiole in °C	Mean Tempera- ture of Midrib in °C	
7 A.M.	11.0 ± 1.0 Difference from air temperature	11.2 ± 1.0 - 2 ± 1.8 Insignificant	11.4 ± 1.1 + 4 ± 1.0 Insignificant	11.3 ± 1.2 + 3 ± 2 Insignificant	11.2 ± 1.4 - 2 ± 2.1 Insignificant	Leaves show air tempera- ture in the morning
9 A.M.	16.5 ± 2.0 Difference from air temperature	21.7 ± 2.5 + 5.2 ± 3.2 Significant	21.8 ± 1.6 + 5.1 ± 2.5 Significant	18.4 ± 1.2 + 1.9 ± 2.3 Significant	18.0 ± 1.8 + 2.4 ± 2.7 Significant	
11 A.M.	25.0 ± 3.0 Difference from air temperature	29.2 ± 2.5 + 4.2 ± 3.9 Significant	29.1 ± 3.0 + 4.1 ± 4.2 Significant	26.7 ± 2.0 + 1.7 ± 3.0 Significant	26.4 ± 1.5 + 1.5 ± 3.3 Significant	The leaves remain warmer through out the day than the air
1 P.M.	26.0 ± 4.1 Difference from air temperature	31.9 ± 3.0 + 5.9 ± 5.0 Significant	30.8 ± 3.5 + 4.8 ± 5.4 Significant	28.2 ± 2.0 + 2.2 ± 4.5 Significant	28.0 ± 2.1 + 2.0 ± 4.6 Significant	but the inner tissue shows air tempera- ture in the even- ing
3 P.M.	36.7 ± 4.5 Difference from air temperature	39.9 ± 4.0 + 3.2 ± 6.0 Significant	39.7 ± 4.2 + 3.0 ± 6.0 Significant	38.8 ± 2.5 + 2.1 ± 5.1 Significant	38.6 ± 2.3 + 1.8 ± 5.5 Significant	
5 P.M.	16.8 ± 3.7 Difference (from air temperature	20.6 ± 2.6 + 3.8 ± 4.5 Significant	20.4 ± 2.3 + 3.6 ± 4.3 Significant	19.1 ± 1.6 + 2.3 ± 4.0 Significant	19.4 ± 1.2 + 2.6 ± 3.8 Significant	
7 P.M.	14.2 ± 4.0 Difference from air temperature	16.0 ± 4.0 + 1.8 ± 5.7 Significant	15.9 ± 3.0 + 1.7 ± 5 Significant	15.0 ± 1.5 + 8 ± 4.2 Insignificant	15.1 ± 1.2 + 9 ± 4.1 Insignificant	

air. After sunrise at 7-30 A.M. the temperature of leaves begins to rise rapidly. The temperatures of upper and lower surfaces continue to rise till 1 P.M., afterwards it falls slowly. At 7 P.M. they are warmer than the air by about 1.5°C. The temperature of the inner tissue remains almost constant after 9 A.M. but at 7 P.M. they show negligible difference from the air temperature.

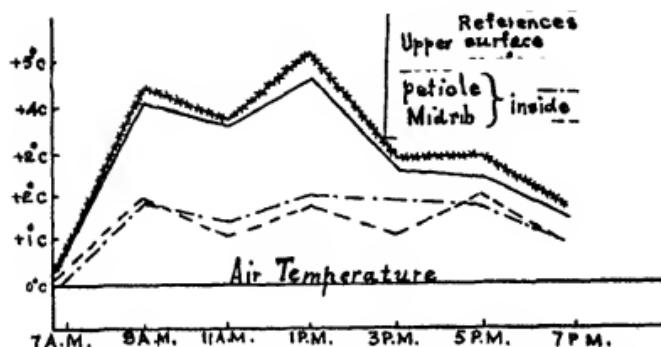


FIG. 5 Monthly mean departures of leaf temperatures of irrigated 4F Cotton Plants from air during November at different periods of the day

#### *IV (B) Leaf temperatures and soil moisture*

It has been observed that the amount of water available in the soil has a marked influence on the temperature of leaves. To study the effect of moisture content, temperature determinations were made on leaves of 4F cotton crop grown in a field which received no irrigation after sowing. But as some rains fell in July and August the crop was as good as the irrigated one.

Sometimes in September, however, the unirrigated crop showed signs of wilting. Determinations of the moisture contents of the soil in the irrigated and unirrigated plots were made from time to time and are given in Table VIII which shows that, as compared with the irrigated crop, the unirrigated crop was deficient in soil moisture.

TABLE VIII

*Percentage of soil moisture at various depths of irrigated and unirrigated fields during different months*

Month	Nature of the field	Depth	Percentage of soil moisture	
August 1931	Irrigated	1st ft	9.10	There was about 4.31 inches of rainfall
		2nd ft	12.04	
		3rd ft	11.77	
	Unirrigated	1st ft	9.10	
		2nd ft	11.72	
		3rd ft	10.86	
September 1931	Irrigated	1st ft	11.02	Samples for moisture determination were always taken 5-6 days after the last irrigation during August, September and October but 8-10 days in November
		2nd ft	12.30	
		3rd ft	13.04	
	Unirrigated	1st ft	1.09	
		2nd ft	5.72	
		3rd ft	8.25	
October 1931	Irrigated	1st ft	10.27	
		2nd ft	11.38	
		3rd ft	12.21	
	Unirrigated	1st ft	94	
		2nd ft	4.59	
		3rd ft	6.39	
November 1931	Irrigated	1st ft	14.74	
		2nd ft	15.81	
		3rd ft	16.67	
	Unirrigated	1st ft	74	
		2nd ft	4.71	
		3rd ft	6.27	

#### *Temperature of Unirrigated Cotton*

September 1931.—The temperature of leaves was measured after every two hours. The data obtained are given in Table IX and Fig. 6. It will be seen that in the morning at 6 A.M. leaves show air temperature. The temperature of leaves afterwards begins to rise till the maximum value is recorded at 2 P.M. when leaves show a departure of about  $+4^{\circ}\text{C}$  and inner tissue of  $+2^{\circ}\text{C}$  to  $+3^{\circ}\text{C}$  from the air temperature. Afterwards the leaf temperature continues to fall but still remains higher than that of the air till 6 P.M. when the temperature of the leaves and the air are the same. This condition is maintained up to daybreak. The inner tissue, however, shows a spasmodic rise and fall throughout the night. It is noteworthy

that leaves of the irrigated crop on the other hand have a lower temperature than the air from 12 to 6 P.M. by about  $-2^{\circ}\text{C}$  to  $-3^{\circ}\text{C}$  (Table I, Fig. 1)

TABLE IX

*Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during September 1931 (Unirrigated crop)*

*"Maximum flowering" Stage*

Time	Air tem perature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
6 A.M.	23.5	+ 23	+ 23	+ .35	+ .30
8 A.M.	27.1	+ 2.00	+ 1.80	+ 1.00	+ 1.3
10 A.M.	32.06	+ 3.00	+ 2.90	+ 2.09	+ 1.90
12 Noon	34.77	+ 3.25	+ 3.00	+ 1.00	+ 1.80
2 P.M.	37.89	+ 4.43	+ 4.61	+ 2.99	+ 2.23
4 P.M.	35.54	+ 2.49	+ 3.15	+ .81	- .24
6 P.M.	31.02	+ 5	+ .25	+ .50	+ .00
8 P.M.	27.8	- 9	+ 61	+ 1.2	+ .81
10 P.M.	27.11	- 51	+ 21	+ .24	+ .24
12 Midnight	27.93	- 63	- .30	+ 1.92	+ 2.66
2 A.M.	26.31	+ 19	+ .38	+ 1.00	+ 2.50
4 A.M.	25.98	+ 61	+ .33	+ .00	+ .20
6 A.M.	24.42	+ 91	- .05	+ .67	+ .25

TABLE IX

*Comparison of mean departures of leaf temperatures  
from air on September 11, 1931*

*" Unirrigated crop "*

Hours of observa- tion	Mean Tempera- ture of air in °C	Mean Tempera- ture of Upper Sur- face in °C	Mean Tempera- ture of Lower Sur- face in °C	Inner tissue		Remarks
				Mean Tempera- ture of Petiole in °C	Mean Tempera- ture of Midrib in °C	
6 A.M.	24.7 ± 25 Difference from air temperature	25.0 ± 15 + 3 ± 29 Insignificant	24.9 ± 18 + 2 ± 30 Insignificant	25.0 ± 13 + 3 ± 28 Insignificant	24.9 ± 16 + 2 ± 29 Insignificant	
8 A.M.	29.4 ± 32 Difference from air temperature	32.3 ± 16 + 2.9 ± 36 Significant	31.4 ± 20 + 2.0 ± 37 Insignificant	31.1 ± 12 + 1.7 ± 34 Significant	31.0 ± 11 + 1.0 ± 33 Significant	
10 A.M.	35.3 ± 40 Difference from air temperature	38.4 ± 20 + 3.1 ± 44 Significant	38.2 ± 20 + 2.9 ± 44 Significant	37.8 ± 16 + 2.5 ± 43 Significant	37.5 ± 15 + 2.2 ± 42 Significant	In the morning and even- ing leaves show air tempera- ture, but remain consider- ably and signifi- cantly warmer
12 Noon	38.7 ± 35 Difference from air temperature	42.3 ± 26 + 3.6 ± 43 Significant	42.2 ± 15 + 3.5 ± 36 Significant	40.9 ± 20 + 2.2 ± 40 Significant	40.8 ± 20 + 2.1 ± 40 Significant	than the air in the fore and afternoon by about 2°C to 5°C
2 P.M.	39.4 ± 43 Difference from air temperature	44.6 ± 50 + 5.2 ± 67 Significant	44.9 ± 45 + 5.5 ± 63 Significant	42.9 ± 35 + 3.5 ± 57 Significant	42.7 ± 30 + 3.3 ± 54 Significant	
4 P.M.	36.1 ± 40 Difference from air temperature	38.3 ± 25 + 2.2 ± 47 Significant	38.8 ± 25 + 2.7 ± 44 Significant	36.6 ± 21 + 1.5 ± 46 Insignificant	36.2 ± 15 + 1.1 ± 45 Insignificant	
6 P.M.	31.4 ± 42 Difference from air temperature	31.8 ± 30 + 4.4 ± 56 Insignificant	31.7 ± 40 + 7 ± 56 Insignificant	31.5 ± 30 + 1 ± 5 Insignificant	31.3 ± 25 + 1 ± 48 Insignificant	
8 P.M.	29.6 ± 30 Difference from air temperature	29.0 ± 15 + 6 ± 33 Insignificant	29.2 ± 20 + 4 ± 36 Insignificant	29.7 ± 16 + 1 ± 34 Insignificant	29.9 ± 12 + 3 ± 32 Insignificant	

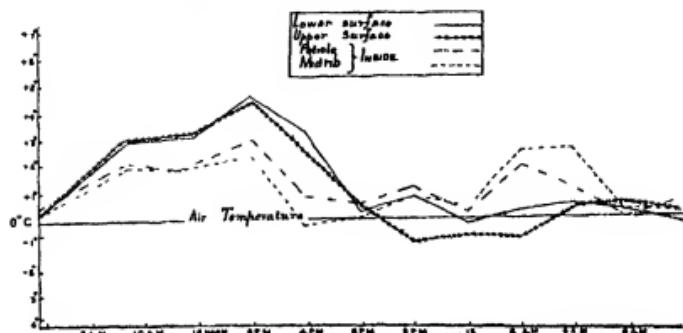


FIG. 6 Monthly mean departures of leaf temperatures of unirrigated 4F Cotton Plants from air during September at different periods of the day

October 1931 (1st half) — At 6 A.M. the temperature of the plants is slightly lower than air temperatures (Table X, Fig. 8). After 6 A.M. there is a rapid rise of temperature in all parts of the leaves and the maxima is reached at 10 A.M. At this time, upper surface, lower surface, petiole and the

TABLE X

Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during October 1931 (1st half) (Unirrigated crop)

"Stray boll picking" Stage

Time	Air tem perature in degrees C°	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
6 A.M.	23.71	-1.37	+2.25	-1.08	-1.53
8 A.M.	30.5	+3.46	+3.14	+1.35	+1.72
10 A.M.	33.34	+6.04	+4.15	+2.87	+2.82
12 Noon	38.29	+5.95	+3.50	+1.41	+0.95
2 P.M.	39.30	+3.92	+2.90	+2.17	+2.03
4 P.M.	35.23	+1.89	+1.22	+0.52	+0.51
6 P.M.	28.72	+0.34	-0.77	+0.14	-0.08
8 P.M.	22.12	+1.15	+0.73	+1.29	+1.07

TABLE X  
*Comparison of mean departures of leaf temperatures  
 from air on October 8, 1931  
 "Unirrigated crop"*

Hours of obser- vation	Mean Temperature of air in °C	Mean Temperature of Lower Surface in °C	Mean Temperature of Upper Surface in °C	Inner tissue		Remarks
				Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
6 A.M.	24.0 ± 20 Difference from air temperature	23.0 ± 15 - 1.6 ± 25 Significant	24.0 ± 13 - 6.0 ± 23 Insignificant	23.1 ± 16 1.5 ± 23 Significant	23.2 ± 15 - 1.4 ± 25 Significant	In the morning the leaves are cool paratively soil than the air, but remain warmer
8 A.M.	30.8 ± 42 Difference from air temperature	34.0 ± 25 + 4.1 ± 48 Significant	34.8 ± 30 + 4.0 ± 51 Significant	32.3 ± 18 1.1 ± 42 Significant	32.8 ± 25 + 2.0 ± 48 Significant	From 8 A.M. to 4 P.M. and acquire air tem- perature again in the even- ing
10 A.M.	34.2 ± 50 Difference from air temperature	41.4 ± 52 + 7.2 ± 7 Significant	39.0 ± 40 + 5.4 ± 64 Significant	37.3 ± 20 1.1 ± 53 Significant	37.2 ± 25 + 3.0 ± 55 Significant	
12 Noon	37.9 ± 78 Difference from air temperature	42.5 ± 40 + 4.6 ± 55 Significant	41.0 ± 35 - 1.0 ± 51 Significant	39.7 ± 20 1.8 ± 42 Significant	39.5 ± 18 + 1.6 ± 42 Significant	
2 P.M.	40.5 ± 40 Difference from air temperature	43.5 ± 30 + 3.5 ± 5 Significant	42.4 ± 25 + 2.4 ± 47 Significant	42.1 ± 20 + 2.1 ± 44 Significant	42.3 ± 15 + 2.3 ± 42 Significant	
4 P.M.	34.5 ± 30 Difference from air temperature	36.8 ± 40 + 2.3 ± 5 Significant	36.5 ± 32 + 2.0 ± 43 Significant	35.2 ± 20 + 7.2 ± 35 Insignificant	35.4 ± 20 + 9.2 ± 35 Insignificant	
6 P.M.	29.2 ± 15 Difference from air temperature	29.7 ± 20 + 5.5 ± 25 Insignificant	29.1 ± 15 - 1.3 ± 21 Insignificant	29.4 ± 12 + 2.2 ± 19 Insignificant	29.6 ± 10 + 3.3 ± 21 Insignificant	
8 P.M.	21.4 ± 30 Difference from air temperature	22.0 ± 15 + 0.6 ± 33 Insignificant	22.1 ± 20 + 7.7 ± 35 Insignificant	22.2 ± 15 + 8.8 ± 33 Insignificant	22.1 ± 13 + 7.7 ± 32 Insignificant	

midrib showed a departure of +6.04°C, +4.15°C, +2.87°C, and +2.82°C, respectively. From 10 A.M. onwards there is a gradual fall till 6 P.M., when the plants acquire air temperature. The inner tissue shows a slight rise of temperature again at 2 P.M. At 8 P.M. again the plants show higher temperature than the air by about 1°C.

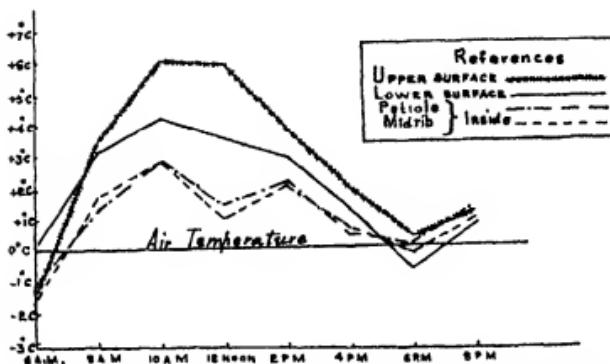


FIG. 7 Monthly mean departures of leaf temperatures of unirrigated 4F Cotton Plants from air during October (1 part) at different periods of the day

October 1931 (2nd half) and November 1931.—The data obtained is compiled in Tables XI and XII and Figs. 8 and 9 respectively. The

TABLE XI

Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during October 1931 (2nd half) (Unirrigated crop)  
"Stray boll picking" Stage

Time	Air tem perature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
7 A.M.	13.6	+ .85	- .03	- .70	- 1.36
9 A.M.	22.48	+ 3.74	+ 3.01	+ 2.6	+ 2.55
11 A.M.	29.52	+ 2.81	+ 2.23	+ .11	+ .73
1 P.M.	32.08	+ 3.17	+ 2.30	+ 1.71	+ 1.51
3 P.M.	30.46	+ 1.13	+ .60	+ .47	+ .90
5 P.M.	23.70	+ 2.23	+ 1.03	+ 2.12	+ 1.48
7 P.M.	13.37	+ .73	+ .03	+ 1.38	+ 1.22

TABLE XI

Comparison of mean departures of leaf temperatures  
from air on October 29, 1931

"Unirrigated crop"

Hours of observa- tion	Mean Temperature of air in °C	Mean Temperature of Upper Surface in °C	Mean Temperature of Lower Surface in °C	Inner tissue		Remarks
				Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
7 A.M.	13.0 ± 40 Difference from air temperature	13.7 ± 35 + 7 ± 53 Insignificant	13.4 ± 30 + 4 ± 5 Insignificant	13.2 ± 20 + 2 ± 44 Insignificant	12.2 ± 10 - 8 ± 53 Insignificant	The leaves in this month show very erratic behaviour
9 A.M.	20.4 ± 45 Difference from air temperature	24.6 ± 40 + 4 ± 60 Significant	24.8 ± 25 + 4 ± 51 Significant	23.4 ± 15 + 3 ± 47 Significant	23.4 ± 10 + 3 ± 48 Significant	
11 A.M.	26.7 ± 35 Difference from air temperature	29.9 ± 30 + 3 ± 46 Significant	29.4 ± 35 + 2 ± 49 Significant	27.2 ± 18 + 5 ± 30 Insignificant	27.8 ± 14 + 1.2 ± 37 Significant	
1 P.M.	27.4 ± 25 Difference from air temperature	30.0 ± 30 + 3.5 ± 39 Significant	30.1 ± 40 + 2.7 ± 47 Significant	29.4 ± 2 + 2.0 ± 32 Significant	29.2 ± 30 + 1.8 ± 30 Significant	
3 P.M.	27.8 ± 32 Difference from air temperature	29.6 ± 20 + 1.8 ± 37 Significant	28.8 ± 20 + 1.0 ± 17 Insignificant	28.6 ± 10 + 1.8 ± 36 Insignificant	28.8 ± 15 + 1.0 ± 35 Insignificant	
5 P.M.	22.0 ± 18 Difference from air temperature	23.9 ± 30 + 1.9 ± 35 Significant	23.9 ± 20 + 1.9 ± 27 Significant	23.5 ± 16 + 1.5 ± 24 Significant	23.8 ± 18 + 1.8 ± 25 Significant	
7 P.M.	14.1 ± 40 Difference from air temperature	14.9 ± 30 + 8 ± 5 Insignificant	14.5 ± 35 + 4 ± 57 Insignificant	15.0 ± 25 + 9 ± 47 Insignificant	14.9 ± 20 + 8 ± 49 Insignificant	

leaves show air temperature at 7 A.M. except the inner tissue during October 2nd half after which there is a rapid rise till maximum is reached at 9 A.M. From this time onwards temperature of the plant rises and falls till 7 P.M., when the plants are warmer than the air by about 1°C to 2°C, but the inner tissue during November becomes cooler at 5 P.M. and remains so till 7 P.M.

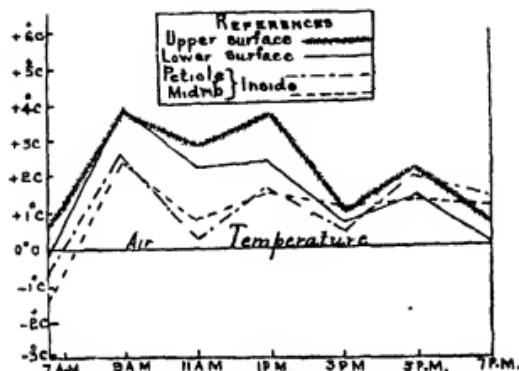


FIG. 8. Monthly mean departures of leaf temperatures of unirrigated 4F Cotton Plants from air during October (II part) at different periods of the day.

TABLE XII

Monthly mean departures of leaf temperatures (columns 3-5) from air (column 2) during November 1931 (Unirrigated crop)

"Peak of boll picking" Stage

Time	Air temperature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
7 A.M.	10.92	- +4.2	- 4.5	- 1.8	- 1.6
8 A.M.	21.19	+ 3.43	1.3.02	+ 2.92	+ 2.72
11 A.M.	27.98	+ 3.41	+ 3.16	+ 1.01	+ 1.14
1 P.M.	32.89	1.2.80	+ 2.77	+ .99	+ .4
3 P.M.	30.90	+ 2.95	+ 3.09	+ 1.27	+ 1.19
5 P.M.	20.60	+ 3.21	+ 2.83	- .72	- .41
7 P.M.	15.71	+ 2.06	+ .94	- .72	- .82

TABLE XII

Comparison of mean departures of leaf temperatures from air on  
November 13 during 1931 "Unirrigated crop"

Hours of observa- tion	Mean Tempera- ture of air in °C	Mean Tempera- ture of Upper Surface in °C	Mean Tempera- ture of Lower Surface in °C	Inner tissue		Remarks
				Mean Tempera- ture of Petiole in °C	Mean Tempera- ture of Midrib in °C	
7 A.M.	12.2 ± 30 Difference from air temperature	12.0 ± 20 - 2 ± 36 Insignificant	11.8 ± 15 4 ± 31 Insignificant	12.0 ± 15 - 2 ± 33 Insignificant	12.0 ± 15 - 2 ± 33 Insignificant	In the morning and even- ing the leaves have temper- ature of the air, but are warmer signif- cantly than the air from 9 A.M. to 1 P.M.
9 A.M.	16.4 ± 40 Difference from air temperature	22.2 ± 25 13.8 ± 47 Significant	22.4 ± 18 14.0 ± 43 Significant	21.8 ± 16 +3.4 ± 43 Significant	21.5 ± 12 1.3 ± 41 Significant	
11 A.M.	28.1 ± 46 Difference from air temperature	31.8 ± 37 13.7 ± 59 Significant	31.6 ± 28 13.5 ± 54 Significant	30.0 ± 11 11.9 ± 47 Significant	29.8 ± 10 11.7 ± 47 Significant	
1 P.M.	32.4 ± 20 Difference from air temperature	35.7 ± 15 +3.3 ± 25 Significant	35.5 ± 16 13.1 ± 25 Significant	33.6 ± 11 +1.2 ± 22 Significant	33.4 ± 10 1.8 ± 22 Significant	
3 P.M.	28.6 ± 40 Difference from air temperature	31.7 ± 25 +3.1 ± 47 Significant	31.8 ± 18 +3.2 ± 43 Significant	30.1 ± 20 11.5 ± 44 Significant	30.2 ± 16 11.6 ± 43 Significant	
6 P.M.	18.8 ± 35 Difference from air temperature	21.3 ± 20 12.5 ± 4 Significant	21.5 ± 22 12.7 ± 41 Significant	18.0 ± 16 - 8 ± 38 Insignificant	18.1 ± 16 7 ± 38 Insignificant	
7 P.M.	15.4 ± 40 Difference from air temperature	17.8 ± 15 12.4 ± 42 Significant	18.1 ± 43 + 7.1 ± 43 Insignificant	14.6 ± 12 - 3 ± 41 Insignificant	14.8 ± 13 - 0 ± 42 Insignificant	

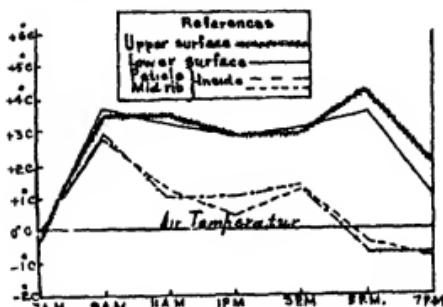


FIG. 9 Monthly mean departures of leaf temperatures of unirrigated 4F Cotton Plants from air during November at different periods of the day.

*V. (C) Moisture content of the leaves during growth period of the plant*

The amount of water present in cotton leaves was determined after an interval of two hours from 6 A.M. to 8 P.M. once a week during the months of August–November. The leaves were dried to a constant weight in an electric oven. Average results obtained for the irrigated and unirrigated crop are given in Table XIII and Figs. 10 and 11.

Reference to these tables and figures shows how the amount of moisture in the leaves of irrigated crop varies in the course of a day. In August, the leaves have 82 per cent moisture in the morning. Then it begins to decrease with the advance of the day till at 2 P.M. the leaves are left with only 61 per cent of moisture. After 2 P.M. moisture content of the leaves again begins to increase. At 8 P.M. it goes upto 73 per cent. In September, the percentages of moisture in leaves is reduced by about 5 per cent as compared with August. Correspondingly the extent of variability is affected. Similarly in October and November there is a marked fall in the percentage of moisture in the leaves and degree of variability. With regard to the unirrigated crop the leaves during the course of the day and during different months from September to November always showed about 5 to 10 per cent less moisture (Table XIII, Fig. 11).

Variability of moisture was also significantly less. The leaves got wilted daily after 10 A.M. and remained so till 4 P.M. It may be pointed out that the rate of transpiration of leaves varies considerably in different months and under different conditions of soil moisture.

TABLE XIII

Percentage moisture in the leaves of 4F cotton plants during different months of the growing seasons  
at different times of the day in 1951

(On dry weight basis)

Kind of crop	Month	Time										Age of plants in days					
		6 A.M.	7 A.M.	8 A.M.	9 A.M.	10 A.M.	11 A.M.	12 Noon	1 P.M.	2 P.M.	3 P.M.	4 P.M.	5 P.M.	6 P.M.	7 P.M.	8 P.M.	
Im- paled	Aug.	88.03	-	80.54	-	74.87	-	69.36	-	60.59	-	63.68	-	72.42	-	72.93	90
	Sept.	78.12	-	75.47	-	75.40	-	73.10	-	63.83	-	65.67	-	75.38	-	75.58	120
	Oct.	76.03	-	73.63	-	72.47	-	70.02	-	68.89	-	68.22	-	72.16	-	73.64	160
Nor.	Nov.	73.90	-	72.50	-	70.22	-	70.80	-	72.80	-	72.30	-	75.20	-	75.30	180
	Sept.	74.89	-	71.62	-	69.27	-	67.10	-	66.72	-	67.81	-	69.24	-	70.79	120
	Oct.	72.22	-	70.01	-	66.64	-	65.43	-	63.71	-	71.66	-	71.96	-	71.96	130
Nor.	Nov.	69.39	-	67.51	-	66.66	-	66.31	-	67.40	-	67.81	-	69.87	-	69.87	180

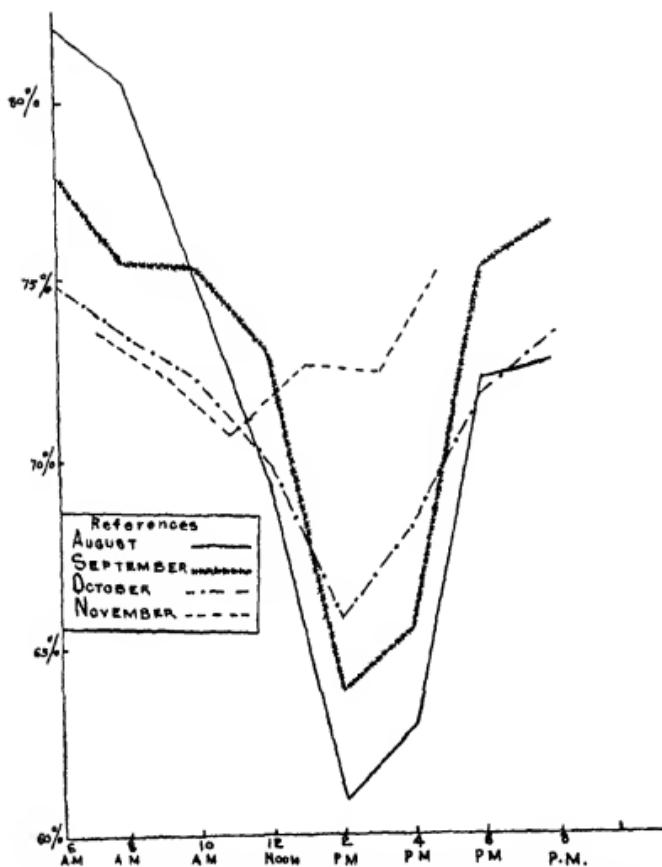


FIG. 10. Variation in the amount of moisture in the leaves of irrigated 4F Cotton Plants during various months at different periods in the day.

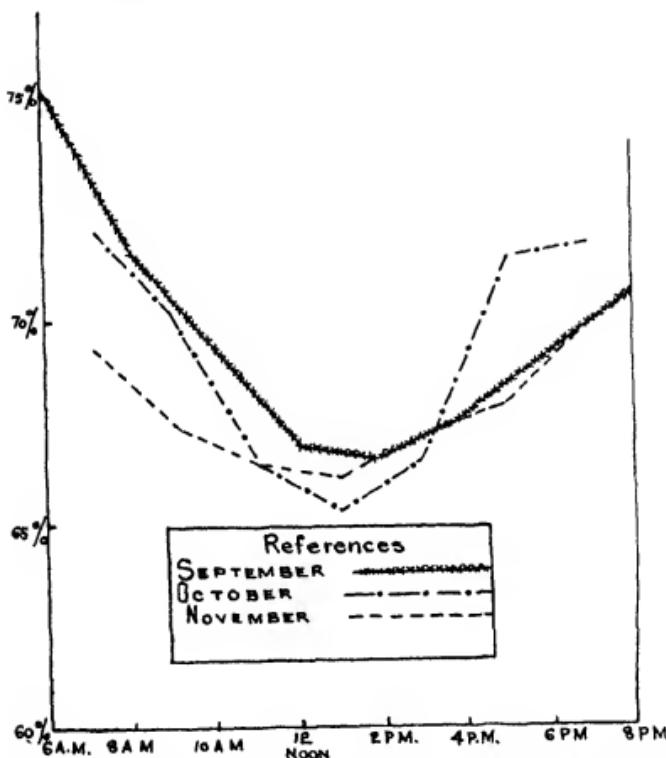


FIG. 11. Variation in the amount of moisture in the leaves of unirrigated 4F Cotton Plants during various months at different periods in the day.

#### *VI (D) Temperature and age of leaves*

Usually plants behave like cold-blooded animals whose temperature varies with the medium in which they live. This, however, is true within certain limits. The temperature regulatory power of the leaves diminishes with age as is indicated by Table XIV.

TABLE XIV.

*Mean temperature departures of old and young leaves from air temperature at different periods of the day during September and October 1931*

*Average of 50 determinations*

Time	Air Temperature in °C	Nature of Leaves	Mean Temperature of Upper Surface in °C	Mean Temperature of Lower Surface in °C	Inner tissue		Remarks
					Mean Temperature of Petiole in °C	Mean Temperature of Midrib in °C	
7 A.M.	21.2	Old	+1.0 ± 30	+1.4 ± 20	+1.0 ± 30	+1.1 ± 35	Old leaves were 40-46 days old and young leaves were 25-30 days old
		Young	+ 5 ± 25	+ 3 ± 28	+ 4 ± 15	+ 3 ± 17	
	Difference (Old - Young)	Old	+1.1 ± 39	+1.1 ± 34	+ 6 ± 25	+ 8 ± 38	
		Young	Insignificant	Insignificant	Insignificant	Insignificant	
1 P.M.	31.0	Old	+ 5 ± 15	+ 3 ± 10	-0.1 ± 16	-0.3 ± 12	25-30 days old
		Young	-1.7 ± 20	+ 1.8 ± 06	-2.0 ± 12	-2.3 ± 10	
	Difference (Old - Young)	Old	+ 2 ± 25	+ 2.1 ± 15	+ 1.0 ± 20	+ 2.0 ± 15	
		Young	Significant	Significant	Significant	Significant	
7 P.M.	28.4	Old	+ 2 ± 16	+ 1 ± 12	+ 4 ± 16	+ 1 ± 18	25-30 days old
		Young	+ 1 ± 10	+ 0.0 ± 13	- 2 ± 16	- 4 ± 20	
	Difference (Old - Young)	Old	+ 1 ± 18	+ 1 ± 17	+ 6 ± 26	+ 5 ± 27	
		Young	Insignificant	Insignificant	Insignificant	Insignificant	

50 determinations were made on the temperature of young and old leaves at different periods of the day. The results obtained show that the leaves when young are able to adjust themselves readily with the air temperature as it fluctuates. In the afternoon when air temperature rises, temperature of young leaves never rises above the air, but on the other hand old leaves get warmed up and show a higher temperature. Perhaps, this is due to the fact that the young leaves have greater moisture content and are thus better able to conduct energy. The old leaves have been found to contain less moisture, and consequently remain warmer than the air.

In the morning and evening mean temperature differences of the old and young leaves are statistically insignificant but at 1 P.M. the old leaves are warmer as compared with young leaves by about 2.0°C. and the difference is highly significant.

#### VII (E) Temperature of wilted leaves

In this connection measurements of temperature were made at 1 P.M. in September and October 1930. The data given in Table XV show that their temperature was 3° to 4° C higher than that of the air. In some cases the difference was found to be as high as 10° C.

TABLE XV  
*Mean temperature departures of normal and wilted leaves at 1 P.M.  
 during September and October 1930*  
*Average of 51 determinations*

Nature of leaves	Air temperature in degrees C	Upper surface	Lower surface	Inner tissue	
				Petiole	Midrib
1	2	3	4	5	
Normal	31.3	+1.5 ± 2	+1.1 ± 30	-1.1 ± 25	-1.2 ± 20
Wilted	31.6	+3.9 ± 15	+2.0 ± 20	+3.4 ± 50	+2.0 ± 30
Difference (Wilted - Normal)		+2.4 ± 25 Significant	+1.80 ± 30 Significant	+4.5 ± 47 Significant	+3.2 ± 30 Significant

Wilted leaves show higher temperature than normal leaves and the mean difference is statistically found to be highly significant

### VIII Discussion of the results

(1) The main data are presented in Tables I to XII and Figs. 1 to 9. It will be observed that leaves of the cotton plant behave differently regarding their temperature during different months. In the hot months of July, August and September as shown in Figs. 1, 1 (a) and 2, there is a rise of leaf temperature from 6 A.M. to 8 A.M. or 10 A.M. and afterwards a fall occurs. The fall is evidently due to the cooling of the plant as a result of transpiration, which increases as the day advances, because greater solar radiation promotes transpiration under certain conditions. During the middle of the day, when air temperature is at its maximum and the solar energy incident on the leaves is correspondingly high, leaves of the irrigated crop show a lower temperature by - 2° C to - 3° C due to active transpiration. Some preliminary experiments on the rate of transpiration in cottons show that it is greater during the middle of the day than in the morning and evening.

In the case of the unirrigated crop, it is found that in September and October (1st half) Figs. 6 and 7, the leaves show air temperature in the morning when the atmosphere is cool and transpiration is very little. But as it gets hotter the leaf temperature rises above that of the air and it is 4° C. higher between 12 noon and 3 P.M. In this respect the unirrigated

crop shows a marked difference from the irrigated crop where the plants have a lower temperature than that of the air during these hours (Figs. 2 & 8)

The explanation seems to be that on account of shortage of moisture in the soil and leaves in the unirrigated crop, transpiration is reduced and sufficient cooling effect is not produced

As the weather becomes colder, the differences of monthly mean departures of the two crops become less striking. This is perhaps due to the fact that moisture content of the leaves of irrigated crop also is depleted with advance in age as shown in Table XIII and also partly because temperature of the soil falls (Table XVIII) and the root absorption is reduced. This has been shown by results of unpublished experiments of the senior author on "Effect of various temperatures on root-absorption in cotton".

(2) Table XVI Appendix A indicates a very interesting behaviour of the leaves during different months. In the case of unirrigated crop there is a successive backward shift of the maxima of the mean departure in temperatures from September to November. In September the maxima occurs at 2 P.M. while in October it is at 11 A.M. or 9 A.M., and in November it comes at 9 A.M.

Reverse is the case in the irrigated crop. The shift of the maxima goes forward (Table XVI Appendix A), i.e., from 8 A.M. in August to 10 A.M. in September and 1st part of October. It comes at 3 P.M. in October (2nd half). There is then a backward shift in November when it comes at 1 P.M.

In seeking an explanation for this, it has to be borne in mind that leaf temperature is correlated with transpiration, which depends upon air humidity, leaf moisture, soil moisture and root-absorption

In August, the average humidity (Table XIX Appendix A) during the day is high. The transpiration, therefore, will be low and this causes a rise in the leaf temperature earlier. But in September and October, humidity is lower and thus the relative transpiration is greater than in August; consequently the leaves attain maximum temperature later. In November with increase in humidity accompanied by shortage of soil moisture, etc., the relative transpiration decreases again and the maximum temperature is attained earlier by the leaves

In the case of unirrigated crop as is shown by Table XIII, Fig. 11, the soil moisture decreases from September to November along with leaf moisture. For this reason a decrease of transpiration occurs and the plants tend to attain the maximum temperature earlier.

(3) Statistical studies made on the coefficient of correlations between surface temperatures and temperatures of the inner tissue show that in the

case of the irrigated crop there is a very high positive correlation which remains almost constant (Table XVII Appendix A) from August to October (1st half). The correlation co-efficient becomes very low during 2nd half of October, perhaps due to the upset of water balance in the plants on account of low soil temperatures (Table XVIII Appendix A). When the plants adjust themselves to low water supply from the roots, the coefficient of correlation again becomes very high in November.

The coefficient of correlation is almost constant from September to November in the case of the unirrigated crop. Probably on account of the adjustment of the plants to low water contents, tendency for variation is checked.

(4) The rise of temperature of wilted leaves above that of the air during September and October is probably due to shortage of moisture in the leaves and soil. In this connection the findings of Molisch (1925) and others on the conversion of starch into sugars in wilting leaves may be of considerable significance. Some preliminary work of the authors of this paper shows that the respiratory activity of wilted leaves is less than that of turgid leaves. This might be due to the accumulation of sugars and inadequate moisture content in the wilted leaves. Déhéram and Manquene (1886) as well as Iljin (1922) and Dastur (1925) have shown how greatly the rate of photosynthesis of leaves is affected by their water content. This points out that the leaves of the crop which become flaccid or wilted due to inadequate soil moisture cannot carry on metabolic functions properly. Moreover Iljin (1922) also investigated that leaves which have once become flaccid or wilted and have regained turgidity and normal appearance after adequate supply of water is given, do not attain their original photosynthetic activity. It follows from these experiments that if a plant gets inadequate supply of water it has not only higher temperatures than the air, but also becomes less active for the performance of metabolic functions properly. Nutritive processes, therefore, are liable to receive a set-back as a result of shortage of soil moisture and the recovery from this disability may not take place to the extent of resuming normal condition. This finding has an important bearing on the irrigation of American cottons in canal colonies.

#### *IX Summary*

(1) The paper deals with the determination of temperatures of 4F American cotton plant at Lyallpur. Temperature was taken by means of a thermo-couple apparatus.

(2) Readings were taken after every 2 hours during day and night in August and September and only during the day in July, October and November. The data are given in the form of departures from the air.

(3) It has been shown that the temperature of the cotton plant is never constant. It changes according to the temperature of the surrounding air. A slight change in the temperature of the air brings about a corresponding increase or decrease in the temperature of leaves. From June to September in the morning and evening the plants show an temperature. In the afternoon when the atmosphere is hot, plants are cooler by about  $2^{\circ}\text{C}$  to  $3^{\circ}\text{C}$ . In October and November however, the plants usually remain warmer than the air and this is perhaps associated with decreasing moisture content of the leaves and consequent decrease of transpiration. At night time, the plants had air temperature. The temperature of the plant is not a constant quantity. Unlike warm blooded animals which have a definite limit of body heat in healthy conditions, plants warm up or cool down just as the temperature of the surrounding air rises or falls. Plant is like a cold blooded organism, whose temperature varies with the medium in which it lives.

(4) Soil moisture has been found to exercise a marked influence on the temperature of the leaves. The plants of the unirrigated crop, which were all the time suffering from shortage of soil moisture, showed a higher temperatures by about  $4^{\circ}\text{C}$  to  $5^{\circ}\text{C}$  during September in the afternoon. Irrigated plants at this time were cooler by about  $2^{\circ}\text{C}$  to  $3^{\circ}\text{C}$ .

(5) On the same plant the temperature of young leaves is found to be lower and that of the old leaves higher than that of the air at 1 p.m., but in the morning and evening there is no significant difference.

(6) Wilted leaves have higher temperatures than that of healthy turgid leaves by  $3^{\circ}\text{C}$  to  $4^{\circ}\text{C}$ . In some cases the difference of about  $10^{\circ}\text{C}$  was observed.

(7) The leaves of the unirrigated crop were found to have lower moisture content as compared with irrigated crop. The amount of moisture decreases with increase in age. The figures of variability in moisture content during the day are 22, 15 and 10 in August, September and November respectively. The leaves of the unirrigated crop show less variation in moisture content.

(8) A positive correlation has been found between leaf surface temperatures and that of the inner tissue. In irrigated cotton it varies between  $+0.94 \pm .032$  and  $+0.78 \pm 0.15$ . For unirrigated crop it is  $+0.77 \pm 0.15$ .

APPENDIX A

TABLE XVI

*Shift of the maxima of the mean departures of leaf temperatures from air during different months in 1931*

Month	Temperature of	Time at which maxima occur
<i>Irrigated plot</i>		
August	Surface	8 a.m.
	Inner tissue	8 a.m.
September	Surface	10 a.m.
	Inner tissue	10 a.m.
October (1st half)	Surface	10 a.m.
	Inner tissue	10 a.m.
October (2nd half)	Surface	1 p.m.
	Inner tissue	9 a.m.
November	Surface	1 p.m.
	Inner tissue	1 p.m.
<i>Unirrigated plot</i>		
September	Surface	2 p.m.
	Inner tissue	2 p.m.
October (1st half)	Surface	11 a.m.
	Inner tissue	11 a.m.
October (2nd half)	Surface	9 a.m.
	Inner tissue	9 a.m.
November	Surface	9 a.m.
	Inner tissue	9 a.m.

TABLE XVII

*Coefficient of correlation between surface and inner tissue temperatures during different months in 1931*

Month	Irrigated plot	Unirrigated plot
August	+ .94 ± .039	..
September	+ .75 ± .15	+ .77 ± .15
October (1st half)	+ .604 ± .26	+ .84 ± .69
October (2nd half)	+ .26 ± .06	+ .61 ± .24
November	+ .78 ± .15	+ .69 ± .19

TABLE XVIII

*Average soil temperature during different months in 1931 at 35 cm. depth*

Month	Soil temperature in degrees C.
August	34.05
September	30.88
October (1st half)	29.9
October (2nd half)	23.0
November	18.5

TABLE XIX

*Average humidity during different months in 1931 at 8 A.M.*

Month	Percentage humidity
August	72.8
September	65.3
October	68.0
November	69.0
December	75.0

*Jai Chand Luthra and Indar Singh Chima.* Proc. Ind. Acad. Sci., B, vol. VI, Pt. X/III





*X. References*

1. Balla, W. L., *The cotton plant in Egypt*, 1912, 202, London
2. Camp, A. F., and Walker, M. N., *Tech Bull Florida*, 1927, 189.
3. Clum, H. H., *Amer Jour. Bot.*, 1926, 13, 194-230
4. Dastur, R. H., *Ann Bot.*, 1925, 39
5. Déherans and Manquene, *Compt Rend.*, 1896, 103, 167
6. Eaton, F. M., and Belden, O. G., *Tech Bull U.S. Dept Agri Washington*, 1929, No. 91, p 1
7. Erakuel and Taubenhau, *Sci.*, 1932, 75, No 1945
8. Iljin, *Jahrb Wiss. Bot.*, 1922, 61, 670
9. Miller, E. C., and Saunders, A. R., *Jour. Agri Research*, 1923, 26, 15-43
10. Mollisch, H., *Zeit P Bot.*, 1925, 17, 577
11. Seely, D. A., *Sci. Arts and Letters, Ann Rpt.*, 1917, 19, 167-196
12. Trought, T., *Ind. Jour. Agri Sci.*, 1931, 4, 319
13. Vant Hoff, J. D., *Studies in Chemical Dynamics*, 1896

## THE PHYSIOLOGY OF DIGESTION AND ABSORPTION IN THE CRAB *PARATELPHUSA* (*OZIOTELPHUSA*) *HYDRODROMUS* (HERBST).

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Received August 3, 1937

(Communicated by Dr S G M Ramanujam, M.A., Ph.D., D.L.C.)

### 1. Introduction

THE various aspects of digestion among Crustacea were studied extensively. The most recent paper on the subject is that of C. M. Yonge (1924). Excepting for a few papers on the structure and mechanism of the gastric armature in Decapods, no aspect of digestion was ever studied in any Indian type. In the present paper, the author proposes to deal with the physiology of digestion and absorption and *Paratelphusa* (*Oziotelphusa*) *hydrodromus* (Herbst). This is the common South Indian field crab which lives in deep burrows having a peculiarly oblique descent on the muddy banks of ponds and small canals. It is often covered by patches of muddy colour and remains at the entrance of its burrow protruding its stalked eyes. At one's approach it rapidly crawls away into its retreat.

This has been selected for study on account of the ease with which it is obtained at all times and the convenience with which it is kept under observation, in the laboratory for long periods.

### 2. Material and Methods

The crabs were collected in and around the University area and kept under observation in the laboratory. Some of them were fixed as soon as they were collected from the field while others were fixed at different periods of starvation to study the extent of the period to which fat within the digestive gland can persist. To study the passage of food in the digestive tract the animals were starved to begin with for a period of three days and then fed on bits of flesh stained with carmine and methylene blue and fixed at different intervals. The preliminary starving forces the crabs to feed on the food stained with unpalatable dyes.

In almost all cases the material was fixed in Bouin's fluid. In the case of the digestive gland 30% alcohol containing 5% corrosive sublimate was used as a fixative. In the case of animals which were fed on olive oil

stained with Sudan III, Fleming's solution without acetic acid was used as fixative. When the animals were fed on ferrum oxydatum saccharatum, they were fixed in 95% alcohol containing 5% ammonium sulphide.

In order to section the heavily chitinous regions of the fore-gut, the material was decalcified for three or four days in 70% alcohol containing 2.5% nitric acid and then transferred to a 10% solution of soft soap in which it was kept for four days and embedded in the ordinary way.

In almost all the cases sections were stained in Delafield's haematoxylin and eosin. In a few cases Heidenhain's iron-haematoxylin was employed.

In connection with the observations made on the peristalsis of the mid- and hind-guts, the apparatus suggested by Hogben and Hobson (1924) in their studies on internal secretions was adopted. The mid-gut or the hind-gut was excised from the living animal and the two extremities were ligatured with silk. By means of one end it is fixed in the bath of saline medium (in proportions by volume of 5/8 molar solutions NaCl 200, MgCl<sub>2</sub> 40, KCl 2, CaCl<sub>2</sub> 2, dextrose 3, with Na<sub>2</sub>HPO<sub>4</sub> to pH 7). The other end is connected with the writing lever. Adrenaline and epinephrine in dilutions of 1/80,000 and 1/1,00,00,00 respectively were used for perfusing the isolated regions of the gut.

### 3 Alimentary Canal

#### (a) General description and histology

The alimentary canal falls into three natural divisions — (i) The fore-gut, (ii) The mid-gut and (iii) The hind-gut.

*Fore-gut* — The mouth is situated on the ventral side of the cephalic region between the mandibles and is bound in front by the fleshy labrum and behind by the metastoma. It leads into the fore-gut which is lined throughout by a cuticle which is continuous with the exoskeleton around the mouth. The fore-gut consists of three distinct parts —

(1) *Oesophagus* — The epithelium consists of cells of great length. They are 85  $\mu$  in length in a crab with a carapace of 2.5 inches in breadth and are 3  $\mu$  in width. External to the epithelial layer is a thin chitinous layer of 12  $\mu$  in thickness. This consists of an outer deeply staining layer and an inner structureless layer. There is a distinct basement membrane beneath which is a layer of connective tissue. This is composed of dense reticulate fibres with a number of small nuclei. External to the layer of connective tissue is a layer of circular muscles — the constrictor muscles. The dilator muscles pass through the connective tissue and are attached to the basement membrane.

Lying in the connective tissue are larger numbers of round glands termed the tegumentary glands (Fig 5). Each gland is globular and consists of numerous narrow conical cells. The apex of each cell is directed inwards. In the centre of each mass is a narrow cavity (Cav Fig 5) which is continued into the cavity of the oesophagus by means of an intra-cellular duct (Int D Fig 5). Each cell of the gland is provided with a distinct nucleus and stains deeply. They have a diameter of 20 to 30  $\mu$ . These glands are scattered through the connective tissue of the oesophageal wall and the labrum and the metastoma are also packed with them. They are also present in the hind-gut.

These glands were supposed to have a digestive function and were termed salivary or intestinal glands (Huet, 1883), (Vitzou, 1882). They secrete a sticky substance which entangles the food in the gut. The secretion does not reveal the presence of any digestive enzyme. As Yonge (1921) has pointed out their presence only in the fore-gut and hind-gut always in connection with chitin suggests the possibility of their rôle in the secretion and preservation of the chitinous lining.

(2) *Cardiac fore-gut*—The oesophagus runs directly upwards and opens into the spacious cardiac fore-gut (C Fig 1). This is a large spherical sac whose chitinous lining is thickened at definite places to form a complicated arrangement of ossicles termed the gastric armature which will be described below. At the posterior wall of the cardiac fore-gut is a ventral invagination termed the cardio-pyloric valve separating the cardiac chamber from the pyloric chamber.

(3) *Pyloric fore-gut*—The cardiac chamber passes downwards into the pyloric chamber (Py Fig 1). This is simple towards its anterior third. In the posterior two-thirds its ventral wall is thickened to form two rounded lateral pouches the "ampoules pyloriques" of Mocquard (1883). These constitute the "gland filter" of Yonge (1924). Each ampulla is thrown into distinct longitudinal ridges beset with stiff setae. The two ampullae meet in the mid-ventral line forming the inter-ampullary ridge (I Amp Fig 2). The ventro-lateral walls of the chamber are also thickened forming the supra-ampullary ridges (S Amp Fig 2). The approximation of the supra-ampullary ridges divides the pyloric chamber into a dorsal comparatively free region and a ventral portion (V D Fig 2) which is further subdivided into a left and a right portion by the presence of the inter-ampullary ridge. Detailed account of the structure and mechanism of the gland filter is found in the works of Jordan (1904), Williams (1907) and Yonge (1924). The cardiac and pyloric chambers have the same histological details as the oesophagus. There is a columnar epithelium and the chitinous layer is thin.

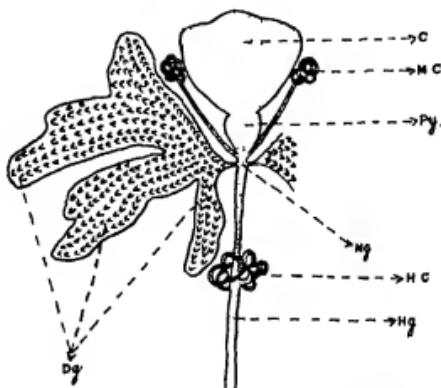


Fig 1.

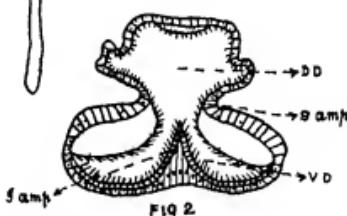


Fig 2.

in places other than those occupied by the masticatory ossicles. There is a distinct basement membrane, and a thin layer of connective tissue with circular and longitudinal muscles.

*Mid-gut*—Posteriorly the pyloric chamber passes into the mid-gut. From the upper side of the posterior end of the pyloric chamber six to seven flap-like structures or valves project into the mid-gut. The views of Cuenot (1893), Mocquard (1883) and Huxley (1880) regarding the physiological significance of these structures are discussed elsewhere in this paper.

From the anterior region of the mid-gut two cæca, the mid-gut cæca arise one from each side. Each cæcum extends over the pyloric chamber and is pressed against the side of the cardiac chamber. The digestive gland also opens into the mid-gut just behind the mid-gut cæca (Fig 1).

The mid-gut is the shortest portion of the alimentary canal and is about 12-13 mm in a full-grown crab (Fig 1). The epithelial cells (Fig 3) are

columnar and are  $50\mu$  in length with small nuclei. They are provided with a striated border with a thickness of 1 to  $2\mu$ . Near the base of the epithelium there are the basal cells (B C Fig 3) of Frenzel (1885) with deeply staining large nuclei. Beneath the epithelium is a basement membrane and beneath this is a layer of connective tissue with circular and longitudinal muscles.

The mid-gut caeca have the same histology as the mid-gut.

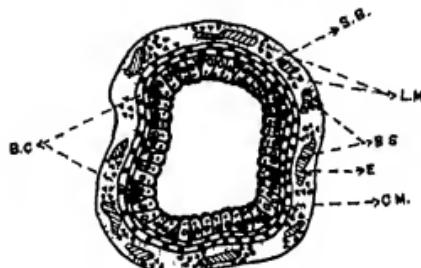


Fig 3

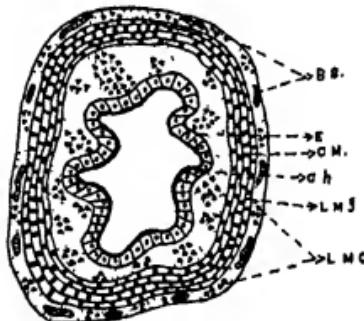


Fig 4

*Hind-gut*—The hind-gut is a long narrow tube which starts from the mid-gut and extends as far as the anus. A long coiled cecum (H C Fig 1), the hind-gut cecum, is given off from the right side of the hind-gut before its entry into the abdomen. The hind-gut and hind-gut cecum have columnar cells of  $60\mu$  in length and have towards their inside a deeply staining

thin chitinous layer (Ch Fig 4) which is produced into longitudinal ridges. The chitinous layer is continuous with the exoskeleton at the anus. The walls of the hind-gut as in the case of the oesophagus are packed with tegumental glands which are found in great profusion towards the anterior region of the hind-gut. The circular and longitudinal musculature (C M and L M I and L M O Fig 4) is better developed in the hind-gut than in the mid-gut.

(b) *Digestive gland*

Almost the entire ventral side of the anterior region of the cephalothorax is occupied by the paired digestive gland or hepatopancreas. This is a yellowish brown lobulated structure. Each half of the gland is composed of three main lobes. There are three main ducts arising from them on each side corresponding to the three lobes. These three finally end in fine blind tubules and the cavity of each tubule is in communication with the mid-gut as in the case of *Cancer*.

In section (Fig 6) the tubules are bounded by connective tissue and are composed of two types of cells beneath a basement membrane. (i) Ferment or secretory cells vary in height from 20 to  $70\mu$  and  $60\mu$  in width (S C Fig 6). Each cell contains small vacuoles and a darkly staining secretion. Some of the cells are packed with secretion while others are found having discharged partly or completely their secretion into the lumen of the tubules.

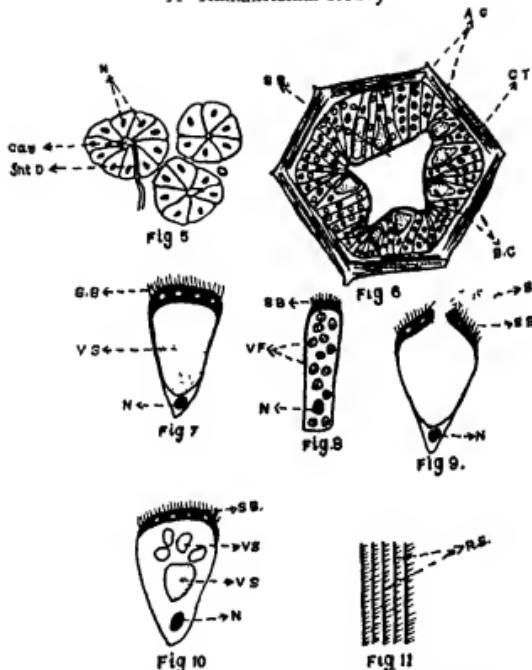
(ii) Absorption cells (A C Fig 6) are vacuolate and vary from  $70$  to  $100\mu$  in length and  $20\mu$  in width. Fat globules are found within each cell. The border of the cell in contact with the lumen of the tubule both in the case of ferment as well as absorption cells is striated.

(c) *Ossicles of the fore-gut*

The ossicles of the fore-gut fall into two distinct sets (i) those of the cardiac chamber and (ii) those of the pyloric chamber. The ossicles of each chamber can be further grouped into (i) main ossicles and (ii) supporting ossicles.

*Main ossicles*—The meso-cardiac ossicle (M Fig 12, A) is a triangular piece of sclerite transversely placed in the anterior region of the dorsal wall of the cardiac chamber. The anterior end is ventrally inclined while the posterior broader end is not distinctly separated from the uro-cardiac ossicle (U Fig 12, A) and the ptero-cardiac ossicles (Pt Fig 12, A).

The uro-cardiac ossicle (U Fig 12, A) is attached to the hind end of the meso-cardiac ossicle and extends posteriorly along the dorso-median



line. Posteriorly, it inclines ventrally and gives articulation to the pro-  
pylionic ossicle (*P P* Fig 12, C). Ventrally is the median tooth (*M D* Fig 12, A) which is differentiated into three regions. There is an anterior  
crescentic denticle and a broader posterior denticle with a concavity  
between them. In front of the anterior denticle is a semi-circular depression  
in which a minute denticle is present.

A pair of ptero-cardiac ossicles (*Pt* Fig 12, A) are attached to the  
meso-cardiac ossicle on either side by means of oblique hinges. Their  
inner ends which are in contact with the meso-cardiac ossicle are broader  
while their outer ends are narrow. The anterior gastric muscles are attached  
to the inner ends of the ptero-cardiac ossicles. The outer ends of the ossicle  
articulate with the zygo-cardiac ossicles (*Z* Fig 12, B) by means of a  
ligament—the antero-lateral ligament of Pearson (1908). The posterior border

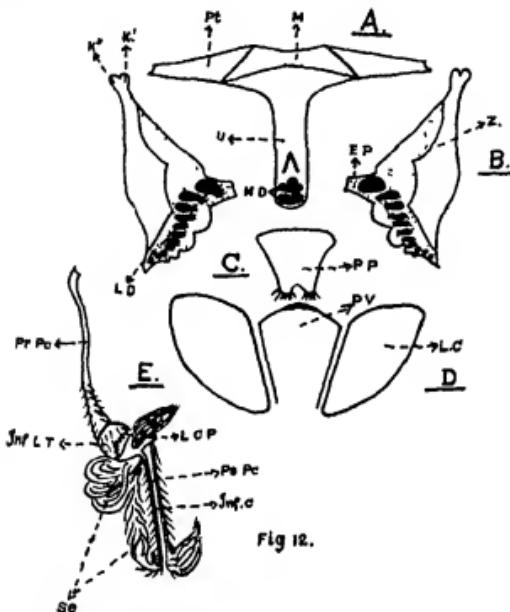


Fig 12.

of the ptero-cardiac ossicle is straight while the anterior has a somewhat curved margin

The pair of lateral zygo-cardiac ossicles (Z Fig 12, B) articulate with the outer ends of the ptero-cardiac ossicles on either side. They pass posteriorly and exhibit a ventral inclination. Each zygo-cardiac ossicle anteriorly is produced into two knob-like processes (K', K'' Fig 12, B). The inner process articulates with the outer end of the ptero-cardiac ossicle of its side. The outer process is attached to the anterior end of the prepectinal ossicle one of the supporting ossicles described below of its side. Anteriorly each zygo-cardiac ossicle is narrow, and rod-like and becomes broadened towards the hind end. About the level of the median tooth the lateral teeth (LD Fig 12, B) are developed on them. Posteriorly, the dorsal border is in connection with the cardio-pyloric valve (PV Fig 12, D) and the exo-pyloric ossicles (EP Fig 12, B). The inner border of the

ossicle is curved ventrally whereas the outer margin is bent inwards so that the ossicle exhibits a cavity towards the inside and a convexity towards the outside. The outer border is folded forming a deep groove below the outer convexity. Though the ossicle is thin, yet as a result of the complicated folding it gives a false impression of thickness.

Each lateral tooth (*L D* Fig. 12, B) of the zygo-cardiac ossicle carries eight to nine hard denticles separated by deep grooves. The denticles are differentiated into three to four definite regions. The nature of the denticles in each region differs from those in other regions.

The posterior border of the pro-pyloric (*P.P.* Fig. 12, C) is rotated to the anterior side so that its position at rest resembles the position of the pro-pyloric ossicle of *Cancer pagurus* (Pearson, 1908). The exo-pyloric ossicles are attached to the posterior border and therefore displaced from the position described in *Gelasimus* (Reddy, 1934). This is provided with setæ and is depressed posteriorly. In these depressions the cardio-pyloric constrictors are inserted. The posterior end is bifurcated while the narrower anterior end is attached to the uro-cardiac ossicle.

The pyloric ossicle is a broad dorso-median ossicle in the roof of the pyloric chamber. Its anterior end is in contact with the pro-pyloric ossicle.

*The supporting ossicles*—The pyloric supporting ossicles do not differ from those of *Cancer* described by Pearson (1908). The cardiac supporting ossicles however differ.

The anterior ends of the pre-pectinal ossicles (*Pr P* Fig. 12, E) articulate with the outer knob-like processes (*K'* and *K''* Fig. 12, B) of the zygo-cardiac ossicles. They closely follow the zygo-cardiac ossicles and gradually get broadened at their attachment with the pectinal ossicles. The pectinal ossicles (*Inf L T* Fig. 12, E) are cup-shaped and bear towards their inside the infra-lateral-cardiac teeth which are in a level with the lateral teeth on either side of the median tooth (*M D* Fig. 12, A). Laterally, the pectinal ossicles are attached to the lateral cardio-pyloric ossicles (*L C P* Fig. 12, E) and posteriorly to the infra-lateral cardiac and post-pectinal ossicles (*Inf* *s*, *Ps P* Fig. 12, E). These are thickly beset with setæ. The infra-lateral cardiac ossicles end in oval setose brushes towards the oesophagus on reaching the same level as the post-pectinal ossicles turn up and end in posteriorly directed long tufts of setæ (*Se* Fig. 12, E) which extend as far as the cardio-pyloric entrance.

(d) *Muscles of the fore-gut.*

The musculature falls into two types, viz., the extrinsic and intrinsic muscles. {

*Extrinsic musculature* — The muscles in this system run from the ossicles of the gastric armature to the exo-skeleton. The anterior gastric muscles extend between the inner ends of the ptero-cardiac ossicles and the anterior region of the carapace. They are in the form of two distinct bands which diverge considerably anteriorly.

There are three bands of posterior gastric muscles. The inner or the central band arises from the posterior region of the carapace and is inserted on the pyloric ossicle. The outer bands also arise from the posterior region of the carapace and are inserted on the dorsal surface of the posterior ends of the zygo-cardiac ossicles.

There are two bands of cardiac levator muscles extending between the lateral walls of the cardiac chamber and the carapace. The pyloric levator muscles extend between the lateral walls of the pyloric chamber and the posterior region of the carapace. Two bands of cardiac depressor muscles extend between the ventro-lateral wall of the cardiac chamber and the anterior region of the sternum. The pyloric depressor muscles proceed ventrally from the ventro-lateral wall of the pyloric chamber.

*Intrinsic musculature* — In this case the muscles run between the ossicles of the armature within the fore-gut itself.

The cardio-pyloric constrictors arise from the posterior border of the meso-cardiac ossicle and are inserted on the posterior border of the propyloric ossicle. They are in three bands, the outer of which diverge slightly towards their insertion. There are three pairs of lateral cardiac muscles. The dorsal pair extends between the dorsal border of the zygo-cardiac ossicle and the upper region of the infra-cardiac ossicle and the pre-pectinal ossicle. The central one extends between the upper region of the infra-cardiac ossicle and the pre-pectinal ossicle. The ventral one runs from the lateral surface of the infra-lateral cardiac ossicle to the dorsal surface of the postero-lateral cardiac plate.

The cardiac constrictors are formed by two sets of muscles the postero-inferior cardiac and antero-lateral cardiac muscles. The postero-inferior cardiac muscles run between the two posterior ends of the infra-lateral cardiac ossicles. The antero-lateral cardiac muscles arise as a bifurcated bend from the antero-lateral cardiac plate and are inserted on the anterior wall of the meso-cardiac ossicle just above the insertion of the cardio-pyloric constrictor muscles. The cardiac depressors and elevators are on either side of the cardiac constrictors.

The pyloric constrictor muscles are constituted by a number of narrow bands between the post-pectinal and infra-lateral cardiac ossicles and the supporting ossicles in dorsal and lateral walls of the pyloric chamber.

(e) *Modus Operandi of the gastric armature*

Mocquard (1883), Pearson (1908) and Patwardhan (1934-35) have pointed out that the gastric armature is put into action mainly by the contraction of the anterior gastric muscles. Huxley (1880) described the active movement as the result of contraction of both anterior as well as posterior gastric muscles. But a study of the gastric armature of *Paratelphusa* and other South Indian Decapod Crustacea shows that the active movement is brought about mainly by the contraction of the posterior gastric muscles while the anterior gastric muscles and cardio-pyloric constrictors are chiefly concerned in the restoration of the armature to its position of rest.

If the anterior gastric muscles were mainly responsible for the collision of the three teeth-bearing ossicles, namely, uro-cardiac and the two zygo-cardiac ossicles, as stated by Mocquard (1883), Pearson (1908) and Patwardhan (1934-35) the most important factor, viz., the pressing down of the uro-cardiac tooth to meet the colliding zygo-cardiac teeth could not be brought about on account of the reverted position of the propyloric ossicle. It is only when the posterior border of the pro-pyloric ossicle is pulled backwards the uro-cardiac tooth is brought downwards. This is made possible only by the contraction of the posterior gastric muscles. Their contraction results in pulling back the pro-pyloric ossicle which owing to the roof of the cardiac chamber presses down the uro-cardiac tooth to meet the zygo-cardiac teeth.

Both the uro-cardiac and propyloric ossicles represent levers of the second order. In the case of the uro-cardiac ossicle the fulcrum is situated at its attachment with the meso-cardiac ossicle while the power is applied at its posterior extremity just behind the uro-cardiac tooth by the anterior border of the pro-pyloric ossicle. The power is the result of the contraction of the posterior gastric muscles and is transferred to that point by means of the exo-pyloric and pro-pyloric ossicles. The work is performed in the region of the uro-cardiac teeth.

In the case of the pro-pyloric ossicle the fulcrum is at the attachment of its anterior border with the posterior end of the uro-cardiac ossicle and the power is applied by the exo-pyloric ossicles at its posterior border while work is done at a point near the fulcrum in pressing down the uro-cardiac tooth.

On the supposition that the operation of the gastric armature is effected by the contraction of the anterior gastric muscles, Pearson (1908) locates both the point of application of power and fulcrum at the place of articulation of the zygo-cardiac ossicle with the outer end of the ptero-cardiac

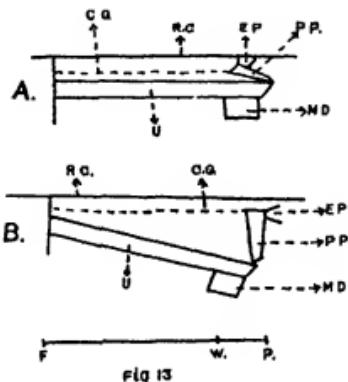


Fig 13



Fig 14

ossicle while the work is turned out at the zygo-cardiac tooth and describes the action of the zygo-cardiac tooth as that of the lever of the second order, considering the zygo-cardiac and exo-pyloric ossicles as a single bar. With the conditions described by him neither the zygo-cardiac ossicle with the exo-pyloric is a lever of the second order, nor is movement possible when power acts at the fulcrum. But if the power were to act at the exo-pyloric ossicles—as it should by the contraction of the posterior gastric muscles—then the action of the combined ossicles is that of a lever of the second order with the fulcrum at the anterior end of the zygo-cardiac ossicle, the work being turned out in the region of the zygo-cardiac tooth and the power being applied at the exo-pyloric ossicles as shown in Fig 15

The pre-pectinal ossicles bearing the infra-lateral cardiac teeth do not seem to come into action during the play of the main ossicles. Their action commences just after the cessation of the activity of the main ossicles. When the armature is in its position of rest, the postero-inferior cardiac

muscles which are attached to the posterior borders of the infra-lateral cardiac ossicles contract. As the post-pectinal ossicles are fused with the infra-lateral cardiac ossicle, both at the posterior and anterior ends, the contraction of these muscles draw together the pectinal ossicles, so that the infra-lateral cardiac teeth meet in the middle line above the median tooth, brushing it as they come together. These clean the furrows of the median tooth and thus serve to keep it clean for the next collision with the zygo-cardiac teeth. The infra-lateral cardiac teeth are brought back to their original position by the contraction of the lateral cardiac systems of muscle which extend between the dorsal border of the zygo-cardiac ossicles and the anterior region of the infra-lateral cardiac ossicles. If we consider

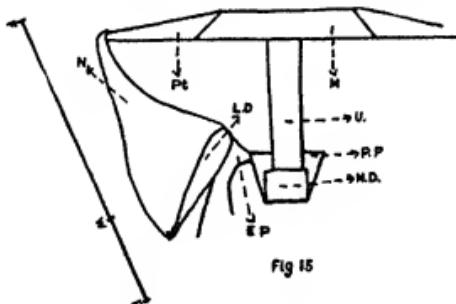


Fig 15

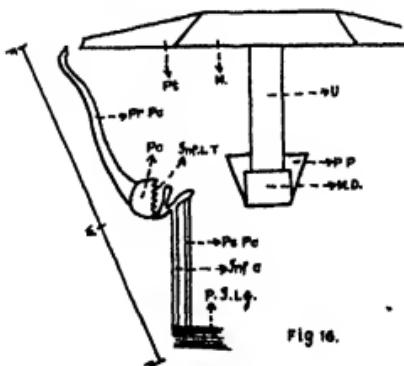


Fig 16.

the pectinal and infra-lateral cardiac ossicles as a single piece, their action is that of a lever of the second order. The fulcrum is at the anterior extremity of the prepectinal ossicle at its articulation with the outer end of the ptero-cardiac ossicle and the work is turned out in the region of the infra-lateral cardiac teeth, while the power is applied by the postero-inferior cardiac muscles at the hind ends of the ossicle.

Thus the entire operation of the gastric armature falls into two types, *viz.*, (i) the action of the main system of ossicles and (ii) the action of the supporting system of ossicles. The former is concerned with the crushing of the hard food materials while the latter is concerned with the cleaning of the crushed material from the furrows of the teeth borne by the main ossicles. The two actions alternate. The first is brought about by the extrinsic musculature while the second is effected by the intrinsic system.

#### *4. Passage of the Food through the Digestive Tract*

The animals after a preliminary starving were fed on pieces of flesh stained with methylene blue and carmine. The animals were killed at intervals of two to three hours each and the various regions of the alimentary canal were fixed in Bouin's fluid.

Food is held by the chelic and passed to the maxillipedes which after preliminary reduction transfer it to the chitinised mandibles. After further reduction it is passed on into the mouth. Through oesophagus it is passed into the cardiac fore-gut by the action of oesophageal musculature.

No trace of the stains used with the food was detected in the mid-gut or the tubules of the digestive gland in the first four hours. During this time the food is retained in the fore-gut and is subjected to the grinding action of the gastric armature and the straining processes of the gland filter. The digestive secretion from the digestive glands makes its way into the cardiac chamber along the ventral grooves (*V D* Fig. 2) of the gland-filter and mixes with the fine particles of food material as shown by Yonge (1924). The fluid with the digested food in very fine particles then passes back through the gland-filter into the mid-gut. All large particles are retained in the cardiac fore-gut till they are reduced to a fine state. After ten hours methylene blue and carmine can be detected in the mid-gut, mid-gut caeca and the tubules of the digestive gland. The absorption of the digested matter takes place within the mid-gut and its connected structures. The undigested matter is forced into the hind-gut by the peristaltic movements of the mid-gut. These peristaltic movements are not very pronounced. It takes about 1.5 to 2 minutes for the completion of every peristaltic wave. When the isolated mid-gut in a bath of saline

medium was perfused with adrenaline and epinephrine in dilutions mentioned before, the peristaltic movement was remarkably accelerated. It took about a minute for the action of the adrenaline to start. 7 to 8 peristaltic waves per minute were produced.

In about 12 to 16 hours, the undigested matter enters the hind-gut and is finally expelled through the anus. The stains used with the feeding of the animals are detected in all regions of the hind-gut during this time.

The peristaltic movements of the hind-gut are more pronounced than those of the mid-gut. Four to six peristaltic waves per minute are noticed. Every peristaltic wave starts from the anterior end of the hind-gut, quite independent of the peristaltic waves of the mid-gut and passes towards the anus which is extended with every wave.

Perfusion of the isolated hind-gut with adrenaline and epinephrine in dilutions mentioned above accelerates the peristaltic rhythm.

Sometimes a few unreduced hard pieces escape the cardio-pyloric valve and are found in the spacious anterior third of the pyloric chamber. The concrecence of the supra-ampullary ridges effects a complete separation of the dorsal and the ventral regions of the posterior two-thirds of the pyloric chamber. Due to the resistance offered by the profuse setae of the gland filter in the ventral division, the unreduced pieces which have escaped the cardio-pyloric valve are forced into the dorsal division whereas the digestive secretion with dissolved food and very fine particles alone pass through the gland-filter of the ventral division. Mention has already been made about five to six flap-like processes projecting backwards from the posterior end of the pyloric chamber into the mid-gut. Huxley (1880) and Mocquard (1883) suggest that these act as valves in preventing regurgitation of the contents of the mid-gut into the fore-gut. Pearson (1908) and Cuenot (1893) state that these are concerned in carrying the hard pieces from the dorsal division directly into the hind-gut, thus preventing the soft walls of the mid-gut from damage. Yonge (1924) agrees with Huxley (1880) and Mocquard (1883). In *Nephrops* which has a long mid-gut such a direct carriage of particles from fore-gut to hind-gut by the flap-like processes is not possible. But in *Paratelphusa* the mid-gut is very short and the valves in all probability are concerned in carrying these hard pieces directly from the fore- to the hind-gut.

### 5 Specificity of Digestive Enzymes

The digestive secretion produced by the secretory or ferment cells of the digestive gland is a thick yellowish brown fluid. This makes its way into the cardiac chamber through the grooves in the gland-filter and is

mixed up with the food. The secretion is easily obtained by pushing a glass tube into the cardiac fore-gut through the mouth and drawing off the fluid therein. For experimental purposes glycerine extracts of the digestive glands were prepared. Only extracts of 20% strength were used in the reactions. Boiled extracts were set up as controls. Toluene was used as an antiseptic and the digests were incubated at 36° C.

The digestive secretion is poured into the lumen of the tubules of the gland in the form of fine yellowish droplets. Sections of the gland reveal some of the secretory cells actually emptying their contents into the lumen of the tubule. The secretion is faintly acidic towards litmus. As *Nephrops* (Yonge, 1924) it shows no trace of free acids. The secretion contains amylolytic, proteolytic and lipolytic enzymes.

The main results of the experiments connected with the amylolytic enzyme are shown in Table I. Starch, glycogen, sucrose are digested. It has no action on inulin and raffinose as in the case of *Nephrops* (Yonge,

TABLE I

No.	Experiment	Time	Result
1	(A) 15 c.c. of 20% extract + 15 c.c. of 1% Starch medium, neutral	5 hours	Titrated into 10 c.c. of Benedict's solution (A) 17.5 c.c.
	(B) .. medium 2 N HCl	..	(B) 38.0 c.c.
	(C) .. medium 2 N Na <sub>2</sub> CO <sub>3</sub>	..	(C) 42.0 c.c.
2	(A) 15 c.c. of 20% extract + 15 c.c. of 5% glycogen	3 hours	Titrated into 5 c.c. of Benedict's solution (A) 5.5 c.c. (B) 17.0 c.c.
	(B) .. Boiled	..	
3	(A) 15 c.c. of 20% extract + 15 c.c. of sucrose	..	(A) 5.0 c.c.
	(B) .. Boiled	..	(B) 17.5 c.c.
4	(A) 15 c.c. of 20% extract + 15 c.c. of 1% inulin	..	(A) 14.0 c.c.
	(B) .. Boiled	..	(B) 14.0 c.c.
5	(A) 15 c.c. of 20% extract + 15 c.c. of 1% raffinose	..	(A) 16.0 c.c.
	(B) .. Boiled	..	(B) 10.0 c.c.
6	(A) 20 c.c. of 20% extract + 1 gm saw dust	8 days	(A) 7.4 c.c.
	(B) .. Boiled	..	(B) 36.9 c.c.

1924) But traces of cytase as in *Astacus* (Bidermann and Moritz, 1898) are revealed by the action of the enzyme on saw-dust which contains hemi-celluloses.

The optimum temperature for the action of the enzyme was then determined. The results of the experiments are shown in Table II and are represented graphically in Fig. 17. The optimum lies at 45° C. Similarly,

TABLE II

No	Experiment	Tempera- ture	Result
A	10 c.c. of 20% extract + 10 c.c. of 2% starch solution was incubated for 6 hours at	30° C	At the end of 6 hours each digest was boiled, filtered, made up to 20 c.c. and titrated into 15 c.c. of Benedict's solution 10.2 c.c.
B	do	35° C	10.1 c.c.
C	do	40° C	10.0 c.c.
D	do	42° C	9.9 c.c.
E	do	44° C	9.8 c.c.
F	do	46° C	9.8 c.c.
G	do	48° C	10.0 c.c.
H	do	50° C	10.2 c.c.
I	do	52° C	10.3 c.c.

TABLE III

No	Experiment	Tempera- ture	Result
A	10 c.c. of 20% extract was kept for 30 minutes at	54° C	10 c.c. of 2% starch was added and incubated at 30° C for 3 days. Then boiled, filtered, made up to the same volume and titrated into 15 c.c. of Benedict's solution 10.4 c.c.
B	do.	55° C	10.4 c.c.
C	do	56° C	10.8 c.c.
D	do	58° C	11.4 c.c.
E	do.	62° C	12.2 c.c.
F	do.	64° C.	16.0 c.c.
G	do.	66° C.	16.0 c.c.
H	do.	68° C.	16.0 c.c.

the temperature of destruction was also determined. The results are shown in Table III. The temperature of destruction lies at 62° C. It is interesting

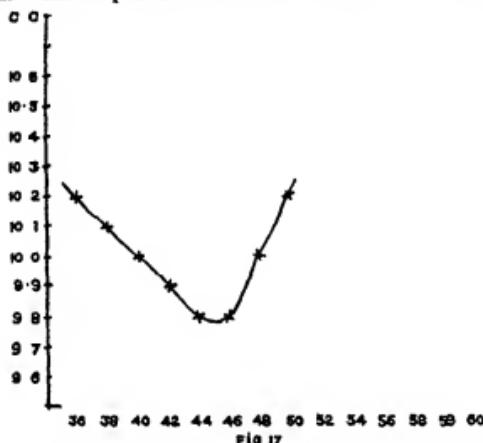


Fig. 17

to compare these results with those obtained by Yonge (1924) in the case of *Nephrops* *Paratelphusa* livers under warmer conditions than *Nephrops* and still the optimum temperature and temperature of destruction of the amylolytic enzyme are lower than those of *Nephrops* (Yonge, 1924).

The proteolytic enzyme present is most active in alkaline medium and is almost inhibited in acid medium. Fibrin, casein and peptone are digested. The optimum degree of alkalinity for the action of the enzyme was then determined. The method of Yonge (1924) was adopted. Digests with various degrees of alkalinity were set up with .5 g of fibrin in each. They were incubated at 36° C for 3 days and then boiled, filtered and made up to the same volume. 20 c.c. of 10% formaldehyde was added to each and titrated into N/10  $\text{Na}_2\text{CO}_3$  with phenolphthalein as indicator. The following results were obtained:

Neutral . . .	5.4 c.c.
N/80 $\text{Na}_2\text{CO}_3$ . . .	6.8 c.c.
N/40 . . .	7.1 c.c.
N/30 . . .	7.2 c.c.
N/20 . . .	8.9 c.c.
N/10 . . .	8.9 c.c.

The optimum alkalinity as in the case of *Carcinus* (Roaf, 1908) and *Nephrops* (Yonge, 1924) is in N/20  $\text{Na}_2\text{CO}_3$  medium

The lipolytic enzyme shows a wide range of action. It hydrolyses methyl acetate, amyl acetate, butyl acetate, ethyl acetate and an emulsion of olive oil

The digestive secretion also hydrolyses amygdalin and salicin. Phloridzin was not acted upon

The digestive gland in addition to the functions of digestion and absorption according to Cuenot (1893) has also excretion, elimination and regulation. Carnine and methylene blue which were injected into the abdomen were detected in the tubules of the digestive gland. This phenomenon as Yonge (1924) has already pointed out, is of little importance in the excretory process

#### 6 Nature of Absorption and Food Reserves

For investigating the absorption within the gut the animals were first starved for three days and then some were fed on olive oil stained with Sudan III while others were fed on ferrous oxydatum saccharatum. Those fed on olive oil were used for detecting absorption within mid-gut and mid-gut caeca. These were fixed in Fleming's solution without acetic acid. In the case of the tubules of the digestive gland where there is a lot of stored fat the olive oil process is of no use. Experiments on starving and subsequent sectioning of the gut have revealed that fat can persist in the cells of the digestive gland up to 38 to 40 days. The absorption cells at this stage are clear and in many cases distorted. For detecting absorption in the tubules of the digestive gland animals fed on iron salts were fixed in ammonium sulphide in 96% alcohol.

The absorptive cells of the mid-gut and mid-gut caeca showed the presence of dark-staining fat droplets. No trace of absorption was noticed in the fore-gut, hind-gut and hind-gut caecum. The ferment or secretory cells of the mid-gut are free from these darkly staining globules.

In sections of the digestive gland iron salts are found profusely in the lumen of the tubules and in the vacuoles in the absorptive cells of the tubules. The presence of iron salts in the absorptive cells of the mid-gut and mid-gut caeca is easily noticed.

Cuenot (1893) and Jordan (1904) considered that the mid-gut and the mid-gut caeca are specialized for fat absorption. There is however no reason for that assumption. Animals were starved up to 40 days till almost all the fat in the absorptive cells disappeared. They were then fed on olive oil and after 4 days fixed in Fleming's solution without acetic acid. Section

of both the digestive gland as well as those of the mid-gut and mid-gut cæca revealed the presence of darkly staining globules. Thus clearly show that absorptive cells of digestive gland tubules also are capable of absorbing fat. These, however, were in greater profusion in the mid-gut and mid-gut cæca. This is evidently due to the fact—as Yonge (1921) has pointed out—that carbohydrates and proteins which are easily split up make their way immediately into the tubules of the digestive gland, from the fore-gut, whereas fats which are more slowly split up are passed on into the mid-gut.

Though the mid-gut is very small the absorptive surface is very much increased by the lumen of the two long mid-gut cæca and the tubules of the digestive gland.

Within the absorptive cells of the digestive gland fats, glycogen and calcium salts are found as reserves. Bernard (1853), Smith (1914), Kirch (quoted by Yonge, 1924), Paul and Sharpe (1919) have shown the close connection of these food reserves with the moulting in the case of many other Decapods.

#### 7 Summary and Conclusion

1. *Paratelphusa (Ornatotelphusa) hydromedus (Herbst)* is the common South Indian field crab. It is a fresh-water animal which lives in deep peculiarly oblique burrows.

2. The alimentary canal falls into three natural divisions, viz., (i) fore-gut, (ii) mid-gut and (iii) hind-gut. The fore-gut consists of the oesophagus, cardiac chamber and the pyloric chamber. The mid-gut is the shortest portion of the gut. Immediately behind the fore-gut it is produced into two cæca. The digestive gland opens into the mid-gut at this region by two lateral ducts. The hind-gut is the longest portion. Anteriorly arising from the right side is a cæcum. The histological structure of the various regions is described.

3. The gastric armature present in the fore-gut is worked by a number of muscles. Its mechanical constitution is such that the force of a single pull is resolved along three teeth-bearing ossicles which collide. The force of collision is so great as to reduce even the hard shells of molluscs to a fine powder.

4. The *modus operandi* of the gastric armature is brought about by the posterior gastric muscles. The cardio-pyloric constrictors and the anterior gastric muscles are concerned in the restoration of the gastric armature to its position of rest. The various views regarding the working of the different ossicles are discussed.

5 After external reduction by the mandibles food is passed into the fore-gut where it is retained for four hours. Here it is subjected to the elaborate processes of mastication and straining by the gastric armature and glandfilter. The digestive secretion from the digestive glands finds its way into the cardiac chamber and mixes with the reduced food. In about ten hours the food mixed up with the secretion passes into the mid-gut and its connected structures. In 12-16 hours it reaches the hind-gut and is finally expelled through the anus. Peristaltic movements of the mid- and hind-guts are responsible for the passage of food in the gut.

6 The mid-gut exhibits feeble peristalsis, each peristaltic wave taking 1.5 to 2 minutes for its completion. The hind-gut shows pronounced peristalsis—4 to 6 waves occur per minute. When perfused with adrenalin and epinephrine in dilutions of 1/80,000 and 1/1,00,000 respectively, the peristaltic rhythm in both the cases was very much accelerated.

7 Amylolytic, proteolytic and lipolytic enzymes are present in the digestive secretion. Amylolytic enzyme acts best in neutral medium and digests starch, glycogen, sucrose. Its optimum temperature is 45°C and temperature of destruction is 62°C. Hemucelluloses in saw-dust were also acted upon showing the presence of a cytase. Proteolytic enzyme has an optimum in N/20 Na<sub>2</sub>CO<sub>3</sub> medium. Lipolytic enzyme has a very wide range of action. It hydrolyses fats and esters.

8 Absorption is confined to the mid-gut, mid-gut cæca and tubules of the digestive gland. It is observed that fat is absorbed both in the mid-gut and the digestive tubules. There is no evidence to show the assumption of Cuenot (1893) and Jordan (1904) regarding the specialisation of the mid-gut for fat absorption.

9 Fats, glycogen and calcium salts are found as reserves in the cells of the digestive glands.

In conclusion, I have great pleasure in expressing my best thanks to Mr K V Reddy, B.A., B.Sc. (Edin.), Forest Officer, Jeypore, Mr R V Seshaiya, M.A., Lecturer in Zoology, Annamalai University, and to Dr B B Dey, D.Sc. (Lond.), F.I.C., and Dr S G M Ramanujam, M.A., Ph.D. (Lond.), F.Z.S., Professors in Chemistry and Zoology, respectively in the Presidency College, Madras, for their valuable suggestions, encouragement and help during the course of this research.

REFERENCES

1. Bernard, C., "Recherches sur une nouvelle fonction du foie," *Ann de Sc Nat, Zool.* (3rd ser.), 1853, 19
2. Biemann, W., und Moritz, "Beitrage zur vergleichenden Physiologie der Verdauung, I.—Über ein cellulose lösendes Enzym im Lebersekret der Schnecken," *Arch ges Physiol.*, 1898, 73
3. Calman, W. T., *A Treatise on Zoology*, edited by Ray Lankester, 1909, Part VII, 3rd fascicle, "Crustacei," London
4. Cuenot, L., "Etudes physiologiques sur les Crustaces Decopodes," *Arch de Biol.*, 1893, 13
5. Elliott, "Action of Adrenaline," *Jour Physiol.*, 1904, 32
6. Frenzel, "Über den Darmkanal der Crustaceen nebst Bemerkungen zur Epithelregeneration," *Arch Mkr Anat.*, 1885, 25
7. Gaskell, "Adrenaline in Annelids," *Jour Gen Physiol.*, 1919, 2
8. Gaja, J., "Ferments des Glucosides et des Hydrates de Carbone chez les Crustaces Marins," *C.R. Soc. Biol.*, 1907, 63
9. Huet, L., "Nouvelles recherches sur les Crustaces Isopodes," *Jour de l'Anat et de la Physiol.*, 1883, 19
10. Hogben and Hohson, "Studies on internal secretions III. Action of Pituitary extract and adrenaline on Contractile tissues of certain invertebrates," *Brit J Exper Biol* 1924, 1, (2)
11. Huxley, T. H., *The Crayfish*, London. International Scientific Series, 1880
12. Jordan, H., "Die Verdauung und der Verdauungsapparat des Flusskrebses," *Arch ges Physiol.*, 1904, 101
13. Jordan, H., "Zur Prage nach der exkretiven Funktion der Mitteldarmdruse bei *Astacus fluviatilis*," *op. cit.*, 1904, 105
14. Lang, A., *Text-book of Comparative Anatomy*, London, 1896
15. Miller, F. R., "On the Rhythmic Contractility of the Annel Musculature of the Crayfish and Lobster," *J Physiol.*, 1910, 40
16. Mocquard, A., "Rechercher anat. sur l'estomac des Crustaces podophthalmaires," *Ann Sc Nat.*, 1883, (6), 16
17. Murtin, J. R., "Absorption and secretion in the Digestive system of the Land Isopodes," *Proc Acad Nat Sci Phil.*, 1902, 34
18. Newbiggin, M. I., "The pigments of the Decapod Crustacea," *Jour Physiol.*, 1897, 21
19. Parker, T. J., "On the stomach of Freshwater Crayfish," *J Anat Physiol.*, 1876, 11
20. Patwardhan, S. S., "On the structure and mechanism of the Gastric Mill in Decapoda I.—The Gastric Mill of *Paratelphus guerini* M. Edw.," *Proc Ind Acad Sci.*, 1934-35, 1, (5)
21. Patwardhan S. S., "On the structure and mechanism of the Gastric Mill in Decapoda II—*Brachyura*," *op. cit.*, 1934-35, 1, (7)
22. Patwardhan, S. S., "On the structure and mechanism of the Gastric Mill in Decapoda III—*Anomura*," *op. cit.*, 1934-35, 1, (8)
23. Patwardhan, S. S., "On the structure and mechanism of the Gastric Mill in Decapoda IV—*Macrura Reptilia*," *op. cit.*, 1934-35, 1, (8)
24. Paul, J. H., and Sharpe, J. S., "The Deposition of Lime salts in the Integument of Decopod Crustacea," *J Physiol.*, 1915, 50

25 Paul, J. H., and Sharpe, J. S., "The Relationship of Lecithin to the Growth Cycle in Crustacea," *Bioch Jour.*, 1919, 13.

26 Pearson, J., "Cancer," *LMB C Memoir*, 1908, No. XVI.

27 Pump, W., "Über die Muskelnetze des Mitteldarmdrüse von Crustaceen," *Arch Mikr Anat.*, 1914, 85.

28 Reddy, A. R., "The Gastric Armature of some South Indian Decapod Crustacea," *Jour Annamalai University*, 1934, 4, (1).

29 Reddy, A. R., "A note on the variations in the Gastric armature of some South Indian Decapoda," *Proc 22nd Ind Sci Cong.*, 1935.

30 Reddy, A. R., "On the structure, mechanism and development of the gastric armature in *Stomatopoda*, with a discussion as to its evolution in Decapoda," *Proc Ind Acad Sci.*, 1935, 1, (10).

31 Reddy, A. R., "On the *Modus Operandi* of certain ossicles in the gastric armature of Decapod Crustacea," *Curr Sci.*, 1935, 4, (1).

32 Roaf, H. E., "The Hydrolytic Enzymes of Invertebrates," *Bioch Jour.*, 1908, 3.

33 Smuth, G., "The effect of Sacculina on the storage of fat and glycogen and on the formation of pigments by its host," *QJMS*, 1914, 59.

34 Vitzou, A. N., "Recherches sur la Structure et la Formation des Téguments chez le Crustace Decapode," *Arch Zool Esper.*, 1882, 10.

35 Wallengren, H., "Über das Vorkommen und die Verbreitung der sogenannten Intestinaldrüsen bei den Decapoden," *Zest J Wiss Zool.*, 1901, 70.

36 Weber, M., "Ueber den Bau und die Thatigkeit der sog. Leber der Crustaceen," *Arch Mikr Anat.*, 1880, 17.

37 Williams, L. W., "The stomach of the Lobster and the food of Larval Lobsters," *37th Ann Rep Comm Inland Fish, Rhode Island*, 1907.

38 Young, "Movements of the isolated small Intestine," *Quart Jour Esper Physiol.*, 1915, 8.

39 Yonge, C. M., "The mechanism of Feeding, Digestion and Assimilation in Lamellibranch Mya," *Brit J. Esper Biol.*, 1923, 1, (1).

40 Yonge, C. M., "The mechanism of Feeding, Digestion and Assimilation in *Nephrops Norvegicus*," *op. cit.*, 1924, 1, (2).

41 Yonge, C. M., "On some aspects of Digestion in Clinary Feeding Animals," *Jour Mar Biol Assoc.*, 1935, 20, (2).

42 Yonge, C. M., and Long, D., "The function of tegumental glands in the statocyst of *Homarus vulgaris*," *op. cit.*, 1935, 20, (2).

#### EXPLANATION OF FIGURES

FIG. 1—Alimentary Canal of *Paratelphusa*

FIG. 2—TS of the posterior third of the pyloric chamber  $\times 43$

FIG. 3—TS of the mid-gut  $\times 72$

FIG. 4—TS of the mid-gut  $\times 58$

FIG. 5—TS of the tegumental glands  $\times 232$

FIG. 6—TS of the digestive tubule  $\times 174$

FIG. 7—A secretory cell just before discharging its secretion  $\times 580$

FIG. 8—An absorption cell with vacuoles of absorbed food material.  $\times 580$

FIG. 9—A secretory cell discharging its secretion.  $\times 580$ .

FIG 10.—A secretory cell showing the formation of the vesicle of secretion  $\times 580$

FIG 11.—The longitudinal chitinous ridges bearing setae in the ampullary pouches  $\times 43$

FIG 12.—The gastric armature with the main ossicles separated. The pectinal system of ossicles of the left side alone is shown

FIG 13.—Diagram to illustrate the mode of action of the uro-cardiac ossicle

A Position at rest

B Position at action

FIG 14.—Diagram to show the action of pro-pyloric ossicle

FIG 15.—Diagram to show the action of zygo-cardiac ossicle

FIG 16.—Diagram to show the action of the pectinal system of ossicles

FIG 17.—Temperature curve of the amylolytic enzyme

#### REFERENCE LETTERS

<i>A C</i>	Absorption Cells	<i>M D</i>	Median or uro cardiac tooth
<i>B C.</i>	Basal Cells	<i>M p</i>	Mid-gut
<i>B S.</i>	Blood Sinus	<i>N</i>	Nucleus
<i>C.</i>	Cardiac Chamber	<i>P.</i>	Power
<i>C G.</i>	Cardio-pyloric Constrictors	<i>P<sub>i</sub></i>	Pectinal ossicle
<i>Ch.</i>	Chitinous layer	<i>P P</i>	Pro-pyloric ossicle
<i>C M.</i>	Circular Muscles	<i>P<sub>t</sub></i>	Ptero-cardiac ossicle
<i>C T</i>	Connective Tissue	<i>P P'</i>	Cardio pyloric valve
<i>D D</i>	Dorsal division of the pyloric chamber	<i>P I Lg</i>	Pyloric Chamber
<i>Dg</i>	Digestive gland		Posterior infra lateral cardiac muscles
<i>E.</i>	Epithelium	<i>Pr P<sub>c</sub></i>	Pre-pectinal ossicle
<i>E P</i>	Exo-pyloric ossicle	<i>Pr P<sub>c</sub></i>	Post-pectinal ossicle
<i>F.</i>	Fulcrum	<i>R C.</i>	Roof of the Cardiac Chamber
<i>H C</i>	Hind-gut Cecum	<i>R S</i>	Longitudinal setose ridges in the ampullary pouches
<i>Hg.</i>	Hind-gut	<i>S</i>	Secretion
<i>I Amp</i>	Inter-ampullary ridge	<i>S B</i>	Striated border
<i>I C</i>	Infra-lateral cardiac ossicle	<i>S C</i>	Secretory cells
<i>Int D</i>	Intra-cellular duct	<i>Se</i>	Setae
<i>Inf L.T.</i>	Infra-lateral cardiac teeth	<i>S Amp</i>	Supra-ampullary ridge
<i>K'</i>	Inner Knob-like process of the zygo-cardiac ossicle	<i>U.</i>	Uro cardiac ossicle
<i>K"</i>	Outer knob-like process of the zygo-cardiac ossicle	<i>V D</i>	Right half of the ventral division of the pyloric chamber
<i>L C</i>	Lateral cardiac Plate	<i>V F</i>	Vacuoles with absorbed food material
<i>L D</i>	Lateral or zygo-cardiac tooth	<i>V s</i>	Small vesicles of secretion in formation
<i>L M</i>	Longitudinal muscles	<i>V S</i>	Vesicle of secretion
<i>L.C P.</i>	Lateral cardio-pyloric ossicle	<i>W.</i>	Work
<i>L.M I</i>	Inner longitudinal muscles	<i>Z.</i>	.. Zygo-cardiac ossicle
<i>L.M.O.</i>	Outer longitudinal muscles		
<i>M.</i>	Meso-cardiac ossicle		
<i>M C</i>	Mid-gut Cecum		

## FUNGI OF ALLAHABAD, INDIA.—PART III.

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Received August 9, 1937

[Communicated by Dr Shri Ranjan, M.Sc (Cantab.), Docteur ès Sciences.]

SINCE the publication of Part II of "The fungus flora of Allahabad" a number of other fungi have been collected in this town which are included in this third list.

Special interest attaches to this list of fungi on account of the following points:—

(1) Allahabad is not specifically mentioned by Butler and Bisby in "The fungi of India" as a locality for any of these fungi.

(2) Some of the species are on new hosts. They are marked with an asterisk.

(3) Some of these fungi are not recorded in "The fungi of India." These are marked with a †.

(4) Fifteen species and one genus in the list are described for the first time in *Annales Mycologici-Fungi Indici*, Nos. 1 and 2 by Sydow and Mitter and No. 3 by Sydow, Mitter and Tandon. These are marked with a double asterisk.

Except a few, the fungi were collected by the authors.

No	Name of the fungus	Name of the host
ARCHIMYCETES		
1	<i>Synchytrium rileyi</i> Syd	<i>Peristrophe bicalyculata</i>
PHYCOMYCETES		
*2	<i>Choanephora cucurbitarum</i> (Berk & Rav) Thaxter	<i>Hibiscus esculentus</i>
3	<i>Choanephora infundibulifera</i> (Currey) Cunningham	<i>Hibiscus rosa-sinensis</i>

No.	Name of the fungus	Name of the host
	<i>Choanephora ? sumsons</i> Cunningham	<i>Zinnia</i> sp (cultivated)
	<i>Cystopus blits</i> (Biv) de Bary	<i>Achyranthes aspera</i>
	<i>Cystopus blits</i> (Biv) de Bary	<i>Amaranthus</i> sp
	<i>Cystopus blits</i> (Biv) de Bary	<i>Digera arvensis</i>
*8	<i>Cystopus ipomae-panduratae</i> (Schw) Stevens & Swingle	<i>Ipomoea hirta</i>
*9	<i>Cystopus ipomae-panduratae</i> (Schw) Stevens & Swingle	<i>Ipomoea pes-tigridis</i>
*10	<i>Cystopus ipomae-panduratae</i> (Schw) Stevens & Swingle	<i>Ipomoea pilosa</i>
*11	<i>Cystopus ipomae-panduratae</i> (Schw) Stevens & Swingle	<i>Merremia emarginata</i>
12	<i>Peronospora arborescens</i> (Berk) de Bary	<i>Papaver somniferum</i>
†13	<i>Peronospora obovata</i> Bon	<i>Spergula arvensis</i>
	<i>Peronospora trifoliorum</i> de Bary	<i>Medicago denticulata</i>
15	<i>Peronospora trifoliorum</i> de Bary	<i>Medicago indica</i>
*16	<i>Peronospora viciae</i> (Berk) de Bary	<i>Vicia sativa</i>
17	<i>Phytophthora colocasiae</i> Raciborski	<i>Colocasia antiquorum</i>
18	<i>Sclerospora graminiscola</i> (Sacc) Schroeter	
	ASCOMYCETES	
**19	<i>Coronophora epistroma</i> Syd	<i>Phyllanthus reticulatus</i>
**20	<i>Diaporthe mitteriana</i> Syd	<i>Porana paniculata</i>
†21	<i>Hypoxyylon necrotiosodes</i> Speg.	<i>Tamarindus indica</i>

No	Name of the fungus	Name of the host
22	<i>Hypoxyylon rubiginosum</i> (Pers.) Fr	Dead wood
†23	<i>Pleospora bataanensis</i> Petr	<i>Agave americana</i>
24	<i>Protomyces macrosporus</i> Unger	<i>Coriandrum sativum</i>
✓ 26	<i>Triblidella rufula</i> (Spreng.) Sacc.	<i>Albizia odoratissima</i>
	BASIDIOMYCETS	
	(a) <i>Ustilaginales</i>	
26	<i>Melanopsichium austro-americanum</i> (Speg.) Berk	<i>Polygonum glabrum</i>
27	<i>Ustilago eleusinis</i> Kulkarni	<i>Eleusine egyptiaca</i>
28	<i>Ustilago</i> Kollers	<i>Avena sativa</i>
	(b) <i>Uredinales</i>	
29	<i>Puccinia kuehnii</i> (Krueg.) Butler	<i>Saccharum</i> sp
30	<i>Puccinia pennisetis</i> Zinni	<i>Pennisetum typhoideum</i>
**31	<i>Uromyces gentilis</i> Syd	<i>Mimusops hexandra</i>
32	<i>Uromyces setariae-stalicae</i> (Diet.) Yoshino	<i>Setaria italica</i>
✓ 33	<i>Uromyces</i> sp	<i>Saccharum arundinaceum</i> (No <i>Uromyces</i> is mentioned on this host)
	(c) <i>Hymenomycetes</i>	
34	<i>Amanita</i> sp	
35	<i>Boletus</i> sp	
36	<i>Dädalea flava</i> Lev	
37	<i>Dädalea unicolor</i> (Bull.) Fr	

No	Name of the fungus	Name of the host
38	<i>Favolus scaber</i> B and Br	
†39	<i>Fomes caryophylli</i> (Rac.) Bres	
40	<i>Fomes durissimus</i> Lloyd	
41	<i>Fomes pallidus</i> Petch	
42	<i>Fomes rimosus</i> Berk	
†43	<i>Fomes robustus</i> Karst	
44	<i>Fomes scener</i> Nels & Mont	
45	<i>Ganoderma colossum</i> (Fr.) Bres	
*46	<i>Hexagonia discopoda</i> Pat & Har	<i>Bassia latifolia</i>
*47	<i>Hexagonia discopoda</i> Pat & Har	<i>Tamarindus indica</i>
48	<i>Irpea</i> sp	
49	<i>Lentinus</i> sp	
50	<i>Lensites repanda</i> (Mont.) Fr	
51	<i>Polyporus calcuttensis</i> Bose	
52	<i>Polyporus chocolatus</i> Bose	
53	<i>Polyporus gramocephalus</i> Berk	
54	<i>Polyporus hookeri</i> Lloyd	
†55	<i>Polyporus nilgheriensis</i> Mont	
56	<i>Polystictus hirsutus</i> Fr	
57	<i>Polystictus leoninus</i> Klotzsch	
58	<i>Polystictus velutinus</i> Fr.	

No	Name of the fungus	Name of the host
✓ 59	<i>Poria lacrymata</i> Fr	
60	<i>Poria</i> sp.	Bamboo
*61	<i>Poria</i> sp	Palm
*62	<i>Poria</i> sp	<i>Vitis negundo</i>
*63	<i>Poria</i> sp	<i>Zizyphus jujuba</i>
64	<i>Trametes cingulata</i> Ber	
65	<i>Trametes floccosa</i> Bres	
66	<i>Trametes persoonii</i> Fr	

## (d) Gasteromycetes

✓ 67	<i>Cyathus</i> sp	
✓ 68	<i>Podaxon</i> sp.	

## FUNGI IMPERFECTI

69	<i>Acrothecium lunatum</i> Wakker	<i>Andropogon sorghum</i>
70	<i>Alternaria solani</i> (Ell and Mart.) Jones & Grout	<i>Solanum tuberosum</i>
†71	<i>Ascochyta cycadina</i> Scalka	<i>Cycas revoluta</i>
†72	<i>Botryodiplodia dracaenae</i> (P. Henn.) Petr et Syd	<i>Dracæna</i> sp
†73	<i>Botryodiplodia mammensis</i> (Sacc.) Petr et Syd	<i>Ricinus communis</i>
†74	<i>Botryodiplodia syconophila</i> (Sacc.) Petr et Syd	<i>Ficus religiosa</i>
75	<i>Cephalosporium</i> sp	<i>Helianthus</i> sp.

No.	Name of the fungus	Name of the host
**76	<i>Cercoseptoria balsamina</i> Syd.	<i>Impatiens balsam</i>
77	<i>Cercospora beticola</i> Sacc	<i>Beta vulgaris</i>
78	<i>Cercospora cruenta</i> Sacc	<i>Vigna catjang</i>
**79	<i>Cercospora indica</i>	<i>Cajanus indicus</i>
✓ 80	<i>Cercospora morscola</i>	<i>Morus alba</i>
**81	<i>Cercospora myxa</i> Syd	<i>Cordia myxa</i>
82	<i>Cercospora occidentalis</i> Cke	<i>Cassia occidentalis</i>
83	<i>Cercospora personata</i> (Berk and Curt) Ell & Ev	<i>Arachis hypogaea</i>
**84	<i>Cercospora pulchra</i> Syd	<i>Crataeva religiosa</i>
**85	<i>Cercospora sissoo</i> Syd	<i>Dalbergia sissoo</i>
✓ 86	<i>Cercospora tinospora</i> Syd	<i>Tinospora cordifolia</i>
*87	<i>Cercospora</i> sp	<i>Coccinia indica</i>
*88	<i>Cercospora</i> sp	<i>Mehlotus alba</i>
**89	<i>Cercosporella peristrophes</i> Syd	<i>Peristrophe bicalyculata</i>
**90	<i>Citochorella mangifera</i> Syd Nov Gen. Nov Sp.	<i>Mangifera indica</i>
91	<i>Cladosporium fulvum</i> Cke	<i>Lycopericum esculentum</i>
**92	<i>Clasterosporium concinnum</i> Syd	<i>Ficus religiosa</i>
†93	<i>Colletotrichum dracaena-fragrans</i> (Movi) Petr & Syd	<i>Dracaena ? lindii</i>
†94	<i>Colletotrichum papaya</i> (P. Henn) Syd.	<i>Carica papaya</i>

No	Name of the Fungus	Name of the Host
**95	<i>Colletotrichum tinosporae</i> Syd	<i>Tinospora cordifolia</i>
**96	<i>Dendrographium mitteri</i> Syd	<i>Tinospora cordifolia</i>
*97	<i>Diplodia hibiscina</i> Cke. et Ell. -Var. <i>Sabdariffae</i> sacc	<i>Hibiscus sabdariffa</i>
98	<i>Diplodia</i> sp	<i>Porana paniculata</i>
99	<i>Dothiorella</i> sp	<i>Porana paniculata</i>
†100	<i>Exotrichum leucomelas</i> Syd	<i>Peristrophe bicalyculata</i>
101	<i>Haplosporella dracaenarum</i> (Penz. et sacc.) Petr	<i>Dracaena</i> sp
**102	<i>Haplosporella phyllanthina</i> Syd	<i>Phyllanthus reticulatus</i>
†103	<i>Isaria pulcherrima</i> Berk. et Br	<i>Phanix dactylifera</i>
**104	<i>Lamyella plansuscula</i> Syd	<i>Mimusops elengi</i>
†105	<i>Macrophoma dianthae</i> Gobotto	<i>Dianthus caryophyllus</i>
106	<i>Microdiplodia agaves</i> (Niessl.) Petr	<i>Agave americana</i>
107	<i>Microdiplodia agaves</i> (Niessl.) Petr	<i>Agave cantala</i>
**108	<i>Phomopsis artocarpi</i> Syd	<i>Artocarpus integrifolia</i>
†109	<i>Phyllosticta</i> ? <i>confertissima</i> Ell. et P.v	<i>Ulmus integrifolia</i>
†110	<i>Phyllosticta dracaenae</i> Griff. et Maubl	<i>Dracaena</i> ? <i>lindii</i>
111	<i>Phyllosticta violae</i> Desm	<i>Viola</i> sp.
*112	<i>Phyllosticta</i> sp.	<i>Antigonum</i> sp.
113	<i>Phyllosticta</i> sp	<i>Phanix</i> sp.
114	<i>Septogladium acaciae</i> Syd	<i>Acacia arabica</i>

No.	Name of the Fungus	Name of the Host
†115	<i>Septoria calystegiae</i> West	<i>Convolvulus arvensis</i>
†116	<i>Trichothecium roseum</i> Link	Old wood
†117	<i>Vermicularia hocktersi</i> Syd	<i>Ricinus communis</i>



## IS THE CHROMOPHOBIC PART OF THE GOLGI APPARATUS AND MITOCHONDRIA THE ERGASTOPLASM?

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Received September 18, 1937

(Communicated by Professor R. Gopala Aiyar.)

### *Introduction*

VERY recently from work on *Lycasis* (Subramaniam and Gopala Aiyar, 1936, 1937) it was postulated that in the vertebrates the idiosome should be present in a masked condition as a core to the strands or plates composing the Golgi network. Since then, the problem is being studied from two directions. First, I proceeded on the assumption that if such a core is present in a masked condition, it should be seen during the active synthesis of secretion granules at least in some cases. During experiments with various types of vertebrate tissues it was found that in the liver cells of the tree frog, actually, a chromophobic part becomes differentiated during secretion of bile components (Subramaniam, 1937). In the second line of attack, the pancreas of the toad was selected, where, the chromophobic part of the Golgi apparatus is not visible at any stage. If as has been postulated (Subramaniam and Gopala Aiyar, 1936 and 1937) a chromophobic part is present, it should be possible to demonstrate it by some suitable technique. Bowen (1928) describing certain methods employed by some workers for the demonstration of the idiosome in germ cells suggests their application to vertebrate somatic cells. The fixatives suggested are Bouin, Perenyi, Zenker, Mann or Osmic Acid. The stains used are erythrosin, light green, acid fuchsin, methyl green and resorcin-fuchsin. It will be seen that most of the fixatives mentioned above contain acetic acid or mercuric chloride and most of the dyes are acid ones. The special technique for cytoplasmic components is being followed rigidly only in recent years and the earlier workers employed mostly acetic acid or sublimate fixatives. From a study of the earlier papers on the pancreas it appears that it is from acetic acid and mercuric chloride preparations of gland cells that the Ergastoplasm theory was formulated. Though not heard of now, the Ergastoplasm theory held sway between the years 1890 and 1910.

*What is the Ergastoplasm Theory?*

**Nebenkern**—In order to get a clear idea of the Ergastoplasm theory, it is desirable to know something about the 'Nebenkern' described in some gland cells. This structure described particularly in the pancreas has nothing to do with structures of similar name described in pulmonate germ cells and insect spermatids. Gaule (1881) and Nussbaum (1882) discovered independently in the pancreas cell a body of uncertain nature which they called the 'Nebenkern'. This structure in a gland cell is a mass often of fibrillar structure, the nature and function of which is still in dispute. According to the description of Nussbaum the development of the 'Nebenkern' is related to the phase of the secretory cycle. Thus cells actively synthesizing secretory products were found to have one or more large 'Nebenkerns', while in cells loaded with secretion the 'Nebenkern' might be absent. Ogata (1883) found during his researches on the frog and other Amphibia, that the 'Nebenkern' was most common in frogs which had not been fed for a short time. It disappeared later and appeared again only after feeding. Curious and fantastic descriptions of the origin and structure of the 'Nebenkern' are seen in literature. Morelle (1927) considered that faulty fixation produced a 'Nebenkern' while Benoit (1926) assigns to it a minor place in cellular economy.

**Ergastoplasm**—Side by side with the Nebenkern conception was developed the Ergastoplasm theory. Briefly, the so-called Ergastoplasm is constituted by filaments and these are usually seen only after acetic acid or corrosive sublimate fixations. It should be pointed out here that some workers considered that these filaments seen in sublimate and acetic acid fixations, formed the so-called 'Nebenkerns' of the pancreas, due to spiral twisting or coiling (Mathews, 1899). A clear account of these fibrillar structures was first given by Eberth and Muller (1892). Solger (1894, 1896 and 1898) described them in detail under the name 'basal filaments' and as 'Ergastoplasm' their importance was emphasized by Garnier (1897, 1900 a, 1900 b) and Pernant (1898-99).

The morphology of this structure was never clearly defined but as the name 'Solger's filaments' suggests, it had frequently a fibrillar structure. Divergent structures have been included under the term 'Ergastoplasm' due to their supposed common affinity for acid stains, like chromatin, but if one has to believe Mathews (1899) both have slightly different staining reactions as the following statement of his would show "In Pl XI, Fig 81c one of these threads, stained red runs over the nucleus and ends in the green mass of chromatin" (p 178). This is in the pancreas cell of the hen, fixed

in Hermann and probably stained in acid fuchsin and methyl green. The most interesting aspect of the Ergastoplasm theory seems to be the conception of its alteration in amount (and staining capacity) at different parts of the secretory cycle. Gurnir (1900 a, 1900 b), Prenant (1899-99), Jacobs (1928) and many others have shown that in cells actively synthesizing secretory droplets the Ergastoplasm is fully developed while in cells loaded with secretion it is greatly reduced or entirely absent. Though most of the workers were against the idea of the Ergastoplasmic fibrille being actually transformed into secretory granules, yet, the majority agreed with the view that the development of the Ergastoplasm is precedent to the secretory substance proper.

Coming actually to the disposition of the filaments or fibrille, they have been known to run chiefly in a vertical direction beside the nucleus, while below the nucleus they run in a horizontal direction forming an intricate skein (Bensley, 1911). The Ergastoplasm has been interpreted as artifacts produced by acid fixation by some, while others opine that the Ergastoplasm has a material existence in the form of an unformed material which becomes fibrillar when treated with fixatives which are too acid.

Bowen (1929) sums up with the observation that two different things have been confused under the name Ergastoplasm (1) the poorly fixed chondriome and (2) the concentrated material in the basal region of the cell coming from the capillaries.

#### *The Ergastoplasm and the Chromophobic Part of the Golgi Apparatus and Mitochondria*

My interest in the Ergastoplasm and Nebenkern concepts has been due to the fact that technique once used for the demonstration of the Ergastoplasm and Nebenkern has been found successful for the demonstration of the idiosome in germ' cells. If structures shown by such technique could be considered the idiosome, cannot similar structures shown by similar technique in gland cells also be partly or wholly due to the idiosome substance? In order to discuss the above question one has to have a definite idea of the structure of the mitochondria and the Golgi apparatus. The mitochondria are filamentous in the pancreas and no chromophobic component has been demonstrated in relation with the outside of these filaments by any worker. Bowen (1922) from his researches on the Nebenkern formation during spermatogenesis in insects comes to the conclusion that these filamentous mitochondria should have a chromophobic area in the centre which though not visible is supposed to be present. In other words,

the chromophobic core is supposed to occupy a position similar to that of the copper wire in an insulated cable (Fig. 1). Similarly, Subramaniam and Gopala Aiyar have suggested that a central chromophobic core should be present in Golgi networks which do not show any visible chromophobe

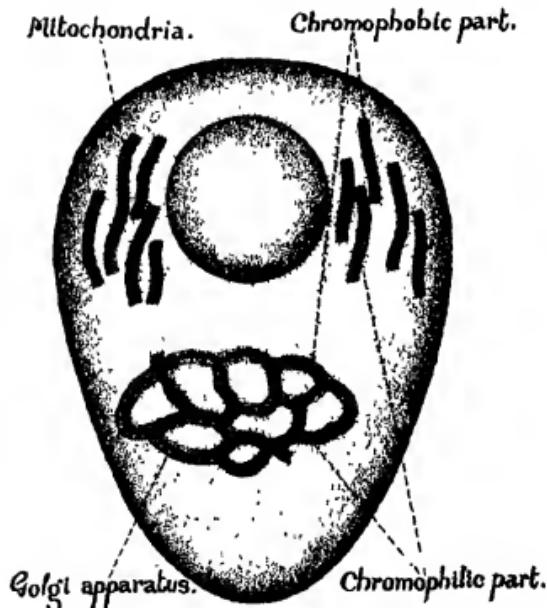


FIG. 1

area. Both the possibilities have been substantiated in part by demonstration of the differentiation of a chromophobic part in mitochondria during Nebenkern formation and the differentiation of a chromophobic area by the strands of the Golgi network in the liver cells of the tree frog during production of bile constituents. The chromophobic part of the mitochondria and Golgi apparatus are supposed to be of a protein composition. It is rather unfortunate that while the chromophobic part of the Golgi apparatus has a definite name—the idiosome—there is no such term

for the chromophobic part of the mitochondria. That the idiosome at least is of a proteid nature could be made from the tests suggested by Bowen (1928) for the demonstration of such a component and with which many workers record a demonstration of the idiosome.

The question that has to be considered is, can the Ergastoplasm be the chromophobic parts of the Golgi and Mitochondria? Bowen (1929) suggests that part of the Ergastoplasm may be only a remnant of the poorly fixed chondriome. The question arises, which part or component of the chondriome is the part that persists? It is common knowledge—though a controversy has been raging on the subject—that many workers consider the mitochondria as having a proteid-lipoid composition. Bowen's suggestion cited above should prove that it is the proteid part that persists, for, lipoids and fats are washed out by corrosive acetic and acetic acid fixatives. A similar proteid-lipoid composition has been attributed to the Golgi apparatus also, but the proportion of the substances are said to be different from that in mitochondria. It is common knowledge that the chromophobic part of the Golgi apparatus in male germ cells has been long known to cytologists under various names, even before the fact that it only forms a part of the Golgi apparatus. Recently, Porson (1927) has demonstrated this component of the Golgi apparatus by a technique originally devised for connective tissue and the detection of mucus. Mucin, it will be interesting to note, is of a proteid nature and hence necessarily the idiosome should also have a proteid composition. Thus we find that the methods employed by the supporters of the Ergastoplasm theory and those who have demonstrated the idiosome were identical. The reason why it escaped the attention of Bowen (1929) who gave a brilliant review of the Ergastoplasm theory as also the methods for the demonstration of the Golgi apparatus and the idiosome (1928) is due to the fact that he paid little attention to the structure of the Golgi apparatus and the mitochondria.

Now it remains to consider the reasons for and against a consideration of the Ergastoplasm as the chromophobic part of the Golgi apparatus and the mitochondria. From a perusal of the literature it appears to me that one of the serious objections is that the filaments of Solger occur only in gland cells, for example in the acinar cells of the pancreas, while the mitochondria occur in acinar cells as well as epithelial cells. It should be pointed out here that great changes in volume and distribution of the Golgi apparatus and the mitochondria occur only in actively secreting cells and not in epithelial cells and a perusal of the papers and Text-books on Histology will show that the shape and complexity of the Golgi apparatus and mitochondria vary in sections of a particular organ composed of different

types of cells.\* Thus the objection cited above is of little value. A more important consideration raised by Bensley (1911) seems to be that the filaments of Solger were not seen in living material whereas the mitochondrial filaments were seen when stained with Janus green. Bensley (1911) seems inclined to believe that the basophile filaments are fixation artifacts due to acid precipitation. He also suggests the other possibility that the Ergastoplasmic fibrillæ may be imbedded in a substance of the same refractive index, and that they may be rendered visible in acid fixations by contraction. On this basis, according to him, it is necessary to assume that the filaments are swollen in chrome sublimate and formalin Zenker preparations, so as to occupy apparently all the space in the cell not taken up by the mitochondrial filaments and fat globules. Another argument of Bensley is that in pancreas fixed in acetic-osmoc-bichromate, stained in amin fuchsin and differentiated in methyl green the mitochondrial filaments are stained intensely red while the basal substance is green. It will be seen that these objections are valid only if both have an independent existence. When we conceive of the possibility of the Ergastoplasmic fibrillæ forming a core to the mitochondria it will be evident that the criterion of differential staining in the manner applied by Bensley does not convince one that the Ergastoplasmic fibrillæ are artifacts.

Bensley makes a distinction between the filamentous mitochondria observed by him and the Ergastoplasmic fibrillæ by their obvious structure, namely, the mitochondria are coarse and bacillus-like, while the filaments of Solger are fine and often form an intricate skein. If the possibility of the chromophobic part of the mitochondrial filaments forming the Ergastoplasmic fibrillæ is admitted, then they have necessarily to be fine filaments.

Thus having considered some of the objections we shall pass on to the resemblances in topography between the Ergastoplasmic or Solger's filaments and the Golgi apparatus and the mitochondria. Bowen (1926) and Hirschler (1918) are of opinion that the mitochondria and the Golgi apparatus have a lamellar structure. This is exactly what has been postulated for the Ergastoplasm by innumerable authors who consider that the fibrillar appearance is only the sectional view of actual plate-like structures. Bensley described mitochondria in the acinar cells of the pancreas as located for the most part in the basal portion of the cell. Recent researches on the pancreas by Hirsch (1931 and 1932) and Duthie (1934) confirm the above observations of Bensley, but it appears to me that these authors have attached too much importance to this basal position of the mitochondria.

\* See Ludford, *J.R.M.S.*, 1925, Fig. 3, p. 357; Cajal, *Histology*, 1934, Fig. 130, p. 167.

and attribute the production of prozymogen to the mitochondria. The Golgi apparatus is network-like and lies just below the nucleus. I believe the fact that during secretory activity the Golgi apparatus and mitochondria increase in number and size needs no reiteration and elaboration. Going further into the resemblances, I find that the earlier workers on Nebenkern and Ergastoplasm have emphasized the fact that in cells loaded with secretion the Ergastoplasmic fibrillae and the Nebenkerns were absent. Thus in almost all cases the development of the Nebenkern and Ergastoplasm has been viewed as in some sense precedent to the synthesis of the secretory material proper. It is exactly at this stage, prior to and during synthesis of secretory materials that hypertrophy of the Golgi and mitochondria occur. Thus having established the probability we shall consider the topography. It should be pointed out here that under the term Nebenkern and Ergastoplasm all sorts of curious structures have been described. The probability considered here is only that between the protein structures usually showing basophilic reactions and the chromophobic part of the Golgi apparatus and mitochondria. In most descriptions of the Ergastoplasmic fibrillae they have been described to have a longitudinal disposition at the sides of the nucleus and a transverse disposition below the nucleus. But even Bensley's descriptions seem to suggest that the images of Solger's filaments may not be superimposable on the pictures of the mitochondria and the Golgi apparatus obtained in the same cell. Here, I believe the cause seems to be the principle of fixation. Students of cytology will be familiar with the fact that the chromosome fixatives are quite unsuited for a demonstration of the mitochondria and the Golgi apparatus. Moreover, solutions containing mercuric chloride and acetic acid do not fix fats and lipoids and hence during the various stages leading up to the clearing agent these have to escape from the tissue leading naturally to the production of currents in the cytoplasm which may be responsible for the production of incorrect pictures of Ergastoplasmic fibrillae which do not fit in exactly with the pictures of mitochondria and the Golgi apparatus. In this connection Mathews' (1899) observation seems to be interesting. He teased living pancreas cells of *Necturus* in saline and while observing them under the microscope introduced mercuric chloride and other fixing agents. When corrosive sublimate is thus introduced he saw these threads rendered visible at the first touch of the fixative, swinging rapidly back and forth in consequence, he suggests, of strong osmosis. Apparently, what he saw was the mitochondria being shorn of its lipoidal part.

*Conclusion.*

I am giving below diagrammatic representations of the position and supposed structure of the Golgi apparatus and the mitochondria (Fig 1) as also the Ergastoplasmic fibrille (Fig 2) in order to demonstrate the

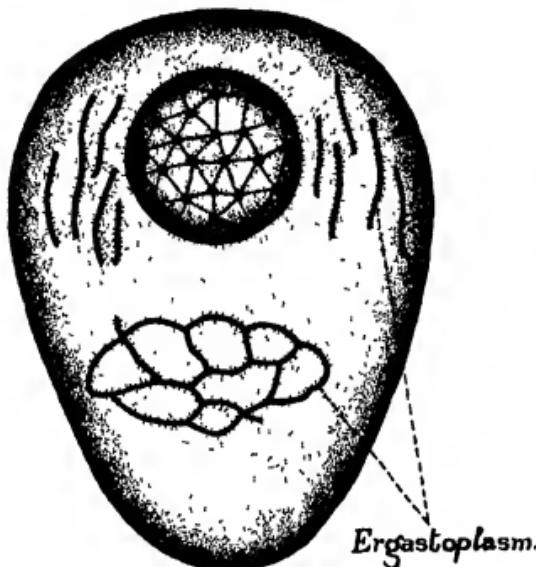


FIG. 2.

the possibility of the chromophobic core of the Golgi and mitochondria being the Ergastoplasmic fibrille. Though only further work on pancreas can justify this assumption, is it not possible that what are described under the terms Ergastoplasm and Nehenkern may, after all, be the chromophobic portions of the Golgi apparatus and the mitochondria?

*Acknowledgments.*

It is a great pleasure to the author to be able to acknowledge his indebtedness to Professor R. Gopala Aiyar, Director of the University

Zoological Research Laboratory, for his advice and criticism and to the University of Madras, for the award of a Research Fellowship

BIBLIOGRAPHY

Benoit, J. . . . . "Recherches sur les voies excretrices du testicule chez les mammifères," *Arch d'Anat Hist et Embryol*, 1926, 4

Bensley, R. R. . . . . "Studies on the Pancreas of the Guinea Pig," *Amer J Anat*, 1911, 12

Bowen, R. H. . . . . "Studies on Insect Spermatogenesis III. On the Nature of the Nebenkern in the Insect Spermatid and the Origin of the Nebenkern Patterns," *Biol. Bull.*, 1922, 42

\_\_\_\_\_. . . . . "Studies on the Golgi Apparatus in Gland Cells IV. A Critique of the Topography, Structure and Function of the Golgi Apparatus in Glandular Tissue," *Quart. Jour. Micro. Sci.*, 1926, 70.

\_\_\_\_\_. . . . . "The method for the Demonstration of the Golgi Apparatus V. The Ithosomic Component, Methods for Lipoids, Troposphongium, Lactonome and Chromidia," *Anat Rec.*, 1928, 40

\_\_\_\_\_. . . . . "The Cytology of Glandular Secretion," (In two parts), *Quart. Rev. Biol.*, 1929, 4.

Dethie, E. S . . . . . "Studies in the Secretion of the Pancreas and Salivary Glands," *Proc Roy Soc., London, (B)*, 1934, 114.

Eberth, C. J., and Müller, K.

Gaule, J.

Garnier, C . . . . . "Untersuchungen über das Pankreas," *Zeit f. wiss. Zool.*, Suppl., 1892, 33

\_\_\_\_\_. . . . . "Kerne, Nebenkerne und Cytozoen," *Centralbl. f. d. Med.*, 1881, 19

\_\_\_\_\_. . . . . "Les filaments basaux des cellules glandulaires," *Bihl. Anat.*, 1897, 5

\_\_\_\_\_. . . . . "Structure et fonctionnement des cellules glandulaires sereuses," *Jour. d'Anat et Phys.*, 1900 a, 35

\_\_\_\_\_. . . . . "Considérations générales sur l'ergastoplasme," *Jour. de Phys. et Path. gen.*, 1900 b, 2.

Hirsch, G. C . . . . . "The Theory of Fields of Restitution with Special Reference to the Phenomenon of Secretion," *Biol. Rev. Camb. Phil. Soc.*, 1931, 61

\_\_\_\_\_. . . . . "Die lebendbeobachtung der Restitution des Sekretes im Pankreas I Teil Die Restitution Der Druse als ganze nach Pilocarplarenzung ihr Ort und ihre," *Zeit f. zellf. u. mikr. Anat.*, 1932, 15.

Hirschler, J. . . . . "Über den Golgischen apparat embryonalzellen," *Arch f. mikr. Anat.*, 1918, 91

Jacobs, W. . . . . "Der Golgische Binnenapparat," *Ergebn. der Biol.*, 1927, 2

\_\_\_\_\_. . . . . "Untersuchungen über die Cytology der sekretbildung," *Zeit. f. zellf. u. mikr. Anat.*, 1928, 8

Mathews, A. . . . . "The Changes in the Structure of the Pancreas Cell," *J. Morph. Suppl.*, 1899, 15.

Morelle, J. . . . . "Les constituants du cytoplasme dans le Pancreas," *La Cellule*, 1927, 37

Nussbaum, M . . . . . "Ueber den Bau und die Thatigkeit der Drusen," *Arch f. mikr Anat.*, 1882, 21

Ogata, M . . . . . "Die Veranderungen der Pankreaszellen bei der Sekretion," *Arch. f. Anat. u. Phys.*, 1883

Poisson, R . . . . . "Recherches sur quelques processus observes dans les elements sexuels jeunes de Notonecta," *Arch Zool exp et Gen.*, 1927, 66

Pienant, A . . . . . "Sur le protoplasma superieur," *J. de l'Anat et Phys.*, 1898-99, 34-35

Solger, B . . . . . "Zur Kenntnis der sezernierenden Zellen der Glandula Submaxillaris des Menschen," *Anat Anz.*, 1894, 9

\_\_\_\_\_. . . . . "Ueber feineren Bau der Glandula Submaxillaris des Menschen," *Festsch zum 70 Geburtstag Carl Gegenbaus*, 1896

\_\_\_\_\_. . . . . "Das Proxymogen der menschlichen Glandula Submaxillaris," *Verhandl. deutsch. Natur u. Artz*, 1898

Subramaniam, M. K . . . . . "The Mechanism of Bile Secretion," *Abst. Ind. Sci. Cong.*, 1937

Subramaniam, M. K., and Gopala Aiyar, R . . . . . "Some Observations on the Possible Mode of Evolution of the network-like Golgi apparatus of Vertebrate somatic cells from discrete Golgi Bodies of Invertebrates," *La Cellule*, 1936, 45

\_\_\_\_\_. . . . . "An Analysis of Shape and Structure of Golgi Bodies in the Eggs of Invertebrates with a Note on the Probable Modes of Origin of the Golgi Network," *Proc. Ind. Acad. Sci.*, 1937, 5, No 4

## FURTHER NOTES ON THE HÆMOPARASITOLOGY OF THE INDIAN BIRDS.

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Received September 30, 1937

IN the present note we will give some statements concerning the Hæmoparasitology of the Indian Birds.

*Trypanosoma gymnorhidis* n. sp.  
Parasite of *Gymnorhitis xanthocollis*, Burton

This bird, shot at Santo Estevão, showed us in its blood smears besides the *Hæmoproteus* which has already been described and named *H. gymnorhidis*<sup>1</sup> some specimens of a Trypanosome, which will be named *Trypanosoma gymnorhidis* n. sp as up to date no Trypanosome has been described in birds of the genus *Gymnorhitis*.

The morphology of this flagellate agrees with the general form of such parasites. Its protoplasm is not uniformly stained by Romanowsky stain, as in some parts it shows some zones, irregularly scattered, taking a denser tinge.

The figure gives a perfect idea of the irregularity of form, number and situation of these zones.

Macronucleus central and surrounded by a large clear area. Very large, it occupies the whole breadth of the parasite, and has an ovoid form. In only one specimen (Fig. 3) its situation is rather subcentral, closer to the posterior pole. The macronucleus has a compact structure in fully grown-up individuals. It suffers a binary division whose initial process seems to begin by a kind of linear split in the middle of the endosomal mass (Figs. 2, 4).

The micronucleus is very small and sometimes not perfectly distinct (Fig. 2). It is situated at a certain distance of the pointed rostrum-like tail of the parasite.

The axoneme of the undulant membrane is generally in the form of a line deeply tinged with blue. Even in slides stained by Heidenhain's iron-hematoxylin, in 50 per cent of the specimens, this organelle does not



*Trypanosoma gymnorhinos*

show a chromatic tinge. In perfectly stained individuals, however, it can be seen, starting from the micronucleus through a fine thread which becomes larger at one to two microns of distance. It continues bordering the undulant membrane and does not end in free flagellum.

*Measurements of Six Individuals (in Microns)*

No. of specimens measured	1	2	3	4	5	6
From the posterior extr. to the kinetoplast						
...    ...	6	2	1	1.5	5	5
From the kinetoplast to trophonucleus						
...    ...	7	11	12	9	12	10
Breadth of the trophonucleus	2	2	2.5	2.5	3	2.5
Free flagellum	0	0	0	0	0	0

*Haemoproteid of Halcyon smyrnensis fusca* (Bodd)

This bird, shot at Santo Estevam, identified by Dr Baimi Prashad, has shown an intense parasitism by an *Haemoproteid* with the following characters.—

Sexual dimorphism, the male gametocytes being colourless or with a slightly blue tone not uniformly spread in the protoplasm when stained by Romanowsky. The small forms are oval and when grown up they are haltheride-like, sometimes surrounding completely the nucleus of the red cell as both extremities of the parasite fuse together leaving often a certain amount of the red cell body unoccupied. The pigment granules are of various sizes, often very minute, often large, isolated or assembled in clusters, showing, however, when compared with the female gametocyte, a tendency to be collected on the poles. In small forms one or two granules only can be stated, but in large forms such granules are more abundant, specially in those very large, surrounding the nucleus of the red cell and in such cases they are scattered all over the body. The nucleus of male gametocyte is very large, generally central, rarely sub-central and occupying a large part of the body, often without definite outline. It is stained in rose by Leishmann's stain, reddish by May-Grunwald-Giemsa.

With this last coloration the protoplasm of the parasite stains slightly reddish violet.

The female gametocyte, oval, fusiform or in the form of a slender haltherid, when young, is definitely haltheridic when grown up. Often the haltherides are somewhat irregular. As in the male gametocytes, both poles of the gametocyte fuse together and surround the nucleus of the red cell, when the parasite is fully grown up. The protoplasm of the parasite is definitely blue with Romanowsky, but the stain is not uniformly spread. The pigment is black-brown, or coffee-brown as in male gametocytes, but generally with a deeper tinge than in the male. With May-Grunwald-Giemsa this pigment in female gametocytes takes sometimes a purple violet tone. Its distribution as well as the size of the granules is irregular. Some specimens show a polar location, but generally subcentral, rather with a polar location.

The red cell is generally not altered when the parasite is young or medium sized. The grown-up specimens displace the nucleus of the red cell to the periphery.

We have found some figures (note Figs. *a* and *b*) which could not be correctly interpreted. Besides the nucleus, they contained small rose corpuscles, which were not however of nuclear nature.



In specimen *c*, we have a very nice type of *accolement* of two female *haltherides*, which could be mistaken for a nuclear division

The outline of both these parasites was clear enough for giving evidence of such *accolement*

#### Classification

*Hemoproteids* have been described in

*Halcyon lindsayi* by Hegner and Chu, 1930 \*

*Halcyon senegalensis* by Theiler, 1930 \*

*Halcyon smyrnensis* (1915) by de Mello, the parasite having been named *Hemoproteus halcyonis* \*

Our parasite is not perfectly the same as *H. halcyonis* from *Halcyon smyrnensis*. We believe that it constitutes a mere variety which will be named *Hemoproteus halcyonis fusca*

*A New Microfilarium of Ardeola grayi Sikes (shot at Santo Estevam)*

Provided with a conspicuous sheath this microfilarid shows its contents packed up in an uniform band-like structure without any distinct separation between them. Its cephalic extremity often does not show any sheath appendix and the nuclear mass begins immediately behind the membrane, sometimes a small clear zone, where one or more granules are noticed, marks a rudiment of anterior sheath, which, in other specimens, is elongated as a glove finger and shows the same structure as the posterior appendix, generally fairly constant and much more developed. The posterior extremity of the nuclear mass may be roundish, twisted or abruptly cut. No definite spot is seen in the body only a split like interruption is found in the centre and rarely near the posterior end.

The sheath appendices are very curious elongated as a glove finger and showing minute granules, scattered all over, fibrils irregularly twisted and patches of substances taking a violet stain with Romanowsky, whose location defies every description and is well figured in the illustrations.

*Measurements in Microns*

No	Total length	Body length	Breadth	Breadth of sheath	Ant. appendix of the sheath		Post. appendix of the sheath	
					Length	Breadth	Length	Breadth
1	162	122	3.5	5	40	4	—	—
2	231.5	193	3.5	7	35	6	3	2.5
3	118	108	4	5.5	10	5.5	—	—
4	216	150	3.5	5	66	5	—	—
5	198.5	110.5	4	4.5	56	7	50	5
6	221	131	4	6	61	4.5	42	5
7	229	144.5	3	6	3.75	7	47	12
8	169	115	3.5	5	56	5	6	3



*Microfilarium limai.*

The senior author has described a microfilarium of this bird under the name *Microfilarium ardeola*<sup>4</sup>. The present microfilarium is a different species which we will name *Microfilarium limas* sp. n. as an homage to Prof J. A. Pires de Lima, from the Faculty of Medicine of Porto.

#### BIBLIOGRAPHY

- 1 de Mello, I. Froulano, "New Haemoproteids of some Indian Birds," *Proc. Ind. Acad. Sci.*, Nov 1935, 2, No 5
- 2 Hegner, Robert and Chu, H. J., "A survey of Protozoa parasitic in plants and animals of Philippine Island," *Philippine J. Sci.*, 1930, 43
- 3 Courtney, G. Robert, "A check list and host index of the genus *Haemoproteus*," *Journal of Parasitology*, 1936, 22
- 4 de Mello, I. Froulano, "Further contributions to the study of the blood parasites of the Indian Birds, together with a list of Haemoparasites hitherto recorded," *J. Royal Asiatic Society of Bengal*, 1936, 2, No 2

ON A CERCOMONAD PARASITE OF THE INTESTINAL  
TRACT OF THE PENTATOMID BUG *ASPONGOPUS*  
*OBSCURUS* (FABR.).

BY I FROLANO DE MELLO, VANTEXA DESSAI

AND

VAMONA XELDENCAR.

(Medical College, Nossa Gôa)

Received April 14, 1937.

OVER fifty per cent of the specimens of the Pentatomid bug *Aspóngopus obscurus* (Fabr.), kindly identified by Dr. Balmi Prashad from the Zoological Survey of India, harbour in their intestinal tract a small cercomonad which will be described in this paper.

In living condition one cannot trace but the movements of the flagellate, whose structure, often very difficult to interpret on account of the small size of the parasite, is fairly well seen in material fixed by sublimate alcohol or Bouin and stained by Heidenhain's iron-haematoxylin.

The general morphology of the parasite is oval, rarely pyriform, the posterior pole having a pointed appearance (Fig. 10). The membrane is of periplastic nature in active forms, the cytoplasm contains often some inclusions of, probably, nutritive material (Fig. 2). The nucleus is oval, of protokaryon type, surrounded by a strongly siderophil membrane and occupies a more or less central position (Figs. 1, 2). On the anterior pole there is a rather large blepharoplast, which seems to us constituted by the fusion of at least two granules, whose union, in such case, takes the appearance of a rod (Fig. 4). From this blepharoplast start three flagella: two very thin, having an equal size and directed forwards, one thicker, of cercomonadic type, adhering to the body which it crosses and becoming free on or near the posterior pole.

The division occurs by mitosis and shows the following phases:—

(a) Premitotic stage where the nuclear karyosome becomes a sort of dust filling completely the endosome (Figs. 3, 9) giving, on a further stage, origin to spireme (Figs. 11, 4).

(b) Mitosis where only the following stages could be observed: prophase with 4 chromosomes and telophase (Figs. 5, 6, 7, 8).



*Trimistus ramaraii* n. sp.

The baso-flagellar apparatus suffers also a division, the daughter granules being attached by a desmose (Figs. 8, 9, 10, 11)

The nucleus in mitosis is considerably enlarged and occupies various positions whose meaning remains for us obscure

We have not been fortunate enough to find figures with the division of the posterior cercomonadic flagellum.

The encystation of the flagellate occurs in the following way: the membrane becomes much thicker than in active forms, the flagella suffer a kind of reabsorption, the remains of the cercomonadic flagellum being the last to disappear. The nucleus seems to increase and the karyosome is reduced to chromatic dust. It is quite possible that some process of division occurs also in the cystic stage, but we have no definite evidence on this point.

*Measurements*—Five flagellates were measured and the statements (in microns) are contained in Table I

*Classification*—Our flagellate belongs to the genus *Trimistus* Alexeïff 1910, sp. typ. *Trimistus motellæ*, parasite of the intestine of the marine fish *Motella tricirrata*

TABLE I

No	Long axis	Breadth	Cercomonadic flagellum	Ant flagella	Nucleus
1	1.5	4	9½	5	1 5
2	3.5	2	7½	4	1
3	3	2.5	6	4	1
4	6	4	15	5	2
5	3	2.5	7	3	1

The characters of the genus *Trimitus* are two anterior flagella and one posterior, this one, thicker, of cercomonadic type, and with its axoneme crossing the body. Nucleus anterior, subterminal, one basal granule giving rise to the three flagella.

The other peculiarities which we find in Wenyon<sup>1</sup> describing the *Trimitus motella*, such as the length of the posterior flagellum, 4 to 5 times longer than the body and that of the anterior flagella, one of which has the same length as the body and the other, half of this length, must be considered as specific characters of *T. motella*.

We do not attach also great importance to the number of the basal granules. In many of these small flagellates, we have seen that such organelles, even when seen under the appearance of one unit are, in reality, provenient of the fusion of more than one basal granule, their anatomic independence being often seen only in mitotic phenomena.

In 1919 Chalmers and Pekkola<sup>3</sup> described under the name of *Dicercomonas soudanensis*, renamed afterwards *Diplocercomonas sudanensis*, a flagellate in human feces much similar to *Trimitus*. Wenyon<sup>1</sup> having examined the original films of those authors, states that they were either *Embadiomonas intestinalis* Wenyon & O'Connor (1917), either *Tricercomonas intestinalis* W & O Con., 1917. If these statements are correct *Diplocercomonas* becomes a *nomen nudum*. If Chalmers and Pekkola are right in their description, *Diplocercomonas* becomes a synonym of *Trimitus*.

The generic name *Trimitus* must be defined in the *Sensu* Alexieff (1910) and not in the meaning which was given to it by Duboscq and Grassé in 1923.<sup>4</sup> The so-called *Trimitus* of these French authors has been shown by Kirby Jr<sup>5</sup> and de Mello<sup>6</sup> to belong to the genus *Tricercomitus* Kirby (1930).

The species we are describing now constitutes the second one belonging to the genus *Trimitus*. We dedicate it to Dr U Rama Rau, the distinguished Director of the "Antiseptic" from Madras and President of the Legislative Council of that Presidency. It will be named, therefore, *Trimitus ramarau* n. sp.

BIBLIOGRAPHY.

- 1 Wenyon, *Protozoology*
- 2 Chalmers, A. J., and Pekkola W., "Diplocercomonas soudanensis," *Jour. Trop. Med. and Hyg.*, 1919, 22, 190.
- 3 Duboscq and Grassé, "Sur quelques protistes d'un Calotermes des îles Loyalty," *Protistologica*, 1929, 14
- 4 \_\_\_\_\_, "Note sur les protistes parasites des Termites de France II. Sur l'évolution de Janickiella grassei Dub and Gr.," *Compt. Rend. Soc. Biol.*, 1924, 90
- 5 Kirby Jr., H. H., "Trichomonad flagellates from Termites I. *Tricercomonitus* gen. nov. and *Hesamastix* Alex.," *Univ. of California Publ. in Zoology*, 1930, 33, 19
- 6 de Mello, Frolano, "Sur un petit flagellé à caractères fixes intermédiaires entre les genres *Trimitus* Nutrichomastix et *Diplocercomonas*," *Sextima Reunión de la Sociedad Argentina de Patología regional del norte* Buenos Aires, 1932 Segunda mitad 907-17.

## ON THE NECTAR SECRETION IN THE COCONUT FLOWERS (*COCOS NUCIFERA*, LINN.)\*

BY G V NARAYANA, B Sc (Ag.)

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Received June 16, 1937

(Communicated by Dr J S. Patel, M Sc, Ph.D.)

Since the floral nectaries have an important bearing on the mode of pollination, the study of the nectar secretion in the coconut flower was pursued. Petch (1913) has recorded the secretion of nectar from three nectaries at the base of the teeth of the pistillode. He has also mentioned the existence of numerous pores which exude a sweet fluid. Sampson (1923) has observed that stigmas when receptive "have a sticky viscous surface in the triradial opening and that honey is secreted by glands at the base of the ovary whose ducts open near the apex of the fruit coat." Huggins (1928) has also observed the presence of nectaries in both the male and the female flowers. Marechal (1928) has recorded the secretion of a sugary fluid from the numerous pores situated round below the stigmas. In the course of the work carried out in this laboratory (Patel, 1937) the presence of three intercarpellary channels which themselves secrete honey was revealed.

### Material and Methods

Male and female flowers in the ordinary tall type of palms under observation were removed from the floral axis at the time of honey secretion. The ovary was fixed in small pieces, but the pistillodes from male flowers were fixed *in toto*. Formalin-acetic-alcohol was used for killing and fixing. After washing in water, dehydrating in graded series of alcohol and clearing in grades of xylol or chloroform, the material was infiltrated with and embedded in paraffin in the usual way. Serial sections 10-15  $\mu$  were cut from the base to the stigma of both the ovary and the pistillode. Sections were stained with safranin and Delafield's haematoxylin, and safranin and light green. Suitable Wratten 'M' filters and point light were used in taking photomicrographs; uniformity of focus in the field was obtained by the use of photo-eyepieces (Zeiss). Freehand sections were made use of extensively for rough examination.

\* Contribution No. 5 of the Oil Seeds Section, Department of Agriculture, Madras.

*Observations*

**Male Flower.**—A clear, sweet fluid is secreted from the triradial opening of the pistillode teeth when the flower opens. The test with Fehling's solution revealed the presence of sugars in the fluid. Secretion is more profuse towards the evening. A delicate scent is also perceived. Bees and ants are some of the more frequent insect visitors.

Serial sections of the pistillode cut from the bottom to the top just at the time when the secretion occurs, reveal three intercarpellary or septal glands. They are narrow, short and free at the base of the pistillode but meet a little way up in the centre and appear like one entire structure with three radiating branches, corresponding to the three carpels of the pistillode (Fig 1). The secreting surface consists of a closely packed palade-like cells with rather large nuclei, dense granular cytoplasm and thin walls. The gland increases in breadth radially and extends upwards to the bases of the pistillode teeth where it opens between the teeth (Fig 2) in the triradial orifice.

**Female Flower.**—After all the male flowers are shed, the female flowers become receptive. The stigmatic region emerges out of the perianth lobes. As the stigmas become receptive, a sweet fluid collects on their inner surfaces. A little below the stigma, there are three orifices (Fig 6, no 9) 1-2 mm long. Each orifice (Fig 6, no 9, Fig 3) is a minute opening alternating with a stigma. Sometime after the stigmatic secretion commences a large drop of nectar collects at each of these three orifices.

Freehand sections showed the nectar oozing from three different centres within the ovary (Fig 4). Each secreting area is situated in between the margins of carpels in a line with each of the arms of the central radial canal (Fig 5). Secretion is most profuse midway between the stigmas and the base of the ovary, and it is very scant (1) in the region of the ovules and (2) in the region between the stigmas and the orifices. It is completely absent at the base of the ovary below the ovules.

Sampson makes mention of three honey glands at the base of the ovary, the "ducts" of which open near the apex of the epicarp. In view of the work reported in this note, it is clear that there are no glands at the base of the ovary and the "ducts" are the nectaries.

The course of the septal glands is indicated in Fig 6. The shape of the gland is like a canal or duct branched and compressed laterally. It is narrow at the upper and lower extremities but broad about the middle (Fig 6). Near the orifices, the glands take a very superficial course and are, therefore, indicated by slightly swollen streaks on the pericarp.

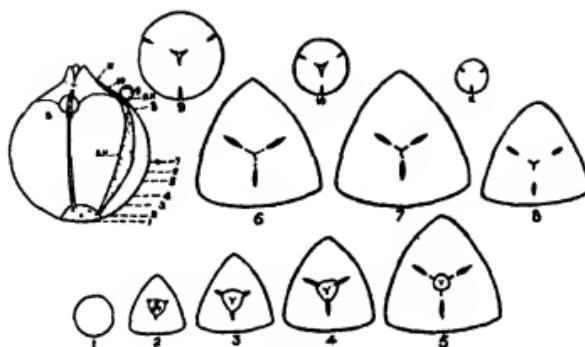


Fig 6.

extending to the stigmas. The gland is lined by closely packed palisade-like secreting cells, as in the male flower and surrounded by a tissue of parenchyma (Figs 7, 8). The nectar from these glands oozes out through the three orifices, mentioned above.

The stigmatic secretion which starts a little earlier than that of the septal nectaries is effected by the epidermal cells of the inner face of the stigma. The triradial canal (Fig. 5) in the centre of the ovary where three carpels meet and which extends from the stigma to the ovule bearing region at the base of the ovary does not appear to take part in secretion.

Petch has mentioned the presence of numerous pores below the stigma "The position of the pores is indicated by whitish spots" round about the base of the stigma. Microscopic examination of this region shows a large number of closely packed needle-shaped crystals (calcium oxalate) in shallow depressions on the epidermis, which is provided with secretory cells (Fig. 9). These cells exude a liquid which on drying leaves incrustations of crystals.

The inner tissue of this region is made up of a mass of parenchymatous cells with a number of scattered, young vascular strands without any secreting elements. But the epidermis consists of closely packed, conspicuously elongated cells. These belong to the category of trichome type of

multi-cellular hydathodes each consisting of a group of 4 to 16 cells, with a narrow base, round head and thick outer wall

Petch has further recorded that the secretion from these "pores" (which are now found to be the secreting elements) forms a ring of liquid preventing the ants from reaching the stigma. In the material examined in the present investigation, this exudation was found to occur long before the stigmas became receptive. The liquid had evaporated and minute deposits of crystals (white specks) were found a week prior to the opening of the stigmas. It is, therefore, very likely that under South Indian conditions the ring of fluid does not exclude the ants from the work of pollination.

#### *Discussion*

Knuth (1909) has stated that the species of genus *Cocos* are anemophilous, but the subsequent investigators have recognized the importance of insects in addition to wind as pollinating agents in the coconut. The secretion of nectar by the male and the female flowers as also the sweet scent of the inflorescence show that the flowers are particularly adapted to attract insects. But there seems to be a difference of opinion as regards the part played by ants in pollination. Petch and Higgins have rejected the possibility of ants acting as agents in pollination, because of their inability to cross the ring of sweet fluid and reach the stigma. It has been shown that the ring of liquid disappears long before the stigmas become receptive. Furtado (1923-25) has shown that ants do help in the pollination of the coconut flower. Kidavu and Namhiar (1925) observe that ants very probably help in transferring pollen.

Septal glands similar to those found in the coconut flowers are reported to occur in *Liliiflorae* and *Scitamineae*. Haberlandt states "The septal nectaries which occur among *Liliiflorae* and *Scitamineae* belong to the category of internal glands. According to Grassman, they arise by the partial non-coalescence of the margins of adjacent carpels. When fully developed they consist of branched or unbranched canals and crevices in the substance of the pericarp, lined by papillose or palisade-shaped secretory elements. Special outlets are provided for the escape of nectar." The "remarkable extra-nuptial nectaries" found at the base of the petiole in the genus *Fagrea* has, according to Haberlandt, some points in common with the septal nectaries. Here the "palisade-shaped secretory cells enclose a cavity which ramifies more or less extensively in the substance of the leaf and opens outside by a passage." The septal or intercarpellary nectaries of the coconut are exactly like those of *Liliiflorae* and *Scitamineae* quoted above.

and bear a remarkable likeness to the extra-nuptial gland in *Fagopyrum* figured by Haberlandt.

Eames and Mac Daniels say "that the secreting cells of the stigmatic surfaces are of the same nature as those of nectaries . . . In all probability the majority of the nectaries are phylogenetically derived from hydathodes." This would mean that the secreting cells of the stigma and nectaries, and hydathodes are closely allied. Thus in the coconut the secretory cells of (1) the septal nectaries, (2) the base of the stigma, and (3) the stigmatic surface, may be essentially of the same origin though of different shape and size. That the secretory cells of the stigma, and the hydathodes are of epidermal origin is obvious from their position. As regards the septal nectaries it may be seen that since they arise by partial non-coalescence of the margins of adjacent carpels they also are of epidermal origin.

#### Summary and Conclusions

(1) Septal nectaries and hydathodes are for the first time reported in the coconut.

(2) Nectar is secreted by the stigmatic surface and by three septal nectaries which are provided with outlets below the stigma. There are no secreting glands at the base of the ovary.

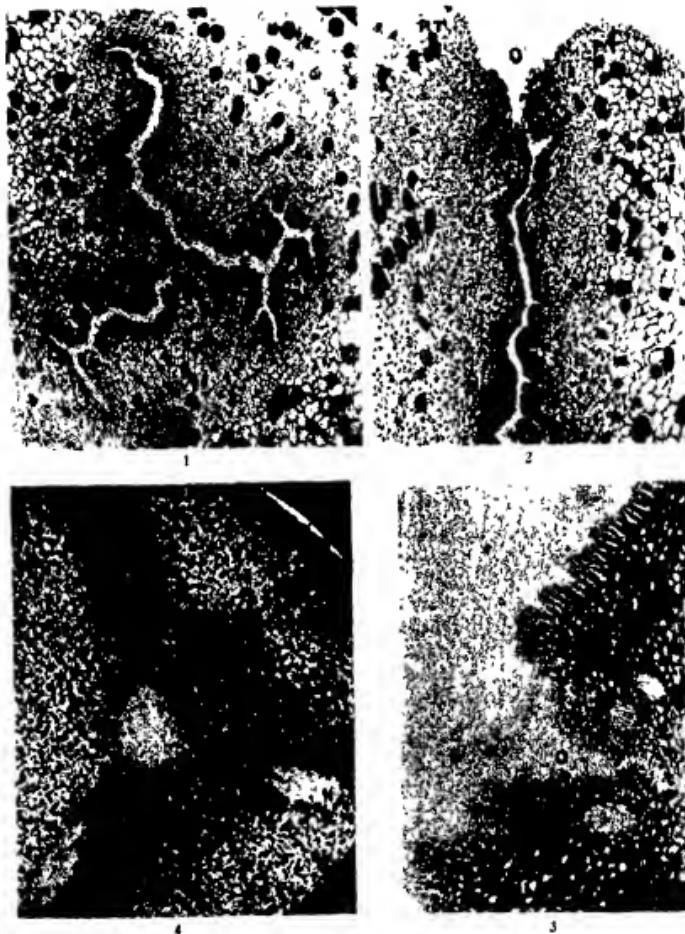
(3) The epidermal hydathodes exude a liquid in the region below the stigma. Under South Indian conditions, in the tall type of palms, this exudation occurs before stigmatic receptivity, and does not exclude ants from reaching the stigma.

(4) In the male flowers also the secretion of nectar is effected by septal nectaries situated in the pistillode.

The author's thanks are due to Dr J. S. Patel, the Oil Seeds Specialist, for much valuable guidance during the course of the work.

#### LITERATURE CITED AND REFERENCES

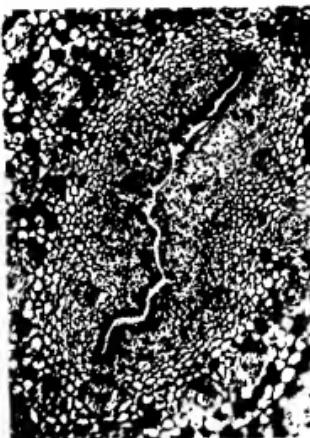
1. Knuth Paul, *Handbook of Flower Pollination* (Eng. edition), 1909.
2. Petch, T., "Coconut flower," *Trop. Agriculturist*, 1913, 41, 449-55.
3. Sampson, H. C., *The Coconut Palm*, 1923.
4. Kidavar, M. G., and Nambyiar, E. K., "Pollination in coconut," *Year-book, Dep. Agric. Madras*, 1925, 33-35.
5. Huggins, H. D., "Pollination and Crop Production," *Agric. Jl. Br. Guiana*, 1928, 1, 164.
6. Marechal, H., "Floral Biology and Pollination," *Agric. Jl. Fiji*, 1928, 1.
7. Patel, J. S., *The Coconut - A monograph*, 1937 (in Press).
8. Furtado, C. X., "A study of the coconut flower and its relation to fruit production," *Gdns. Bull.*, 1923-25, 3, 261-73.







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9. Chamberlain, *Methods in Plant Histology*, 1932.
10. Haberlandt, G., *Physiological Plant Anatomy* (Eng. edition), 1928
11. Eames, A. I., and Mac Daniels, L. H., *An Introduction to Plant Anatomy*, 1925.

LIST OF ILLUSTRATIONS.

FIG. 1 TS of pistillode showing three septal nectaries meeting in the centre.  
C Cavity.  $\times 75$

FIG. 2 LS of pistillode showing one nectary with the outlet O between the pistillode teeth P T  $\times 80$

FIG. 3. TS of part of ovary at the time of secretion showing orifice O and epidermal hydathodes.  $\times 100$ .

FIG. 4. TS of ovary showing three septal nectaries N.  $\times 11$

FIG. 5 TS of ovary showing the central triradial canal  $\times 55$ .

FIG. 6. Ovary—diagrammatic representation of the course of septal nectaries  $\times 1$

FIG. 7. TS of a septal nectary of the ovary C, cavity; SS, secreting surface, P., parenchyma of the nectary.  $\times 84$ .

FIG. 8 TS of part of septal nectary of ovary C, cavity, SS, secreting surface, P., parenchyma.  $\times 320$

FIG. 9. TS of ovary (part of) below stigma showing epidermal hydathodes E H and bundles (broken) of crystals, B C.  $\times 310$ .

## A NOTE ON APLANOSPORES IN A SPECIES OF *OEDOGONIUM*.

By M. S. RANDHAWA, M.Sc., I.C.S.,  
Punjab

Received July 14, 1937.

IN 1928 Handa described certain spore-like bodies in a species of *Oedogonium* which he diagnosed as akinetes. The present author also came across such bodies in a fertile species of *Oedogonium* last year.

These spore-like bodies are 20-24  $\mu$  broad and 30-72  $\mu$  long, and are obovoid in shape. Unlike those observed by Handa, these bodies were

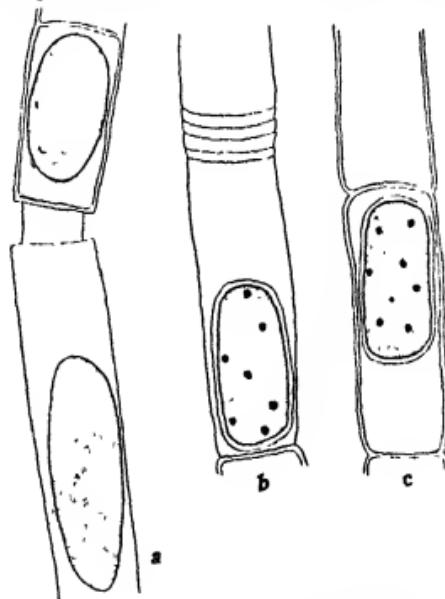


FIG. 1.—Aplanospores in a species of *Oedogonium*

(a) Shows two recently divided cells with aplanospores, (b) and (c) Thick-walled aplanospores in vegetative cells. All  $\times 600$

always found singly inside each cell and never in pairs. In some cases they have a thick walled covering (Figs. b and c), and in one case where these were found inside two recently divided cells, the covering wall was very thin (Fig. a).

The morphology and origin of these spore-like bodies is interesting. Handa collected his material from a hilly place in southern Shan States of Burma, and surmised that a high altitude with its accompanying low temperature may influence their production. The material described in this paper was collected from the plains of the Punjab from a pond in Hoshiarpur District in the month of March 1930. So by no stretch of imagination can one ascribe their formation to low temperature and high altitude.

In the chapter on "Vegetative and Asexual Reproduction of Oedogonials" in his *Structure and Reproduction of the Algae*, Fritsch states in a foot-note, "Handa's record of such stage (referring to resting cells) is open to doubt, as the presence of a parasite is to be suspected." As to what the grounds are on which such a suspicion is based, none are given. Chytridiaceous fungi which are common parasites of some species of *Spirogyra* and *Oedogonium*, are usually globular in shape and much smaller in size. None of these fungi were noticed in the present material. So to call these spore-like bodies as fungi or to suspect their origin due to fungal organisms seems rather far fetched.

How far is it right to call these bodies as akinetes is also open to doubt. Akinetes are usually produced by the transformation of whole cells by the secondary thickening of the cell-wall as in *Pithophora*, *Oedocladium* and *Zygnema giganteum*. In this case these spore-like bodies are formed inside individual cells by the rounding off of the contents and not by the thickening of the walls of the mother cell. Most probably these are zoospores, which having been unable to escape due to certain circumstances, have lost their flagella and developed thick walls. This is apparent from their resemblance in shape with immature zoospores as seen in Fig. a. Hence it is more appropriate to call these bodies as "Aplanospores" rather than "Akinetes". As to whether these aplanospores are capable of germination and producing new individuals can only be found by further observations and experiments.

#### REFERENCES

1. Fritsch, F. E., *The Structure and Reproduction of the Algae*, Vol. I, Camb. Uni. Press.
2. Handa, M. R., "Akinetes in a species of *Oedogonium*," *Journ. Ind. Bot.*, 1928, 7, 1.

# PALÆONTOLOGICAL STUDY OF GASTROPODS FROM LAKI AND BAGATORA, SIND.

BY RAJ NATH, M.Sc., PH.D. (LOND.), D.I.C.

AND

G. W. CHIPLOKER, M.Sc.

(From the Department of Geology, Benares Hindu University)

Received September 20, 1937,

(Communicated by Prof. L. Rama Rau, M.A., F.G.S.)

## I. Short History and Introduction

BEFORE the extensive survey of Sind by W. T. Blanford<sup>4</sup> and Fedden in 1872-74, the stray collections of fossils made by Blagrave and by Capt. Vicary were described by d'Archiac and Haime,<sup>5</sup> who had assigned a lower Tertiary age to all the Tertiary formations of Western India. Later work, however, by Prof. Jenkins<sup>17</sup> on *Vicarya verneuilis*, and the associated forms definitely proved the existence of Miocene beds in Sind.

Of the large collection of fossils made by Blanford and Fedden, during their survey, the Foraminifera were worked out by Prof. R. Jones. Corals were studied by Duncan,<sup>14</sup> while the Echinoids and Mollusca were provisionally identified by Blanford and Fedden—Echinoids were revised by Duncan and Sladen,<sup>15</sup> and the Mollusca were described by Cossmann and Pissarro,<sup>16</sup> Vredenburg<sup>17-19</sup> and Douville<sup>18,19</sup>.

With the idea of doing more detailed and systematic stratigraphical work on lithological and palæontological bases, a large collection of fossils was made, bed by bed, from the neighbourhood of Bagatora (lat. 26° 21' long. 67° 55') and the dome north-west of the village of Laki (lat. 26° 16' long. 67° 57') \*

\* The collection party consisted of the late Prof. K. K. Mathur and Messrs. G. W. Chiplonker, M. L. Misra and V. Bhasker Rao.

N.B.—The following general sequence in the tertiary formations of North-Western India is given for the sake of convenience.

Manchar	Upper	.	Lower Pliocene
Series	Lower	..	Upper Miocene.
Gai Series		.	Burdigalian.
Nari	Upper	..	Aquitanian.
Series	Lower	..	Stampian.
Kirthar	Upper	..	Upper Lutetian and probably Bartonian.
Series	Middle	..	.. Lower and Middle Lutetian.
	Lower (absent in Sind)	.	
Laki	Laki Limestone	..	
Series	Meting Shales	..	
	Meting Limestone	..	
Ranikot Series	Basal Laki Laterite	..	
	..	..	Ypresian
	..	..	.. Landenian.

## ERRATA.

Vol VI, No 4, October 1937

Page 232, line 21—

for Vredenburg<sup>37-48</sup> read Vredenburg<sup>38-48</sup>

Page 246, Species No 64—

for *Magnasperula*, Sacco, var *bagatorenensis*, var nov  
read „ *magnasperula*, Sacco, var *bagatorenensis*, var nov



The present paper is intended to communicate the results of the palaeontological study of Gastropods from the two above-mentioned localities. Other groups of fossils represented in the collection are Foraminifers, Corals, Echinoids, Lamellibranchs, Nautiloids, vertebrate teeth and bones and silicified wood.

*II Description of Species*

*Family TURRITIDÆ, H and A Adams*

*Genus TURRICULA, Herrmann, 1783*

*Turricula lakiensis, sp. nov*

Pl XVI, Fig 1

The solitary specimen has a rather slender spire with three whorls, shouldered at a little distance from the posterior margin. Height of spire whorls is about two-thirds of the width. Posterior to the shouldering the whorls are nearly flat, and convex anteriorly.

Ornament consists of prominent axial ribs, seven or eight per whorl, crossed by spiral threads alternating in two sizes.

Body whorl is large and much inflated, and passes through a slight concavity, into the terminal stem (terminal portion of the body whorl was slightly damaged due to scraping before the specimen was photographed).

Aperture is angulated posteriorly to the shouldering. Anteriorly, the outer lip runs nearly vertical and suddenly turns towards the terminal stem, giving to the aperture an elongate ovate outline. Height of the aperture is about two-thirds of that of body whorl. The inner lip is well defined, rather thin and deflected to the left at the beginning of the posterior canal.

Height of the spire	.	18 mm
Height of the body whorl	.	26 ..
Width of the body whorl	.	17 ..
Height of the aperture	.	17 ..
Width of the aperture	.	11 ..

*Comparison*—Very close resemblance is offered to the present specimen, by *Surcula (Pleurofusia) scala*, Vred, Var? from the Miocene of Myanktin, Burma,<sup>11</sup> but the Sind fossil differs in having a shorter spire, the penultimate whorl markedly smaller than body whorl and the inner lip receding to the left a little sooner.

*Occurrence*—The middle division of the Upper-Middle Kirthar of Laki-Dome, north-west of Laki village.

*Family CONIDÆ, Adams**Genus CONUS, Linnaeus, 1758**Conus vredenburgi, sp. nov*

## Pl. XVI, Fig. 2

The conical spire shows three or four whorls with height about one-fourth of their maximum width, corresponding with the anterior margin. Spire angle is about  $63^\circ$ . Whorls are angulated anteriorly at about one-third height. Region anterior to the angulation is sub-vertical, while the posterior region is conspicuously concave. The slightly raised posterior margin and the angulation give the appearance of two closely situated cords winding round the spire upto the apex. This two-cord structure is likely to suggest six or eight spire whorls. On the concave posterior slope, fine, crowded growth-lines are concave forward. They are antecurrent to the angulation and retrocurrent to the posterior margin. They give to the angulation and to the posterior margin an aspect of a row of scaly plates facing backwards.

Body whorl is conical with feeble convexity. Here the growth lines are convex forward. Anteriorly to the angulation, body whorl is decorated with rather broad, almost flat, ribbon-like spiral threads alternating in two sizes to which the fine growth-lines give a rope-like appearance. Angle of the body whorl is about  $19^\circ$ .

*Comparison*—Due to the two-cord aspect of the spire-whorls, the present species resembles *Conus (Leptoconus) brocki* (Brown)<sup>1</sup>, the latter has, however, wider spire angle, taller spire, wider angle of the body whorl and no rope-like decoration on the body whorl.

*Occurrence*—Middle division of the Upper-Middle Kirthar of Laki-Dome north-west of Laki village.

*Family VOLUTIDÆ, Gray**Genus VOLUTA, Linné, 1758**Voluta* sp. indet. A

The two fragmentary specimens show a very low spire and the axial ribs of the body whorl, giving rise to prominent spines at the posterior margin. These specimens show some resemblance to the Alpine *V. baricornis*, Oppenheimer<sup>2, 3</sup> which however, has a slightly taller spire.

*Occurrence*—Nari Zone 4 B and 4 A of the hill west of Bagatora Railway Station.

*Family CYMATIIDÆ*

*Genus SASSIA*, Bellardi, 1871

*Sassia indica*, Vred var *bagatorensis*, var nov

Pl XVI, Fig 9

This variety differs from Vredenburg's species<sup>43</sup> in having nearly twice (s.e., 22 to 24) as many axial ribs as the species is described to have

*Occurrence*—Nan Zone No. 2, from hill west of Bagatora

*Family CASSIDIDÆ*, Adam

*Genus CASSIDARIA*, Lamarck, 1812

*Cassidaria archiact*, Coss and Piss

1853 *Cassidaria carinata*, Lamarck, *Descr an foss gr Num Inde*, p 317.  
Pl XXXI, Fig 1

1909 *Cassidaria archiact*, Coss and Piss, *Pal Ind*, n.s., Vol III, Mem  
No 1, pt 1, p 39, Pl IV, Figs 8 and 9

1928 *Cassidaria archiact*, Coss and Piss, *Pal Ind*, n.s., Vol X, Mem  
No. 4, p 45

If the specimens, representing this species, from the Upper Rankot series, the Laki Limestone and the Lower division of the Upper-Middle Kirthar be arranged in series their spire shows a progressive shortening as we come up to the younger formations

*Occurrence*—Laki Limestone and Lower division of the Upper-Middle Kirthar of Laki Dome, north-west of Laki village

*Family APORRHAIDÆ*, Philippi

*Genus APORRHAIUS*, de Costa, 1778

*Aporrhais lakiensis*, sp nov

Pl XVI, Fig 5

The spire is slightly conoidal and rather short, about two-fifths of the total height, consisting of four or five slightly convex whorls. Height of the whorls is about two-fifths of their maximum width, situated at the anterior margin. Spire angle is about 75°. Whorls are decorated with thick spiral threads nearly as broad as the intervening spaces

Dorsal ovoid bulge of the body whorl passes anteriorly into an excavation at the neck. At a short distance from the posterior margin a prominent keel separates the posterior concave region, carrying thick spiral threads like those on the spire-whorls. Anteriorly to the keel threads are spaced at about twice their width

At the narrow aperture the keel is turned suddenly towards the apex reaching a little beyond the posterior margin of the third spire whorl, thus showing the extent of attachment of its wing

Following are the dimensions of three of the specimens representing this species —

Total height	63 mm	75 mm	60 mm
Height of spire	26 ..	35 ..	23 ..
Height of body whorl	45 ..	50 ..	40 ..
Maximum width	38 ..	43 ..	37 ..

*Comparison* — This species may resemble, at first sight, the Ranikot fossil *Chenopus dimorphospira*, Cossmann and Pissarro<sup>10</sup>. But the latter has a narrower apical angle of about 43° and taller and fewer spire-whorls.

*Occurrence* — Laki Limestone and Lower division of the Upper-Middle Kirthar of Laki Dome, north-west of Laki village

*Family CERITHIIDÆ, Flenuig*

*Genus CERITHIUM, Brug*

*Cerithium* sp. indet A

The three incomplete specimens have an apical angle of 16° or 17°, height of whorls is two-thirds of their width, sutures are situated in shallow sulcus. Whorls are nearly flat and carry five granular spiral threads, sub-equal and sub-equally spaced, the anterior-most thread becoming a little prominent, two of the specimens show a feeble sixth thread near the posterior suture of the later whorls.

*Comparison* — These specimens resemble *Cerithium vivarium*, mut *alpinum* from the Priabounien and Oligocene of the Alps<sup>5</sup>. Better preserved material is, however, necessary to identify these specimens with any known species.

*Occurrence* — Nari Zone, No. 1 and No. 2, from hill west of Bagatora Railway Station

*Genus BELLARDIA, Meyer-Faymer, 1870*

*Bellardia* cori, sp. nov \*

Pl XVI, Fig 4

1930 *Bellardia* (?) sp. indet, *Pal Ind*, n.s., Vol XV, pt VIII, p. 145.  
Pl XVII, Fig 9

The single available specimen has only three of the spire whorls and the body whorl. Apical angle is 26°. Height of the whorls is about two-fifths

\* This species is named after Mr. L. R. Cox who had first recorded it from Samana Range.

of their maximum width corresponding with the anterior margin. Whorls are nearly flat ornamented with sixteen axial ribs, much narrower than the interspaces on the later whorls, but equal to them on the early whorls, their number thus remaining constant throughout, ribs become less and less significant towards the later whorls. Five spiral striations cross the ribs giving rise to feeble elongate nodes. Posterior margin is immediately followed by a circumsutural ribbon formed by a narrow, shallow groove crossing the axial ribs.

The convex base is much depressed and carries several spiral threads. At about  $200^{\circ}$  from the aperture, there is a conspicuous varix, and another on the second spire whorl, the presence of varices on the intermediate portions cannot be ascertained due to hard incrustation.

*Remarks*—This specimen is identical with the Samana Range fossil described by L. R. Cox, except the spire angle which is  $29^{\circ}$  in the latter. This difference, however, would not separate Samana specimens from the present one.

*Occurrence*—The Nari Zone No 2, from hill behind Bagatora Railway Station.

*Genus TYMPANOTOMUS*, Adams

*Tympanotomus sub-levis*, sp. nov.

Pl. XVI, Fig. 8

The shell is moderately sized, scalariform with apical angle measuring  $22^{\circ}$ . Whorls are one-third as high as their width, and carry two prominent spiral threads; alternating with these on their anterior side are two threads of a little lesser significance, while a still finer thread passes medially. Posterior margin of the whorls consists of a broad band twice as broad as the prominent threads.

*Comparison*—This species differs from the associated *T. levis*, Vred " by the number, the disposition and the degree of differentiation of the spiral threads, as also by its wider apical angle.

*Occurrence*—Nari Zone No 2, from hill behind Bagatora Railway Station.

*Family TURRITELLIDÆ*, Gray

*Genus TURRITELLA*, Lamarck, 1799

*Turritella magnasperula*, Sacco, var *bagatorensis*, var nov.

Pl. XVI, Fig. 3

Shell has a fairly steep apical angle. Height of the whorls is a little more than half their width. The sub-vertical anterior part of the whorls

carries three (or four) very prominent threads, posteriorly to these is a region of fair width. Here due to unfavourable preservation it is difficult to observe any ornamentation. Region posterior to this carries five or six spiral threads which are of lesser significance and also more close-set than those of the anterior region. After this a short slope reaches upto the suture, a corresponding but wider slope near the anterior margin carries two threads sub-equal to those of the posterior region.

*Comparison*—From *T. magnasperula* *sacco* var. *crassoringulata* Vred.<sup>44</sup> The present specimens differ in having three or four prominent threads in the anterior region, wider apical angle, two threads on the anterior slope and five or six threads on the posterior region of the whorls.

*Occurrence*—Nari Zone No 1 and No 4 C, from hill behind Bagatora Railway Station

*Turritella sindensis*, sp. nov.

Pl XVI, Fig. 6

The shell is fairly stout, with apical angle measuring 20° or 21°. Height of the whorls is about half the greatest width corresponding with the anterior keel situated at one-third the height of the whorl. A smaller keel passes at about two-thirds the height of the whorl. The slopes flanking both the keels are feebly convex, except immediately at their bases where they are sharply concave a thread passes close to this concavity. Both the margins have a spiral thread closely following them. Sutures are deeply incised. The shell gives the general appearance of a Pagoda.

*Comparison*—This species has a very characteristic appearance, but when ornamentation is effaced, it might resemble *T. ranikot* Vred. var. *leilanensis*, Vred.<sup>44</sup> from Upper Ranikot beds, the present form, however, has a wider apical angle, the region between the two keels slopes posteriorly and carries no threads, while the corresponding region in the Ranikot fossil is concave and carries three spiral threads.

*Occurrence*—Nari Zone No 1, No. 2 and No 4 B, from hill west of Bagatora

GENUS MESALIA

*Mesalia tricarinata*, sp. nov.

Pl XVI, Fig. 7.

The shell is stout, the spiral angle measuring 20°. Whorls are strongly convex, with height about two-thirds of their maximum width corresponding to the median spiral thread. Sutures are deep. Whorls carry three prominent threads almost like keels, equal and equally spaced from suture to

spire. The concave intervals carry two to four very fine intercalary threads. The convex base carries two threads additional to those continued from the spire whorls. Aperture is sub-circular to oval.

*Comparison*—This species resembles *Mesalia cochlearia*, (Br) var *crassocinctata*, Sacco<sup>1, 20</sup>. The latter, however, has its whorls less convex, sutures a little shallow, narrower apical angle, and spiral threads more numerous and differentiated. *Mesalia fasciata*, Lamarck, from the Cardita Beaumonti beds,<sup>12, 13</sup> has its whorls much less convex.

*Occurrence*—Nari Zone No 1, No 2 and No 4 C, from hill west of Bagatora

*Family EUSPIRIDÆ*

*Genus AMPULLINA*, Lamarck, 1821

*Ampullina*, sp indet A

The two fragmental specimens representing this species have a large sub-oblate body whorl, with the greatest convexity nearer the posterior margin than the anterior extremity. As judged from the disposition of the more or less flat penultimate whorl, the spire should be rather low and depressed with wide apical angle. Aperture is sub-circular. The callus "limb" is not clearly seen.

*Comparison*—In general aspect these specimens resemble the Priabonien and Bartonian Alpine fossil *Natica* (*Ampullina*) *picteti*, Herbert and Renevier<sup>8</sup> from which they differ by shorter (?) spire, a smaller sub-angled aperture, more depressed whorls and more inflated body whorl.

*Occurrence*—Nari Zone No 2 and the upper division of the Upper-Middle Kirthar of the hill west of Bagatora

*Family EPITONIIDÆ*

*Genus EPITONIUM*, Bolton, 1798

*Epitonium mathuri*, sp nov \*

Pl XVI, Fig 10.

A single specimen represents this species. It has five strongly convex whorls, about half as high as their maximum width situated medially. The slightly oblique, acute axial threads are thirty or thirty-two per whorl. The seven spiral threads, of about the same magnitude, as the axial ones, are equally spaced from suture to suture, and give rise to minute tubercles at points of their intersection with axial threads. Fine spiral intercalary

\* This species is named after the late Prof. K. K. Mathur.

threads bisect the spiral intervals, which are narrower than the axial ones. Sutures are deeply incised. Spiral angle measures 18°.

*Comparison*—*Scala gajensis* Vred <sup>44</sup> from the Gaj Beds of Kachh differs from the present species, in not having the spiral threads, while *Scala sub-tenuislamella*, d'Archiac and Haime, <sup>5,44</sup> from the Gaj Beds of Sind has its whorls angulated, a smaller apical angle measuring only 14° and the spiral threads very feeble as compared to the prominent and more close-set axial lamellae. Similar features and a still narrower apical angle excludes the Ranikot form *Scala (Acrilla) colpophora*, Coss and Piss <sup>10</sup> from consideration.

*Occurrence*—Nari Zone No 4 B, from hill west of Bagatora Railway Station

### III General Results from Palaeontological Study

Table II, showing the stratigraphical distribution of the Gastropod species from Laki and Bagatora, shows that twenty-seven species are confined exclusively to the Laki and the Kirthar Beds, fifty-nine species are restricted to the Lower Nari Series, while ten species are common to the upper division of the Upper-Middle Kirthar and the Lower Nari Series. Of these *Ampullospira (Euspirocrommium)* Owens, d'Archiac and Haime and *Ampullina*, sp. indet. B, extend from the upper division of the Upper-Middle Kirthar upto the Nari Zone No 4 E, and *Turbinella*, sp. indet. A, ranges from the Laki Limestone upto Nari Zone No 2. But as these three species are represented by ill-preserved casts, they are left here provisionally as heterogeneous groups.

As regards the zonal distribution of the Gastropod species in the Lower Nari Series, the Zone No 2 stands out very prominently by its richness in the Gastropod species. Out of the fifty-nine species restricted to the Lower Nari Series, forty-three are represented here. The next richest zone is Zone No 4 E, containing twenty-two species, the Zone No 4 B, with twenty-one species comes third. Zone No 1 with twenty species and Zone No 4 C with thirteen species are moderately rich in Gastropod species, while Zone No 3, Zone No 4 A and Zone No 4 D are strikingly poor.

*Cerithium (Pythocerithium) aff perlamellosum*, Vred and *Trochus (Tectus) lucassianus*, Bron are confined to Zone No 1, Zone No 2 has sixteen species confined to it while three species are common to Zone No 1 and Zone No 2, seven species from Zone No. 2 are not represented in Zone No 1, but are continued into the upper zones, so that Zone No 2 can be taken as a unit separated from Zone No 1 on the one hand and Zone No 3 on the other. Now Zone No 3, Zone No. 4 A and Zone No. 4 D do not call

for any special treatment due to their conspicuous poverty in Gastropod species. The Zone No 4 B out of its twenty-one species has five species exclusively confined to it, while only one species it has in common with Zone No 4 C and three species from it reach up to Zone No 4 E, on the other hand, four species are common to Zone No 4 B and Zone No 2. Out of the twenty-two species represented in Zone No 4 E, seven are restricted to it.

From the foregoing discussion it can be concluded that the Lower Nari Series, on the basis of vertical distribution of the Gastropod species, can be divided into Zones No 1, No 2, No 3, No 4 A, No 4 B, No 4 C, No 4 D and No 4 E.

From Table I, showing the zonal sequence in the Lower Nari Series as observed in the field on lithological basis, it can be seen that the thick variegated layers of shales and sandy shales constitute a very prominent feature in the field, separating the three lower zones from the top zone (consisting of five sub-zones). The shale beds intervening between the lower three zones are less sandy and are not as thick as those underlying zone No 4, but are too conspicuous to be overlooked in the field in comparison with the thin but highly fossiliferous beds of limestones, on the other hand, the shale beds intercalated with the five sub-zones, of Zone No 4 are not so prominently thick, as compared to the fossiliferous limestone beds, and yet they cannot be missed in the field.

Thus the zonal sub-division of the Lower Nari Series as arrived at on the basis of the palaeontological study of Gastropods is in full agreement with the one adopted in the field on the basis of lithology. A preliminary study of the Nummulites, Corals and Echinoids also suggests similar results.

As mentioned above, fifty-nine species are restricted to the Lower Nari Series, and ten species, it has in common with the upper division of the Upper-Middle Kirthar. Among the former group, there are thirteen species which are new occurrences to the Bagatora area, eight of these species are known to occur in Miocene beds of Sind,<sup>43, 44</sup> Kuchh,<sup>43, 44</sup> Kathiawar,<sup>43, 44</sup> Baluchistan<sup>43, 44</sup> and Burma<sup>19, 20, 22</sup>. Thus Miocene affinities had begun to be manifest in the Nari fauna of Bagatora area. This would suggest that the bright red to purple coloured, unfossiliferous, calcareous sandstones succeeding the Lower Nari Series at Bagatora<sup>4</sup> are very probably the representatives of the Gaj Series.

In conclusion, the authors take this opportunity of expressing their indebtedness to Dr A Morley-Davies, lately of the Imperial College of Science and Technology, London, for his kind suggestions and valuable

criticism. Thanks are also due to the Director, Geological Survey of India, for kindly lending the necessary literature from time to time and also for the access to the type specimens preserved in the Indian Museum, Calcutta.

TABLE I  
*Table showing the Zonal sequence of the Lower Nari Series  
as observed in the Field*

Zones	Thickness	Lithological and faunal characters
No 4 E	1 ft	Brown shaly limestone breaking into thin slabs rich in flat Echini nodes and Gastropods, Nummulites and Lamellibranchs are present in moderate numbers
Shales	1 to 2 in	Unfossiliferous, coloured brown
No 4 D	1 ft	Greyish brown limestone breaking into irregular blocks. Rich in Nummulites, Gastropods, Echinoids and Lamellibranchs are not very abundant
Shales	1 to 2 in	Unfossiliferous, coloured brown
No 4 C	1 ft 3 in	Greyish brown limestone breaking into irregular blocks. Moderately rich in Nummulites and Gastropods, poor in Lamellibranchs
Shales	1 to 2 in	Unfossiliferous, coloured brown
No 4 B	1 ft 3 in	Reddish yellow limestone breaking into thin plates. Rich in Gastropods, Corals, Echinoids, Nummulites and Lamellibranchs are moderately represented
Shales	1 to 2 in	Unfossiliferous, coloured brown
No 4 A	1 ft 6 in	Yellowish limestone breaking into thick long slabs. Poor in Gastropods; Nummulites and Lamellibranchs are represented fairly well
Shales	43 ft, 3 in	Unfossiliferous, coloured yellowish brown and containing gypsum
Sandy shales	9 ft	Unfossiliferous, coloured grey
No 3	3 ft	Yellowish brown limestone breaking into thick slabs. Very poor in Gastropods, rich in Echinoids and Nummulites, Lamellibranchs and Corals are fairly represented.
Shales	55 ft	Unfossiliferous, coloured yellowish and pink
No 2	9 in	Greyish brown limestone breaking into thin slabs. Rich in Nummulites, Gastropods and Corals; Echinoids and Lamellibranchs are present in moderate numbers
Shales	14 ft 10 in,	Unfossiliferous, coloured white, red, brown
No 1	2 ft	Brownish and yellowish limestone hard, rather compact and not breaking very easily. Rich in Nummulites; moderately rich in Gastropods; Lamellibranchs and flat Echinoids are also present, tubes of <i>Terebra</i> are very common, occurring in all possible positions.

TABLE II  
Table showing the Stratigraphical Distribution of Gastropods from Laki and Bagatora, Sind.

Species	Upper Middle Kirthar				Lower Nan Seine			
	Laki Lam- sition	Lower Middle	Upper Middle	Zone	Zone	Zone	Zone	Zone
Order Opisthobranchia				1	1	1	1	1
1 <i>Acone sericea</i> , Vred								
2 <i>Sophularia oligostoma</i> , Sacco				1 (2)	1			
3 <i>Parmaea latissima</i> , sp. nov								
4 <i>Ovula (Lithoxoidea) brevis</i> , J. de C. Sow								
5 " " <i>medialis</i> , Nicht				2	1	2	2	2
6 " " <i>mediterranea</i> , sp. nov				2				
7 " " sp. indet				1				
8 <i>Ovula rugosa</i> , d'Archia and Haane				2				
9 " <i>(Strophium) medialis</i> , Dürck, var <i>indica</i> , Vred					2		4	4
10 <i>Atrita (Spirula) radiata</i> , Vred								2
11 <i>Eupra (Eocula) sericea</i> , Vred							2	2
12 <i>Atrita (Volutospira) sinuosa</i> , Vred						7		
13 " " <i>dentata</i> , Sow " var <i>spicata</i> , d'Archia and Haane						1	1	1

TABLE II—(Cont'd.)



TABLE III—(Contd.)

71.	<i>Metaphysaria</i> , sp. nov.	1
72.	<i>Prostomia strobigena</i> (d'Archim.)	1
73.	" <i>reniformis</i> (d'Archim and Haine)	3
74.	" <i>retrocurvata</i> , Vind	2
75.	" <i>oblongum granulatum</i> , A. de G. Rose	7
76.	<i>Archidistomella varia</i> (Trem.)	8
77.	<i>Notosia angulifera</i> , d'Orb	1
78.	" <i>(Vivipara) adhaes</i> , Cote and Prie.	2
79.	" <i>longipora</i> , Lym.	1
80.	<i>Symploca</i> , sp. indet	1
81.	<i>Gymnola</i> , sp. indet A	3
82.	" sp. indet B	1
83.	<i>Argylospira (Papirio-criniformis) oceanica</i> d'Arch and Haine	1
84.	<i>Argylospira (Globularia) gallorensis</i> , Graulholz	1
85.	" <i>rotundata</i> , d'Arch and Haine	3
86.	" <i>spiculifera</i> , Lamark	3
87.	" sp. indet A	1
88.	" sp. indet B	1
89.	<i>Epidictyon sebastenae</i> , sp. nov	12
90.	<i>Foliuma schmidiana</i> , Chenu	1
91.	<i>Turbo</i> , sp. indet A	1

TABLE II—(Concl'd)

Species	Lata Lime- stone	Upper Middle Kurnar				Lower Narm Series					
		Lower	Middle	Upper	Zone 1	Zone 2	Zone 3	Zone 4 A	Zone 4 B	Zone 4 C	Zone 4 D
92 <i>Terebratula</i> , sp. indet. B	1	1									
93 " sp. indet. C											
94 " sp. indet. D	1				1						
95 <i>Terebratula heteromera</i> , Brön										3	1
96 <i>Terebratula sub-circularis</i> , d'Orb											
Total Number of Species in each Bed	17	11	7	17	20	43	1	4	21	13	5
											22

N.B.—Numbers in the different columns indicate the number of specimens representing the species in the particular bed.

BIBLIOGRAPHY

1. d'Archiac, *Histoire des Progrès de la Géologie*, 1850, 3
2. d'Archiac and Haime, *Description des animaux fossiles du groupe Nummulitique de l'Inde*, 1853
3. Bellardi, *Moll. terr. terz. del Piemonte e della Liguria*, 1890, 4
4. Blanford, W. T., "Genlogy of Western India," *Mem. G.S.I.*, 1879, 17, Pt 1
5. Boussac, J., "Étude Paléontologique sur la Nummulitique Alpin," *Mem. Ép. Spéc. carte Géol. détail France*, 1911.
6. Brogniart, *Mémoire sur les terrains de Sédiment du Vicentin*, 1823
7. Chemnitz, *Illustrations Conchyliologiques*, 1835
8. Cossmanu, *Essays de Paléoconchologie Comparée*, 1899, fasc. III
9. ———, *ibid.*, 1903, fasc. V
10. ———, and Pissarro, "The Mollusca of the Ranikote Settes," *Pal. Ind.*, n.s., 1909, 3, mem. I
11. Cox, L. R., "Fossil fauna of the Samana Range and some neighbouring areas," *Pal. Ind.*, 1930, n.s., 15, pt. viii
12. Douville, H., "Les Couche à *Cardita beaumonti*," *Pal. Ind.*, n.s., 1928, 10, fasc. I
13. ———, "Les Couche à *Cardita beaumonti*," *Pal. Ind.*, n.s., 1921, 10, mem. III, fasc. II
14. Duncan Martin, P., "Sind fossil Corals and Alcyonaria," *Pal. Ind.*, 1880, (14), 1, pt. ii
15. ———, and Sladen, "The fossil Echinoids of Sind," *ibid.*, 1883, (14), 1, pt. iii.
16. Grateloup, *Atlas Conch. foss. terr. terz. bassin de l'Adour*, 1840
17. Jenkins, H. M., "On some Tertiary Mollusca from Mount Stela in the Island of Java," *Q. J. G. S.*, London, 1864, 20
18. Michelotti, *Etude Mio Inf. Ital. Septentrionale*, 1861
19. Noetling, F., "On some marine fossils from the Miocene of Upper Burma," *Mem. G.S.I.*, 1895, 28, pt. 1
20. ———, "Fauna of the Miocene Beds of Burma," *Pal. Ind.*, n.s., 1901, 1 mem. 3
21. Oppenheim, *Palaeontographica*, 1900, 43
22. ———, *ibid.*, 1906, 30
23. Reeve, *Conchologia Ionica, Monograph of the genus Buccinum*, 1849
24. ———, *Conchologia Ionica, Monograph of the genus Oliva*, 1851.
25. Sacco, *Moll. Terr. terz. del Piemonte Liguria*, 1890, 7
26. ———, *ibid.*, 1891, 8
27. ———, *ibid.*, 1893, 13
28. ———, *ibid.*, 1894, 15
29. ———, *ibid.*, 1895, 17.
30. ———, *ibid.*, 1896, 19.
31. ———, *ibid.*, 1896, 21.
32. ———, *ibid.*, 1897, 22
33. ———, *ibid.*, 1904, 30.

34 Sowerby, J. de C., *Trans G. & Soc. Lond.*, 1840, (2), 5.  
 35 Tryon, *Manual of Conchology*, 1881, 3.  
 36. ———, *ibid*, 1883, 5.  
 37 Vredenburg, E., "Comparative diagnosis of Pleurotomidae from the Tertiary formations of Burma," *Rec. G. S. I.*, 1921, 53, pt. 2.  
 38 ———, "Analysis of the Singu fauna founded on the Rao Bahadur S. Sethu Rama Rau's collection," *ibid*, 1922, 53, pt. 4.  
 39 ———, "Indian Tertiary Gastropoda—IV," *ibid*, 54, pt. 3.  
 40 ———, "Indian Tertiary Gastropoda—V," *ibid*, 1923, 55, pt. 2.  
 41 ———, "Indian Tertiary Gastropoda—VI," *ibid*, 1923, 55, pt. 2.  
 42 ———, "Description of the Mollusca from the Post-Eocene Tertiary formations of North-Western India—Cephalopoda, Opisthobranchiata, Siphonostomata," *Mem. G. S. I.*, 1925, 50, pt. 1.  
 43 ———, "Review of the genus *Gisortia*," *Pal. Ind.*, n. s., 1927, 7, mem. iii.  
 44 ———, "Description of the Mollusca from the Post-Eocene Tertiary formations of North-Western India—Gastropoda (in part) and Lamellibranchiata," *Mem. G. S. I.*, 1928, 50, pt. 2.  
 45 ———, "Supplement to the Mollusca of the Ranikote Series," *Pal. Ind.*, 1928, n. s., 10, mem. iv.

#### EXPLANATION OF PLATE

FIG 1 *Turritella lakunensis*, sp. nov., from the middle part of Upper-Middle Kirthar, near Laki, (B H U No G/148)  
 FIG 2 *Conus vredenburgi*, sp. nov., from the middle part of Upper-Middle Kirthar near Laki, (B H U No G/141a)  
 FIG 3 *Turritella magnasperula*, Sacco, var. *bagatorenensis*, var. nov. from Lower Nari Zone No 1, near Bagatora, (B H U No G/75b)  
 FIG 4 *Bellardia costi*, sp. nov., from Lower Nari Zone No 2 from Bagatora, (B. H. U No G/94)  
 FIG 5 *Aporrhais lakunensis*, sp. nov., from lower part of the Upper-Middle Kirthar near Laki, (B. H. U No G/128b)  
 FIG 6 *Turritella sudensis*, sp. nov., from Lower Nari Zone No 2 from Bagatora, (B H U No G/77a)  
 FIG 7 *Mesalia trikarmata*, sp. nov., from Lower Nari Zone No 2 from Bagatora, (B H U No G/78e)  
 FIG 8 *Tympanotoma sub-lirata*, sp. nov., from Lower Nari Zone No 2 from Bagatora, (B H U No G/93)  
 FIG 9 *Sussia indica*, Vred. var. *bagatorenensis*, var. nov., from Lower Nari Zone No 2 near Bagatora, (B H U No G/152)  
 FIG 10 *Epitonium mathura* sp. nov., from Lower Nari Zone No 4 B near Bagatora, (B. H. U No. G/131)

N.B.—All Figures are natural size

Fig. 5 is a photograph from a drawing made faithfully to the Type specimen, except that the details are shown more clearly than on the weathered specimen itself.





# ON THE ORIGIN AND DEVELOPMENT OF THE CEMENT GLANDS IN *ETROPLUS MACULATUS* (BLOCH).

BY S JONES

(From the University Zoological Research Laboratory, Madras.)

Received October 1, 1937

(Communicated by Prof R. Copala Aiyar)

## Introduction

WHILE engaged in working out the development of some brackish water fishes of Adyar,\* I had the opportunity to investigate the development of the cement glands in two Cichlid fishes, *Etroplus maculatus* and *E. suratensis*. These glands have the same function as in the larvae of Ganoids, Teleosts, Dipnoids and Anura in which similar organs have been discovered. The developmental origin of these larval organs has been worked out in the case of Ganoids, Dipnoids and Anura. In the first group they arise from the endoderm whereas in the latter two, they are of ectodermal origin (Kerr, 1919). Though cement glands are known to occur in some tropical Teleosts like *Etroplus*, *Pterophyllum*, *Sarcodaces* and *Hyperoplus*, no work on their origin has been done so far to my knowledge †. The present paper deals with the cement glands in one of the local forms, *Etroplus maculatus*.

These structures, found only in the aquatic larvae of some fishes and amphibians, are all purely larval organs primarily concerned with the attachment of the larvae to some foreign object to prevent their being swept away by currents. Except in the case of the larvae of *Acipenser*, where they are transformed into barbels, the cement glands degenerate and disappear in course of time. In the case of *Etroplus* also the glands are most active during the earliest larval stage and then atrophy as soon as the young fish is capable of swimming about.

## Historical

*Ganoids*—It was Agassiz (1879) who first worked on the cement glands of fishes when he dealt with the function, degeneration and disappearance of the gland (equatorial disc) in *Lepidostoleus*. He did not mention anything

\* S Jones, "Observations on the Breeding Habits and Development of Certain Brackish Water Fishes of Adyar, Madras," *Proc. Ind. Acad. Sci. (B)*, 1937, 5, No. 6.

† S Jones, *Curr. Sci.*, 1937, 5, No. 9.

as to its origin. In 1881, Balfour dealing with the same form, considered the suctorial papillæ of the gland as epiblastic sensory thickenings and later in 1882, Balfour and Parker speak of them as modified cells of the mucous layer of the epidermis pouring out a sticky secretion. Phelps (1899) as a result of her work on *Amia calva* said that the gland is endodermal in its origin. Subsequently, Reighard and Phelps (1908) gave a detailed account of the origin and development of the gland in *Amia calva* and they attributed a similar (endodermal) origin to the gland in *Lepidosteus*. About the same time appeared the work of Eycleshymer and Wilson (1908) and these authors agreed with regard to the endodermal nature of the glands in *Amia calva*.

Kerr (1906 and 1907) worked out the development of the gland in *Polypterus senegalus* (Budgett's collection) and found it to take its origin from the endoderm. He was not at the time aware of the work on *Amia calva* by Phelps. Kupffer (1893) thought that the cement organs of *Acipenser sturio* was of ectodermal origin. Sawadsky (1911) basing his work on *Acipenser ruthenus* said that the organ in question actually originates from the ectoderm. Thus the endodermal development of the cement glands in Ganoids was finally settled.

*Dipnoids and Anura*—Among the Dipnot cement glands are known to be present only in *Lepidosiren paradoxa* and *Protopterus annectans*, where they have been found to be of ectodermal origin (Kerr, 1903 and 1919). Assheton (1896) and Bhaduri (1935) worked out their development in *Rana temporaria* and *R. afghana* respectively and found them to originate from the outer layer of ectoderm. The difference in these two groups is that in the Dipnot the secretory cells of the glands develop from the inner layer of ectoderm whereas in the Anura they develop from the superficial layer of ectoderm.

*Teleosts*—Cement organs are known to occur only in a very few tropical Teleosts. It was first discovered by Budgett (1901) in the larvae of two African fishes, *Sarcodaces ota* (Characidae) and another which he provisionally determined as *Hyperopisus bebe* (Mormyridæ). Assheton (1907) when dealing with the sense organs and mucous canals in the larvae of *Gymnarchus niloticus* and *Helicopis niloticus* mentions about the presence of mucous secreting glands on the head. It is possible that they are similar to the glands in *Hyperopisus* and *Sarcodaces* though Budgett who examined the living specimens did not observe any adhesive organs. Probably they are degenerate. They are known in some Cichlids like *Eretroplus suratensis* (Willey, 1911), *F. maculatus* (Sundara Raj, 1916) and *Pterophyllum cuneiceps*

(Lieberkind, 1931). Of these Cichlids the last one is South American and the other two are South Indian forms.

From the literature available it appears no work has been done so far on the origin of the gland in Teleosts. Lieberkind (1931) described the well-developed and active gland of the five days old larva of *P. cinctus* and mentions its absence in a 10 days old one. But due to lack of sufficient material he could not work out the origin of the gland.

*Cement Organs of Etroplus maculatus*—In the case of this fish, as soon as it hatches out, the glands can be seen as three pairs of symmetrically arranged conical projections each with a depression at the top. One pair is situated at the anterior extremity of the head near the olfactory organs, whereas the other two pairs are found close together dorsal to the eyes (Figs 1 and 2). These are larger than the anterior-most pair. The glands

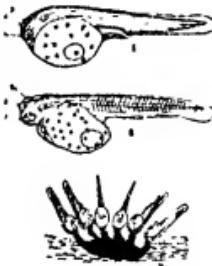


FIG. 1.—Newly hatched larva.  $\times 22$  1, 2 and 3 = Cement organs

FIG. 2.—Larva 2nd Day.  $\times 22$

FIG. 3.—A group of young larvae resting at the bottom attached by their mucous threads.  $\times 6$ .

secrete mucus till the fourth day after which they become inactive and gradually atrophy. During this short period the secretion helps to keep the larvae in one place under the care of the parents, thus affording protection (Fig. 3).

*Origin and Development*—The cells of the gland rudiment gradually become distinct in the thirty-eight hours old embryo. The rudiment develops as a thickening of the inner layer of ectoderm (Fig. 4). The cells are long and conical with the nuclei at their base. They soon curve over enclosing a space, the gland cavity, roofed over by the thin superficial

layer of ectoderm. Fig. 5 is a transverse section of the head passing through the middle pair of glands in a fifty-four hours old embryo

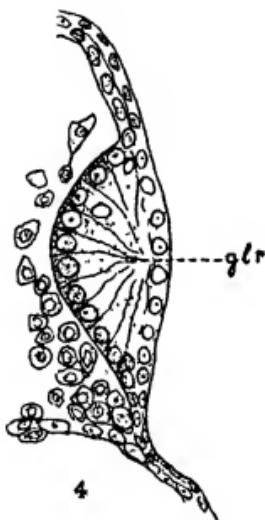


FIG. 4.—Section through a gland rudiment of a 38 hours old embryo glr, gland rudiment

Hatching takes place seventy-two hours after fertilization at which time all the three pairs of glands are active. The secretion of the gland cells fills the cavity and bursts open the superficial layer of ectoderm which gets peeled off. Now the cavity widens. This is accompanied by the raising of the sides of the cavity which results in the formation of a deep gland cavity opening to the outside. The superficial layer of ectoderm extends up to the edge of the gland cup (Fig. 6). No "brush border" of the kind mentioned by Bhaduri (1935), lining the gland cells in *Rana afghana*, has been observed.

Several larvae are found anchored to one place by mucous threads emanating from the cement glands. The parents have, as has been already observed, the curious habit of transferring their broods from place to place and each time, the young ones attach themselves once more by mucous

threads. In the attached condition the larvae remain with the dorsal surface of the head touching the substratum with the body held at an angle to the ground and the tail vibrating incessantly. When disturbed they break off from their moorings and move off in this characteristic position trailing a quantity of dirty mucus behind them. They again collect together in groups and a fresh attachment is effected.

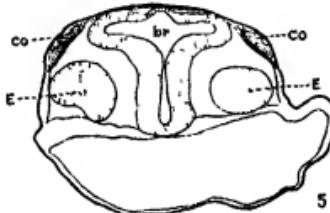


FIG 5.—Section through the head of a 54 hours old embryo passing through the middle pair of cement glands.

FIG 6.—Section through the cement organ 1st Day  $\times 380$

FIG 7 " " " 2nd Day  $\times 380$ .

Abbreviations br, Brain, co, Cement Organ, E, eye, glc, Gland cavity

The glands grow in size (Figs 7 and 8). Meanwhile the inter-space between the pairs of glands increases. Gradually, they get flattened out and the cells become vacuolated. The cell walls become indistinct and degeneration sets in. The rapidity of the process can be understood by the change

the glands have undergone by the fifth and the sixth day (Figs. 9 and 10). As soon as they shrink in size the superficial layer of ectoderm from the

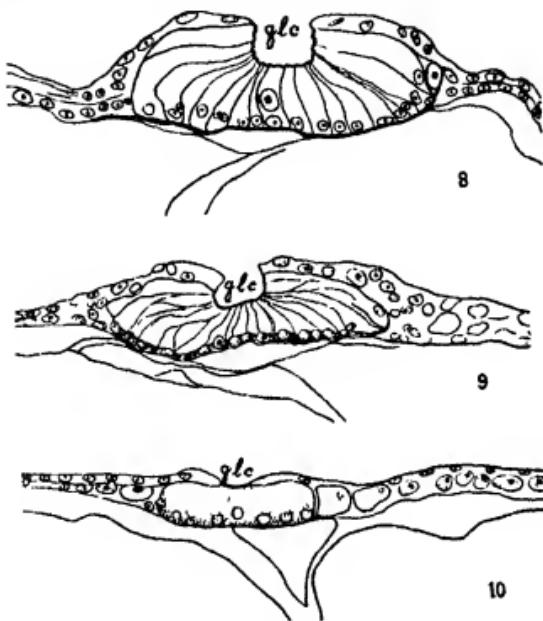


FIG. 8.—Section through the cement organ 4th Day.  $\times 380$

FIG. 9. " " " " 5th Day.  $\times 380$

FIG. 10. " " " " 6th Day.  $\times 380$ .

sides, which at this time gets considerably thickened, grows over the rapidly degenerating structures which are soon lost. By the seventh day it is hardly possible to locate their position.

The structure and position of the cement glands of *E. suraensis* are exactly as in *E. maculatus*. Though their origin and development have not been worked out it is not likely that there will be any difference.

The following table gives the origin and fate of the cement organs in the vertebrates in which these structures have been studied

Group	Genus	Origin	Fate
Ganoidel	<i>Amsa</i>	Endodermal	Disappear
	<i>Acipenser</i>	"	Transformed into barbels
	<i>Lepidosteus</i>	"	Degenerate
	<i>Polypterus</i>	"	Disappear
Teleostei	<i>Etroplus</i>	Ectodermal (Inner layer)	Disappear
Dipnol	<i>Protopterus</i>	Ectodermal (Inner layer)	Disappear
	<i>Lepidostren</i>	"	"
Anura	<i>Rana</i>	Ectodermal (Outer layer)	Disappear

#### Discussion

The present work has shown that the cement organs of *E. maculatus* take their origin from the inner layer of ectoderm. The condition is similar to that in the lung fishes, *Protopterus* and *Lepidostren*, but differs from the cases described in Anura where the glands develop from the superficial layer of ectoderm.

In the case of *Hyperoplus bebe* and *Sarcodactylus odae* the two African fishes described by Budgett, the former has three pairs of cement glands as in *Etroplus* and the latter a large single median one on the dorsal side of the head. The larvae in both cases hang suspended by mucous threads secreted by the cement organs. The arrangement and structure of the glands in active condition of *Pterophyllum* are exactly as in *Etroplus*. Both behave in a similar manner by resting at the bottom. As they belong to the same family, Cichlidae, in all probability, the origin of the organs is the same.

The fact that the cement organs are meant solely to help the animals to attach themselves to some object without being carried off by water currents, shows why they are found only in the aquatic larvae of vertebrates and not in those of terrestrial ones, where we find all traces of them omitted in their ontogeny. Only very rarely do we come across fishes having these structures. Among the Amphibians the larval Anura generally possess them. They are completely unrepresented in the Cyclostomes, Elasmobranchs and Apoda and absent in their typical form in the Urodela. But in the last group, many possess two rod-like structures one on either side, known as balancers, the true homology of which is doubtful. They have glandular tips and develop as ectodermal projections from outside the mandibular arch and serve as organs of support and attachment (Harrison, 1925). A branch of the hyoidean artery supplies these structures and the blood is returned to the jugular veins (Maurer, 1888). Goodrich (1930) has suggested that the balancers are homologous to the cement organs of Anura and judging from their development, function and early atrophy, one is disposed to accept this view. The stalked condition will have to be considered as a secondary feature developed in the course of evolution. This does not appear in any way improbable when we find the cement organs of larval *Acipenser* getting modified into barbels in the adult. Also, just as the cement organs degenerate in fishes when the pectoral fins are developed, the balancers of Urodèles atrophy as soon as the fore-limbs are sufficiently developed to support the head.

The origin of the cement organs as endodermal pouches in the Crossopterygii and Kerr (1906) to compare them with the pre-mandibular head cavities of other vertebrates. Reighard and Phelps (1908) are of opinion that these glands are homologous with the anterior gut pouches of Elasmobranchs, which in turn, have been homologized with the anterior gut pouches of Amphiomorpha, one of which is converted into the elaiotroch organ (Neal, 1898 and van Wyhe, 1914). It is very doubtful if this view could be accepted from the fact that in the Teleostei, Diplopoda and Anura these organs are of ectodermal origin, though the condition in Crossopterygii is in support of it. This would mean that the organs have arisen independently in the different groups of vertebrates as suggested by Eycleshymer and Wilson (1908) who also support the suggestion of Balfour with regard to the origin of the barbels in Teleosts as seen from the fate of the cement organs in *Acipenser*. As against this view Kerr (1919) is of opinion that during ontogeny certain phases in the development of the organs have been slurred over or even omitted giving rise to a condition in Diplopoda and Anura where the organs were known to take their origin from the ectoderm.

But instances, wherein different or intermediate conditions in origin within the same group, which would have gone in favour of this theory, are lacking. Still the fact that in Teleostei and Diplopoda, the two intermediate groups of vertebrates possessing cement organs, they are known to originate from the *inner* layer of ectoderm whereas in the more highly evolved Anura the secretory cells of the glands are derived from the *superficial* layer of ectoderm, cannot be without some phylogenetic significance. However, more work on the cement glands in fishes, especially of Teleosts, is required before this and allied questions can be solved in a satisfactory way.

#### *Summary*

1 There are three pairs of cement glands in *Etraphus maculatus* situated on the dorsal side of the head, the anterior-most pair being smaller than the posterior two pairs.

2 They produce mucus which help to attach the larva to the bottom till it is capable of independent swimming movement.

3 The gland rudiments first appear when the embryo is thirty-eight hours old as thickenings of the inner layer of the ectoderm.

4 The gland cells soon enclose a cavity into which the secretion is poured and by the time of hatching the outer layer of ectoderm over the cavity is lost.

5 The glands grow in size and remain active till the fourth day after which they begin to degenerate. They shrink in size, get flattened out and the superficial layer of ectoderm grows over them. They disappear by the seventh day.

#### *Acknowledgment*

I wish to thank Professor R. Gopala Atavar, Director, University Zoological Research Laboratory, Madras, for his constant help and valuable suggestions during the course of this work. My thanks are also due to the University of Madras for awarding me a Research Studentship.

#### *BIBLIOGRAPHY*

- 1 Agassiz, A\* .. "The Development of *Lepidosteus*," *Impr Acad Arts & Sci Proc (N.S.)*, 1879, 14
- 2 Assheton, R. .. "Notes on the Citation of the Ectoderm of the Anuran Embryo," *Quart Journ Micro Sci* 1896, 38,
- 3 ——— .. "The Development of *Gymnarchus niloticus*," The work of John Samuel Budgett (*Budgett Memorial Volume*), Cambridge, 1907a
- 4 ——— .. "Report upon sundry Teleostean Eggs and Larvae from the Gambia River," *ibid*, 1907b

5 Balfour, F. M. *Treatise on Comparative Embryology*, London, 1881

6 Balfour, F. M., and Parker, W. N. "The Structure and Development of *Lepidosteus*," *Phil Trans Roy Soc London*, 1882, 173, 2

7 Bhaduri, J. L. "The Anatomy of the Adhesive Apparatus in the tadpoles of *Rana aythana* Günther, with special reference to the adaptive modification," *Trans Roy Soc Edin*, 1935, 58

8 Bridge, T. W. "Fishes," *The Cambridge Natural History*, London, 1904

9 Budgett, J. S. "On the Breeding Habits of West African Fishes, with an Account of the External Features in the Development of *Protopterus annectans* and a Description of the Larva of *Polypterus lopezi*," *Trans Zool Soc London*, 1901a, 16, 2

10 ———— "The Habits and Development of some West African Fishes," *Proc Camb Phil Soc*, 1901b, 11

11 Dean, Bashford "On the Larval Development of *Annia eiselti*," *Zool Jährl*, 1896, 9

12 Eycleshymer, A. C., and Wilson, J. M. "Adhesive Organs of *Annia*," *Biol Bull*, 1908, 14

13 Frankenberg, Z. "Über der Morphologische Bedeutung der Haftorgane bei den Larven einiger niederer Vertebraten," *Zool Anz*, 1926, 69

14 Gadow, H. "Amphibia and Reptiles," *The Cambridge Natural History*, London, 1920

15 Goedert, E. "Die Entwicklung des Mundes und der Mundhöhle mit Drüsen und Zunge, die Entwicklung der Schwimmblase der Lung und der Kehlkopfes der Wirbeltiere," *Hertwig's Handbuch der vergleichenden und Experimentellen Entwicklungsgeschichte der Wirbeltiere*, Jena, 1906, 2, 1

16 Goodrich, E. S. *Development and Structure of Vertebrates*, London, 1930

17 Harrison, R. G. "Development of Balancers in *Ambystoma* studied by the method of Transplantation and in relation to the connective tissue problem," *J Exp Zool*, 1925, 41

18 Hora, S. L. "Development and Probable Evolution of the suctorial disc in the Tadpoles of *Rana aythana* Günther," *Trans Roy Soc Edin*, 1932, 57

19 Kerr, J. G. "The External features in the Development of *Lepidomren paradoxus*, Fitz," *Phil Trans Roy Soc London*, 1900(B), 46

20 ———— "The Development of *Lepidomren paradoxus* III. The Development of the skin and its derivatives," *Quart Journ Musc Sci*, 1903, 44

21 ————\* "The Development of *Polypterus*," *Proc Roy Phys Soc Edin*, 1906, 19

22 ———— "The Development of *Polypterus senegalus* Cuv," The work of John Samuel Budgett (Budgett Memorial Volume), Cambridge, 1907.

23 Kerr, J. G. \* "Normal Plates in the Development of *Lepidostoma paradoxa* and *Protopterus aenescens*" Keibel's *Vorlesungen zur Entwicklung Würbels*, Jena 1909 3

24 ————— Text-Book of Embryology 2, London, 1919

25 Keibel, F. "Die Entwicklung der ausseren Körperform der Wirbeltiere embryonen, insb. während der menschlichen Embryonen aus der ersten 2 Monaten," Hertwig's *Handbuch der Entwicklung Würbels*, Jena, 1906

26 Kupffer, C. W. \* "Die Entwicklung des Kopfes von *Acipenser sturio*" *Studien zur vergleichenden Entwicklungsgeschichte des Kopfes der Knochenfische* I. *Heft*, München u. Leipzig, 1893

27 Lieberknecht, I. "Über die Haftorgane bei Jungen von *Pterophyllum scalare* F. Abl." *Zool. fuz.* 1912 97

28 Maurer, F. \* "Die Keimen und ihre Gefäße bei Anuren und Urodelen Amphibien und die Umwandlungen der beiden ersten Arterienbogen bei Tektostern," *Morph. Jahrb.* 1888, 14

29 Neal, H. W. "The segmentation of the Nervous System in *Squalus acanthias*" *Bull. Comp. Zool.* 1891 31, 7

30 Panikkar, N. P. "Notes on the Cichlid Fishes of Madras, *Utroplus maculatus* and *E. maculatus*" *Mad. Fish. Bull.* 1920 12

31 Phelps, J. "Development of the Adhesive Organ of *Amaia*" *Science N. S.* 1899 9

32 Reighard, J., and Phelps, J. "The Development of the Adhesive Organ and Head Mesoblast of *Amaia*" *J. Morph.* 1908 19

33 Sawadsky, A. M. \* "Die Entwicklung des larvalen Haftapparates beim sterlet (*Liparis ruthenus*)," *Amat. Anz.*, 1911, 40

34 Stohr, P. "Über die Haftorgane des Anuren Larven," *S. B. Phys. med. Ges. Wurzburg* 1881

35 Sundara Raj, B. "Notes on the Fresh water Fish of Madras," *Rec. Ind. Mus.*, 1916, 12

36 Thiele, J. "Der Haftapparat der Batrachienlarven" *Zeit. wiss. Zool.* 1887, 46

37 Wyhe, J. W. v. "On the Metamorphosis of *Amphibius laevis* (L.)" *Ver. K. Akad. Amsterdam* 1914, 16, 2

38 Willey, A. "Notes on the Fresh-water Fisheries of Ceylon," *Spot. Zts.*, 1911, 7

\* The references marked thus \* have not been directly consulted by the author



# REPORT ON SOME NEMATODE PARASITES OF KABUL, WITH DESCRIPTIONS OF NEW SPECIES.

BY S A AKHTAR

(From the Department of Biology, Faculty of Medicine, Kabul)

Received October 18, 1937

(Communicated by Dr M. B. Mirza, D.Sc.)

The present paper deals with some Nematode parasites of different animals of this place, which the writer has come across during the last few months. One new species of the genus *Ascaridia* Dujardin, 1845, one of the genus *Subulura* Molin, 1860, and one of the genus *Tachygonetria* Wedl, 1862, have also been described in it.

The writer expresses his grateful thanks to Dr M. B. Mirza, Director, Zoological Laboratories, Muslim University, Aligarh, for kindly going through the manuscript. Sincere thanks are also due to Dr B. Prashad, Director, Zoological Survey of India, for facilities in consulting literature and examining certain specimens in the collection of the Indian Museum at Calcutta.

## 1 *Ascaris lumbricoides* Linnaeus, 1758

Only one male specimen of this species, 248 mm long and about 3 mm thick, was obtained from a man.

## 2 *Ascaridia razia* n. sp.\*

A pair of the worms of genus *Ascaridia* Dujardin (1845) was recovered from the intestine of a wild pigeon (*Columba livia*).

*Morphology*—Body is white and cuticle with transverse striations. Anterior end is with three large and well-defined lips, each bearing two cephalic papillae. Cervical alae are present. Oesophagus is club-shaped and is without a posterior bulb.

*Male*—Transverse striations over the body are 0.015 mm apart. There is a pre-anal sucker and a few rounded or oval bodies are found in front of the sucker within the body cavity. There are oblique muscles near the sucker, all terminating in the centre of the sucker. Caudal alae are small and narrow beginning in front of the posterior end of the body and reaching up to the level of the pre-anal sucker.

\* The species is named after the writer's daughter Razia Khatoon.

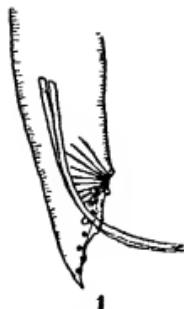


FIG. 1. *Ascidiella racia*. Posterior extremity of male, laterally

There are only nine pairs of big caudal papillæ, four pairs of which are pre-anal, one adanal, and four pairs are post-anal. Spicules are two, subequal and alate. There is no gubernaculum. Tail ends in a conical spine and is slightly incurved.

*Measurements* —

Length of the worm	.	.	20.812	mm
Maximum thickness	.	.	0.787	"
Length of lips	.	.	0.045	"
Diameter of head	..	..	0.146	"
Length of oesophagus	..	.	1.575	"
Diameter of pre-anal sucker (lengthwise)	.	.	0.135	"
Distance between pre-anal sucker and cloaca	.	.	0.200	"
Length of spicules	.	.	1.732 and 1.687	"
Length of tail	..	.	0.450	"

*Female* — Transverse striations over the body are 0.022 mm apart. Vulva is a little behind the middle of the body. Tail is slightly truncate at the level of the cloaca ventrally and then compressed dorso-ventrally ending in a blunt point.

*Measurements* —

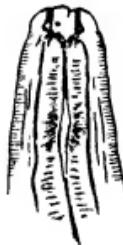
Length of the worm	..	..	..	32.075	mm
Maximum thickness	.	.	..	1.132	"
Length of cervical alæ	..	.	..	2.700	"
Length of oesophagus	..	.	..	2.002	"
Distance between vulva and posterior end	.	.	..	15.480	"
Length of tail	..	..	..	0.825	"
Size of egg	..	..	..	0.180 $\times$ 0.157	"

*Ascaris rasia* differs from all its allied species in different measurements. It also differs from *A. columbe* (Gmelin, 1790) Travassos, 1913, in the diameter of head, distance of body striations, lengths of tails, diameter of pre-anal sucker and in the distance between the pre-anal sucker and the cloacal aperture. The new species *A. rasia* resembles *A. stroma* (v Linstow, 1899) Railliet and Henry, 1914, in having comparatively small number of big caudal papillæ, but differs much from it in the length of oesophagus (about 1/13 of the length of the body) which is longer, the length of the tail in male (1/46 of the body length) which is shorter, and in the length of the tail of female (1/38 of the body length) which is longer. Oblique muscles of the sucker are strong and they terminate together in the centre of the sucker. There are only nine pairs of caudal papillæ, four pairs of which are pre-anal, one pair adanal (biggest of all the papillæ) and four pairs are post-anal. Moreover the eggs of *A. rasia* are larger than those of all the other species of *Ascaris*, also larger than the eggs of *A. stroma*, whose eggs are exceptionally large in the genus. All these are salient characters to differentiate *A. rasia* from all other species of the genus.

### 3 *Subulura kabulana* n. sp.

Five specimens (3 males and 2 females) of the genus *Subulura* Molin, 1860, were recovered from the intestine of an orange-billed pheasant (*Tetraogallus* sp.).

**Morphology**—The body is yellowish white, cuticle striated coarsely and the anterior portion of the body is curved in both the sexes. Anterior end of the body is blunt and lateral alæ are present. The buccal capsule is with chitinous walls and has three teeth at its depth one being dorsal and two sub-ventral. There is a pharynx at the anterior portion of oesophagus.



2

FIG. 2. *Subulura kabulana*. Anterior extremity, laterally.

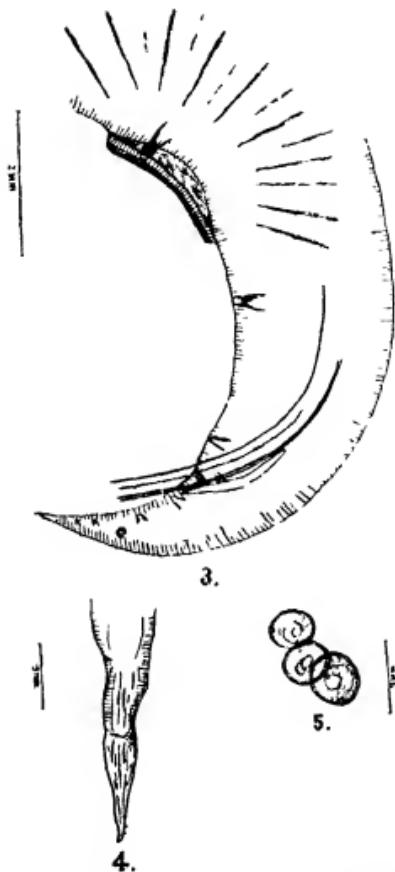


FIG. 3 *Subulura kabulana* Posterior extremity of male, laterally  
 FIG. 4 " " " female, laterally  
 FIG. 5 " " Eggs

which is somewhat thicker. Oesophagus is club-shaped, jointed to a posterior bulb by a narrow portion. The bulb is subglobular

**Male**—The posterior portion of the male is curved and is with a terminal spine. Small caudal alae as well as pre-anal sucker are present. The sucker is with a chitinous rim surrounded by strong oblique muscles. The interior of sucker is somewhat deep and its surface is with fine longitudinal striations. The spicules are equal, curved, alate and fine-pointed. Gubernaculum is long and there are ten pairs of caudal papillæ, three of which are pre-anal, two adanal and five pairs are post-anal.

**Measurements**—

Length of the worm	.	11 587 mm	9 593 mm
Maximum thickness		0 458 ..	0 450 ..
Diameter of head		0 090 ..	0 067 ..
Length of buccal capsule		0 058 ..	
Width of buccal capsule at post end		0 031 ..	
Height of teeth	.	0 018 ..	
Length of oesophagus with bulb		1 192 ..	1 170 ..
Diameter of pre-bulbular swelling of oesophagus		0 135 ..	0 090 ..
Length of the bulb		0 225 ..	0 205 ..
Diameter of the bulb	.	0 225 ..	0 215 ..
Length of pre-anal sucker		0 210 ..	0 202 ..
Distance between pre-anal sucker and cloacal aperture		0 450 ..	0 212 ..
Length of spicules		1 350 ..	1 408 ..
Width of spicule		0 036 ..	
Length of gubernaculum		0 135 ..	
Length of tail		0 256 ..	0 283 ..

**Female**—The tail of female is compressed dorso-ventrally and is bluntly spear-shaped. Vulva is a little in front of middle of the body at about 4/5 of the length. Eggs are with fully-formed embryo and sub-globular.

**Measurements**—

Length of the worm		16 583 mm	19 467 mm
Length of buccal capsule		0 063 ..	
Width of buccal capsule at post end		0 036 ..	
Height of teeth	.	0 015 ..	
Length of oesophagus with bulb		1 237 ..	1 609 ..
Width of pre-bulbular swelling of oesophagus		0 202 ..	
Length of the bulb		0 225 ..	
Distance of vulva from anterior end		7 448 ..	8 212 ..
Length of tail	.	1 193 ..	1 450 ..
Width of caudal spear-shaped portion		0 225 ..	
Size of eggs	..	0 063 — 0 072 × 0 045 — 0 058 mm.	

*Subulura kabulanus*, discovered by the writer, differs from all other species of the genus in its measurements and structure. Through the kindness of Dr B. Prashad, the writer could examine two or three species of *Subulura*, particularly *S. galloperdixis* Baylis and Daubney, 1922, in which oblique muscles are not so strong and the margin of the sucker is not well-defined. *S. kabulanus* appears to resemble, to some extent in its general form and structure as well as in the number of caudal papillæ, *S. andersoni* (Cobbold, 1876) Railhet and Henry, 1914, but differs markedly from it in its measurements, showing its spicules to be longer, gubernaculum smaller, sucker longer, oesophagus smaller and the distance between the pre-anal sucker and cloacal aperture being less than in the named species. Above all, *S. kabulanus* possesses a chitinous rim of the sucker of quite peculiar shape, appearing to be made up of upright bars placed side by side. These characters are sufficient to differentiate it from all other allied species and in this connection it is suggested that the presence or absence of chitinous rim of pre-anal sucker be added to the generic characters of *Subulura*.

#### 4 *Tachygonetria microstoma* Drasche, 1884

Two male specimens of this species were obtained from the caecum of a tortoise (*Testudo* sp.). The specimens were, however, found to differ from the published account of the species in the following respect —

Maximum thickness	.	0 240 mm
Length of spicule	.	0 102 ..
Length of tail	..	0 220 ..

#### 5 *Tachygonetria inflatocervix* n. sp.

Worms of the genus *Tachygonetria* Wedl, 1862, were recovered from the caecum of a tortoise (*Testudo* sp.)

**Morphology** — Small worms, mouth surrounded by six inconspicuous lips and head bearing four big cephalic papillæ. Cervical cuticle is inflated symmetrically, forming a projecting cuticular collar, covering four or five annuli in the middle of the collar. Cuticle of the body is coarsely striated, sometimes except on the dorsal surface and the tail, where the striations are fine. Vestibule is short and without any chitinous armature. Oesophagus is cylindrical and long with posterior bulb containing a valvular apparatus. Excretory aperture is behind the bulb. Intestine is simple and without diverticulum. Lateral flanges are absent.

**Male** — The body is obliquely truncate ventrally at the level of the cloaca. There is only one spicule and that is acicular. Gubernaculum present. Tail or the posterior portion of the body is coiled and is with narrow alæ.

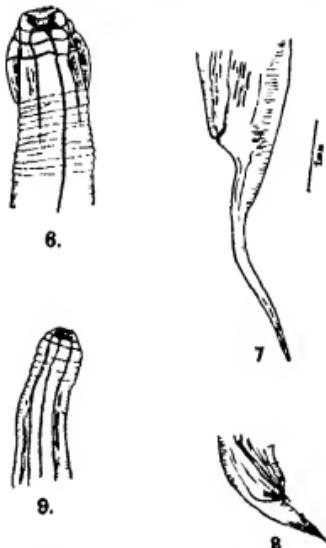


FIG. 6 *Tachygonetra inflatocervix* Anterior extremity  
 FIG. 7 " " Posterior extremity of female, laterally  
 FIG. 8 " " male laterally  
 FIG. 9 " " Anterior extremity of male

Tail is conical and fine-pointed, bearing three pairs of pericloacal papillæ—one pair of pre-anal, one pair latero-ventral and one pair of adanal papillæ. In addition to these there is a pair of voluminous papillæ on the middle of the tail.

*Measurements*—

Length of the worm ..	2 844 mm
Maximum thickness ..	0 187 ..
Diameter of mouth ..	0 225 ..
Length of vestibule ..	0.009 ..
Diameter of cervical collar ..	0 033 ..
Length of oesophagus with bulb ..	0 603 ..
Length of spicule ..	0 117 ..
Length of gubernaculum ..	0 031 ..
Length of tail ..	0 072 ..

*Female*—Cervical collar is well-developed and fully-distended covering up to twelve annules, often another collar develops behind the former. Vulva is behind the middle of the body and the tail is conical, long and tapering to a point. Uteri are two, eggs large, numerous and ellipsoidal and are segmented at deposition.

*Measurements*—

Length of worm	2 970 mm	4 905 mm
Thickness	0.250 "	0.382 "
Diameter of mouth	0.040 "	
Diameter of collar	0.119 "	0.202 "
Length of vestibule	0.027 "	
Nervous ring behind the mouth	0.234 "	
Length of oesophagus with bulb	0.900 "	1.170 mm
Vulva from posterior end	1.427 "	2.430 "
Length of tail	0.346 "	0.517 "
Size of eggs	0.076 — 0.184 x 0.081 — 0.090 "	

*Tachygonetria inflatocervix*, discovered by the writer, differs from all species of *Tachygonetria* in its measurements, it also differs in the shape of its head. Cervical collar, which is not well developed in males, is fully distended in females and is often followed by another collar of the same type. It only resembles *T. microstoma* (Drasche, 1884) and *T. uncinata* (Drasche, 1884), in having a conical tail, bearing a pair of voluminous papillæ, but differs markedly from them in the shape and size of tail, particularly in female and also in the position of the last pair of voluminous papillæ, which is situated on the middle of the tail of male, instead of on the posterior third. These characters are sufficient to create a new species and in this connection it is suggested that "a pair of voluminous papillæ on the middle or posterior third of the tail" be added to the generic characters of *Tachygonetria*.

6 *Chabertia ovina* Railliet and Henry, 1909

Five specimens of this species were obtained from the intestine of a sheep

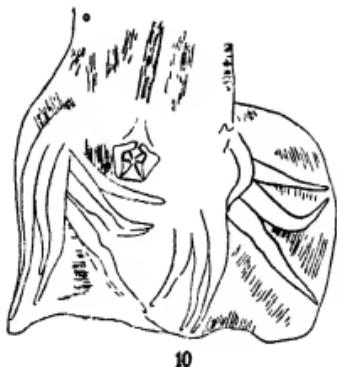
7 *Ostertagia marshalli* Ransom, 1907

Many specimens were obtained from abomasum of a sheep

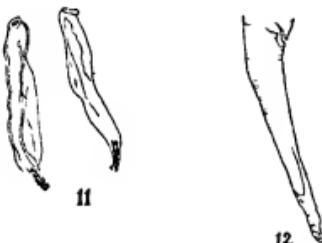
8 *Ostertagia tricuspidis* Marotel, 1910

The species was originally described by Marotel in 1910, from Lyons in France. After that probably the species has not been reported, as yet, from any part of the world. It is interesting to record its occurrence for

the first time in Asia. After dissecting a large number of intestines of sheep, many worms of this species were recovered from one case only. These worms are yellowish brown in colour and the cuticle is finely striated transversely. There are twenty-four longitudinal ridges over the body. The cuticle of the anterior end is dilated and the buccal cavity is very small.



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FIG. 10 *Ostertagia tricuspid* Posterior extremity of male, showing telamon  
 FIG. 11 " " Spicules  
 FIG. 12 " " Posterior extremity of female, laterally

**Male**—The pre-bursal papillae are present and there is no accessory piece. The spicules are equal and are divided into three in the posterior fifth of their lengths. The median process is blunt at the tip while others are pointed. There is a pentagonal structure (telamon) at cloaca.

**Measurements —**

Length of the worm	..	..	10.249 mm
Diameter of head	..	..	0.018 ..
Length of oesophagus	..	..	0.819 ..
Length of spicules	..	..	0.252 ..

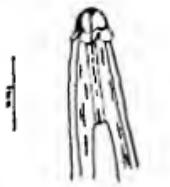
*Female* — Vulva is situated at about posterior fifth of the body and is with a flap on the anterior margin. Tail is digitiform with fine transverse lines a little in front of the tip. Five small papillæ-like dots appear to be on the tip of the tail.

**Measurements —**

Length of the worm	..	..	15.390 mm
Diameter of head	..	..	0.922 ..
Length of oesophagus	..	..	0.922 ..
Vulva from posterior end	..	..	3.240 ..
Length of flap	..	..	0.285 ..
Length of tail	..	..	0.315 ..
Size of eggs	..	..	0.166 x 0.076 ..

**9 *Parabronema* sp**

Only one female of this parasite was found in the beginning of the small intestine of a sheep. The anterior extremity is provided with dorsal and



13



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FIG. 13. *Parabronema* sp. Anterior extremity.

FIG. 14. " Posterior extremity

ventral cuticular shields and is ornamented with six horse shoe-shaped cordon<sup>s</sup>, lateral flanges are absent. The buccal capsule is long and its posterior portion is cylindrical without annular or spiral thickenings and the oesophagus is also without any swelling. The tail is short, bluntly conical and dorsally curved. Anterior part of the body is reddish brown and the remaining portion yellowish brown. Excretory aperture is near the beginning of the oesophagus, and the vulva is behind the oesophagus.

**Measurements —**

Length of the worm	25 437 mm
Length of buccal capsule	0 175 ..
Length of oesophagus	2 340 ..
Distance of excretory pore from anterior end	0 220 ..
Distance of vulva from anterior end	7 200 ..
Distance of vulva from posterior end of oesophagus	4 685 ..
Length of tail	0 180 ..

From the above description the specimen appears to belong to the species *Parabronema skryabinii* Rasovska, 1924, but as it differs from *P. skryabinii* in the position of its vulva and as the male is wanting its correct specific determination is hardly possible.

**10 *Polymorphus boschadis* Schrank, 1788**

Two specimens of this species were found in the intestine of a wild duck (*Anas* sp.)

Type-specimens of the new species as well as of certain others have been deposited in the Museum of the Zoological Laboratories, Muslim University, Aligarh, *Ascaridia razia* under No 935, *Subulura kabulanus* under No 936, and *Tachygonetria inflatocervix* under No 937.

**REFERENCES**

Baylis, H. A., *A Synopsis of the Families and Genera of Nematoda*, 1926  
 \_\_\_\_\_ and Daubney, R., "Report on the parasitic Nematodes in the collection of the Zoological Survey of India," *Mem. Ind. Mus.*, 1922, 4, 7  
 \_\_\_\_\_, *A Manual of Helminthology*, 1929  
 \_\_\_\_\_, *Fauna of British India, Nematoda*, 1936, 1  
 Cram, E. B., "Bird parasites of the Nematoda, Strongylida, etc.," *U. S. Nat. Mus.*, 1927, Bull. 140  
 Hall, M. C., "Nematode parasites of Mammals of the order Rodentia, etc.," *Proc. U. S. Nat. Mus. Washington*, 1916, 50  
 Mirza, M. B., "Subulura hindii n. sp. A New Nematode Parasite of *Sciurus palmarum*," *Proc. Ind. Acad. Sci.*, Bangalore, Feb 1936, 3, 2  
 Thapar, G. S., "Studies on Oxyurid Parasites of Reptiles," *Jl. Helminth.* 1925, 3, 3-4  
 York, W., and Maplestone, P. A., *Nematode Parasites of Vertebrates*, 1926.

## STUDIES ON THE TREMATODE PARASITES OF BIRDS.

### Part II. Morphology and Systematic Position of Some New Blood-Flukes of the Family Schistosomidae.

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Received September 28, 1937.

[Communicated by Dr G S Thapar, M.Sc., M.B., B.S. (London)]

THE discovery of the first genus of this family was originally made by Bilharz in 1851 when he recovered some worms from the mesenteric veins of a native of Cairo. This was reported by Bilharz in 1852 who named it *Distomum haematobium*. Later this genus was reported by other workers under different names, viz., *Gynaecophorus* Diesing, 1858, *Bilharzia* Cobbold, 1859, *Thecosome* Moquin-Tandon, 1860. The name "Distomum" for this blood-fluke was a misnomer and was replaced by *Schistosoma* by Weinland in 1858, and although later workers tried to immortalize the name of Bilharz, the discoverer of this blood-fluke, by naming it after him, the name had to be given up in view of priority of *Schistosoma*.

Looss (1899) created the family Schistosomidae for the genus *Schistosoma* Weinland (1854) and added the second genus *Bilharziella* to the family which was described earlier by Kowalewski (1895) under the name of *Bilharzia polonica*. Odhner (1910) described *Gigantobilharzia* and later (1912) added another genus *Ornithobilharzia*. Johnston (1917) reported the genus *Austrobilharzia* from Australia. Three years later Skrjabin and Zakharow (1920) removed an earlier form *Bilharziella pulvifulenta* Brau, 1901, to their new genus *Dendrobilharzia*. In the same year they recorded another new genus *Trichobilharzia*. Tanabe (1923) created the genus *Schistosomatium* for his experimentally obtained forms in white rats and mice. Travassos (1923) added the genus *Macrobilharzia* to the family. The latest work on the family is that of Price (1929) where he recognizes all these forms, except *Macrobilharzia* of Travassos which he regards as a synonym of *Ornithobilharzia*, and creates three new genera *Microbilharzia*, *Heterobilharzia* and *Paraschistosomatium*. He also divides the family Schistosomidae into two sub-families, *Schistosominae* and *Bilharziellinae*. Subsequently (1931) the

same author in a brief note revives the genus *Macrobilharzia* and suppresses one of his newly created genera, viz., *Paraschistosomatium* as a synonym of *Macrobilharzia*. The family *Schistosomidae* is of recent occurrence in India. Montgomery (1906) described *Schistosoma indica*, from mammals. Christophers and Stephens (1905) discovered an egg of *Schistosoma spindalis* in the urine of a Madrasai. The larval stages have been recorded by Soparkar (1921) and Sewell (1923). Jasper (1923) mentions that several cases of infection occurred in the neighbourhood of Hyderabad after the return of infected troops from Egypt.

From birds, however, the family has been very recently reported in India. The earliest case, so far as the writer is aware, is from Rangoon when Gogate (1934) obtained two immature male specimens of blood-flukes from wild ducks. Although he has neither given diagrams nor adequate description of his forms, he tentatively refers them as *Ornithobilharzia* sp. under the subfamily *Schistosominae*. The writer, in the course of his investigations, has also come across two cases of blood-fluke infection in birds. These forms, which are being described in the present communication, come under the subfamily *Bilharziellinae* Price, 1929, but owing to certain peculiar characters presented by them necessitate the establishment of the subfamily *diagnosis*.

*Subfamily Bilharziellinae* Price, 1929, emended

*Schistosomidae* Suckers present or absent. Gynacophoric canal absent or imperfectly formed or sometimes well developed. Paired intestinal caeca short, uniting cephalad of the middle of the body, common caecum long, with or without lateral dendritic branches. Testes numerous and situated behind the caecal union along the course of the common caecum. Uterus short, containing a single egg.

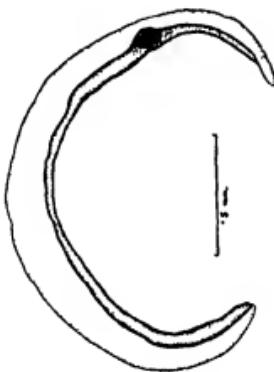
Type genus—*Bilharziella* Loos, 1899

*Chinhuta indica* N G N Sp

A very large number, exceeding 200, of specimens of this species were obtained from the main blood vessels and internal organs of the common teal, *Nettion crecca*. The bird which was captured at the Chinluti Lake, about 8 miles from Lucknow, was kept under observation but died next morning. The post-mortem examination revealed a very heavy infection with *Chinhuta*. There were signs of lesion formation in liver, kidney and even lungs.

The animals were inactive showing little movement when liberated in salt solution. They appeared white in colour and were sticking fast to the tissues by their powerful ventral sucker which is a cup-shaped structure with slightly pedunculated base.

*Male*—It is a long, thin animal with blunt anterior end and tapering posterior end. The length of the specimen is 2.95 mm and the maximum breadth which is a little behind the caecal union is .375 mm. The lateral edges of the body are rolled inwards to form a deep gynæcophoric groove which extends right from the hinder end of the oral sucker up to the posterior end.



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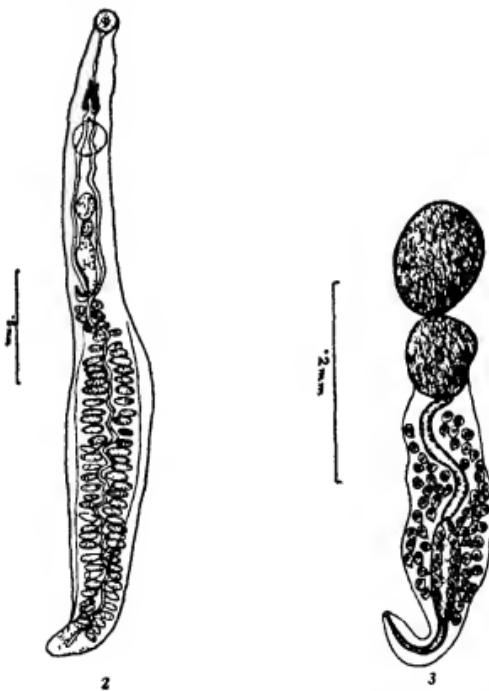
TEXT FIG 1 *Chinhuis indica* male—Lateral view showing the Gynæcophoric groove.

The circular oral sucker lies ventrally and is rather weakly developed. It measures .1 mm in diameter. The ventral sucker, which is muscular and very powerful, is larger than the oral sucker and measures 15 mm in diameter and lies at a distance of 5 mm from the anterior end.

The mouth leads into a long thin oesophagus which measures .375 mm. There is no pharynx in these forms. There are a large number of unicellular glands surrounding the base of the oesophagus and they appear to be arranged in a grape-bunch manner. It is rather difficult to surmise the exact nature of these glands but probably they may be producing some secretions which prevent the clotting of blood on which these parasites feed.

The oesophagus divides into two intestinal caeca at its posterior end just in front of the ventral sucker. There is a small caecal projection directed forwards just at the intestinal bifurcation which probably is a representative of the H-shaped caecal bifurcation seen in some other blood-flukes. The two intestinal caeca run dorsal to the ventral sucker and after a brief course

join together in the pre-equatorial region to form a common caecum at a distance of .625 mm behind the ventral sucker and 1.275 mm from the anterior end. The common caecum which is longer than the separate intestinal caeca runs behind in a zigzag manner ending blindly at a distance of about 1 mm from the posterior end. The common caecum shows thickenings of its walls at various places which also become sharply pointed



TEXT FIG. 2. *Chikungula indica* male—Ventral view showing general anatomy

TEXT FIG. 3. Cirrus sac showing bilobed vesicula seminalis, prostate cells and pars prostatica—ventral view

The testes, which vary between 70-80 in number, lie on the two sides of the common caecum, beginning just behind the caecal union and ending

a little in front of the caecal end. They are oval bodies of variable size, the largest measuring

$125\text{ mm} \times .04\text{ mm}$  and the smallest measuring  $.05\text{ mm} \times .025\text{ mm}$ . The vesicula seminalis is a lobed structure, situated behind the ventral sucker. It consists of a large oval lobe measuring  $117\text{ mm} \times .0875\text{ mm}$  and a small lobe, irregular in shape measuring  $.087\text{ mm} \times .07\text{ mm}$  and joined by a very narrow constriction in the middle. The smaller lobe of the vesicula seminalis is enclosed within the cirrus sac which is a long cylindrical body containing the ejaculatory duct, pars prostatica and a large number of prostate gland-cells. The terminal end of the cirrus sac is bent and opens at the genital pore on the left side of the median line just in front of the level of the caecal union. A small S shaped duct leads from the vesicula seminalis to open into an elliptical pars prostatica measuring  $1\text{ mm}$  in length. A small duct terminating in the cirrus arises from the pars prostatica, takes a bent course and opens out at the genital pore.

**Female**—The female is much smaller in size than the male. It measures  $1.8\text{ mm}$  in length and has a maximum breadth of about  $19\text{ mm}$  behind the ovary in the level of the vitellaria. The body is flat and narrower towards the two ends. It does not show any inrolling of its lateral edges. The oral sucker is ventral and measures  $.04\text{ mm}$  in diameter. The ventral sucker is strongly developed as in the case of the male and measures  $.075\text{ mm}$  in diameter.

The mouth leads into a long oesophagus which measures  $.225\text{ mm}$  and is surrounded at its base by the unicellular oesophageal glands. It bifurcates into the two intestinal caeca at its posterior end, about  $.075\text{ mm}$  in front of the ventral sucker. The two intestinal caeca run for a short distance and unite to form a common caecum at the level of the anterior end of the ovary at a distance of  $.25\text{ mm}$  from the ventral sucker and about  $.675\text{ mm}$  from the anterior end. The common caecum follows a zigzag



*Cchinhuta indica* female—ventral view showing general anatomy.

course behind, shows peculiar thickenings of its wall and ends at a distance of .05 mm from the posterior end.

The ovary is an elongated sinuous body, broader anteriorly and tapering posteriorly. It lies at a distance of .27 mm behind the ventral sucker and measures .155 mm in length and .05 mm in maximum breadth. The oviduct arises from the right side of the broad anterior end of the ovary and after a sharp curve enters the ootype. The ootype is surrounded by small unicellular shell-glands. The short straight uterus arises as a broad tube in front of the ootype and ends at the genital pore, .04 mm behind the ventral sucker. No eggs were found in the uterus.

The vitellaria consist of distinct and large follicles which extend right behind the ovary to the posterior end. They lie on the two sides of the common caecum and pour their yolk by narrow ducts into a large U-shaped vitelline reservoir which lies at the level of the ovary and leads by a thin duct to open into the ootype.

To sum up, the genus *Chinhula* is characterised thus—

Male with a well-developed gynaecophoric canal, extending from a little behind the oral sucker up to the posterior end, female with flattened body and smaller than the male. Suckers present. Oesophagus provided with unicellular oesophageal glands. The two intestinal caeca unite a little in front of the middle of body, common caecum long, without lateral branches but provided with angular thickenings of its wall. Testes vary between 70-80 in number and extend from the caecal union to the posterior end of the animal. Cirrus pouch well developed, enclosing a part of vesicula seminalis, prostate gland-cells, pars prostatica and the cirrus. The terminal end of the cirrus sac is slightly bent to the left side. The male genital pore lies slightly to the left of the median line near the middle of the body. The ovary is elongated and sinuous, situated at the level of the caecal union. Uterus short and straight. The female genital opening is a little behind the acetabulum. Vitellaria situated on the sides of the common caecum, in distinct follicles. A vitelline reservoir is present.

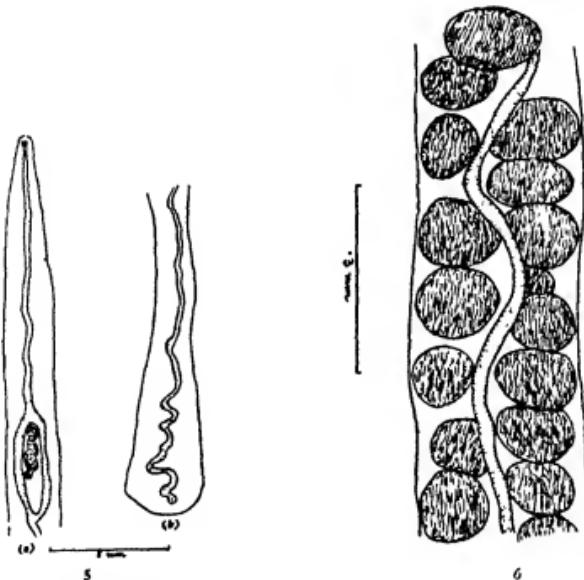
Type species—*Chinhula indica*

**Remarks**—The new genus differs from the members of the subfamily Bilharziellinae in having an extremely well-developed gynaecophoric canal in which feature it stands near the subfamily Schistosominae. But it differs from the subfamily Schistosominae in having cephalad union of the intestinal caeca and testes behind the caecal union, features in which it closely resembles the subfamily Bilharziellinae. The genus, in fact, forms a connecting link between the two subfamilies. The nearest ally of the new

genus is the genus *Bilharziella* with which it shows several features of resemblance. It differs, however, from the genus *Bilharziella* in having an extremely well-developed gynaecophoric canal, in the possession of the oesophageal glands, bent terminal end of the cirrus sac and in the position of the female genital opening. It is, therefore, considered necessary to create a new genus which is designated *Chinhula* owing to its recovery from a bird caught at the lake, Chinhut.

*Gigantobilharzia egreta* N. Sp.

Only one male specimen of this parasite was obtained from the renal vein of the cattle egret, *Bulbulcus ibis coromandus*. The worm was quite inactive and white in colour when liberated in normal salt solution. It is a long thread-like, thin worm and is somewhat fragile. The length



TEXT FIG. 5. *Gigantobilharzia egreta* male—Ventral view of (a) Anterior end showing intestine and cirrus sac and (b) Posterior end showing the termination of intestinal cæcum

TEXT FIG. 6. *G. egreta*, middle of the body showing the relation of testes with intestine

of the specimen, obtained by the writer, is 38.85 mm of which about 35.6 mm is occupied by the testes. It is broadest in the middle where it measures 275 mm. The body tapers gradually towards the anterior end, posteriorly also it tapers but widens again to end in a dilated and blunt posterior end. There is no trace of an oral or a ventral sucker. The gynaecophoric canal is also absent. The mouth begins in a shallow depression and is continued behind into a long oesophagus 1.2 mm in size. The intestinal caeca arise from the posterior end of the oesophagus and run for a very short distance to meet to form the common intestinal caecum. The caecal union takes place at a distance of about 4 mm behind the caecal bifurcation. The common caecum extends in a zigzag manner to almost up to the dilated posterior end of the animal.

The exceedingly large number of testes, more than 600, are round or oval bodies and lie on the two sides of the common caecum. They vary in size, the largest measures 1.5 mm  $\times$  1 mm and the smallest 0.75 mm  $\times$  0.5 mm. The vesicula seminalis lies in the space enclosed by the bifurcation and re-union of intestinal caeca. It is a sinuous structure, about 2 mm in size and ends in a small papilla on which is situated the genital aperture which lies at a distance of 175 mm from the caecal union and 225 mm from the caecal bifurcation. The writer has not been able to observe clearly a cirrus sac in this species which probably is present.

**Remarks**—So far only two species of the genus have been described, *G. acotylea* by Oehlner (1910) from Sweden and *G. monacotylea* by Szidat (1930) from East Prussia. The present form, of which the writer has obtained a male specimen, differs from both the existing species in several important characters. It differs from *G. acotylea* in the absence of a gynaecophoric canal, in having its genital opening slightly dextral and in possessing a comparatively much longer oesophagus. From *G. monacotylea*, it differs in the absence of an oral sucker, in the position of the genital pore, in much smaller vesicula seminalis and in having a much longer oesophagus. It is therefore, designated as a new species for which the name *Gigantobilharzia egretta* is proposed.

#### *Remarks on the Family Schistosomidae*

The family Schistosomidae contains 11 genera and these have been grouped into two subfamilies by Price (1929). The subfamily Schistosominae is distinguished from the other subfamily Bilharziellinae on the basis of a well-developed gynaecophoric canal and re-union of the intestinal caeca behind the middle of the body. The discovery of the new genus *Chinhula*, with an extremely well-developed gynaecophoric canal and

cephalad union of the intestinal caeca, is very interesting as it appears to form a connecting link between the two existing subfamilies. The difference between these subfamilies, therefore, narrows down to only one character, viz., the level of the re-union of the intestinal caeca. This naturally creates a doubt as to the validity of the subfamily division under the family.

Another interesting feature of systematic value is the occurrence of the cirrus sac which has been described in some species of the family. Unfortunately, the writer could not have an access to all the literature on the subject but feels that a re-study of some of the forms is needed. Species have been described under the same genus in which a cirrus sac is either present or absent. It is rather strange to find this discount on the value of the cirrus sac, which has been regarded as a subfamily character in various cases. The writer, therefore, urges for a revision of the genera and species of the family Schistosomidae in order to remove the prevailing confusion and thereby to place the group on a sound basis.

The writer wishes to record here his sincere thanks to Dr. G. S. Thapar for his valuable criticism and suggestions during the course of this work.

#### LIST OF REFERENCES.

- Bilharz, T. .. . "Fernere Beobachtungen über das die Pfortader des Menschen bewohnende *Distomum hematobium*," *Zeitschr. f. wiss. Zool.*, 1852, 4, 15.
- Christophers, S. R., and Stephens, J. W. W. .. . "Note on a peculiar *Schistosomum* egg," *Journ. Trop. Med.*, 1905, 8.
- Cobbold, T. S. .. . "On some new forms of entozoa," *Trans. Linn. Soc. Lond.*, 1859, 22, 4.
- Diesing, K. M. .. . "Revision der Myzohelminthen," *Sitzungab. d. k. Akad. d. Wiss. Wien*, 1858, 32, 21.
- Gogate, B. S. .. . "On trematodes from wild ducks in Rangoon," *Rec. Ind. Mus.*, 1934, 36.
- Johnston, S. J. .. . "On the trematodes of Australian birds," *Journ. and Proc. Roy. Soc. N. South Wales*, 1917, 50.
- Kowalewski, M. .. . "Bilharzia polonica sp. nov.," *Akad. Umies. w. Krakow*, 1895, 31.
- Leiper, R. T. .. . "Medical Helminthology. A review," *Trop. Diseases. Bull.*, 1923, 20.
- Looss, A. .. . "Westere Beiträge zur Kenntniss der Trematoden Fauna Aegyptens," *Zool. Jahrb. Abt. f. Syst.*, 1899, 12.
- Montgomery, R. E. .. . "Observations on bilharziosis among animals in India—I," *Journ. Trop. Vet. Sci. Calcutta*, 1906, 1.
- ..... .. . "Observations on bilharziosis among animals in India—II," *Journ. Trop. Vet. Sci. Calcutta*, 1906, 1.
- Moquin-Tandon, A. .. . *Elements de Zoologie Médicale*, Paris, 1860, 16.



A



B



C

FIG. A -- Section of Liver of *Nettion crecca* showing lesion formation due to the presence of *Chinhuta indica*

FIG. B -- Section of Kidney of *Nettion crecca* showing *Chinhuta indica* in the cavity of the kidney

FIG. C. --Section of Kidney of *Nettion crecca* showing *Chinhuta indica* in the renal vein, obliterating the cavity



Odhner, T.     "     "Gigantobilharzia acotylea, n. g. n. sp., ein mit den Bilharzien verwandter Blutparasit von enormer Lange," *Zool. Anz.*, 1910, 85

Price, E. W.     "     "Zum natürlichen System der digenen Trematoden," *Zool. Anz.*, 1912, 41

Price, E. W.     "     "A synopsis of the trematode family Schistosomidae," *Proc. U. S. Nat. Mus.*, 1929, 75.

Sewell, R. B. S.     "     "Note on *Macrobilharzia Travassosi*," *Jour. Parasit.*, 1931, 17

Skrjabin, K. I. and Zakharov, N. P.     "     "Cercarie Indicum," *Ind. Journ. Med. Res.*, 1922, 10

Soparkar, M. B.     "     "Zwei neue TrematodenGattungen aus den blutgefassen der Vogel," *Dansk Vet. Inst.*, 1920, 2

Szidat, L.     "     "The cercaria of *Schistosomum spinosalis*," *Ind. Journ. Med. Res.*, 1921, 9

Tanabe, B.     "     "Gigantobilharzia monocotylea, n. sp.," *Zeitschr. für Parasitenkunde*, 1930, 2

Travassos, L.     "     "The life-history of a new schistosome *Schistosomatium pathocopticum*, found in experimentally infected mice," *Journ. Parasit. Urbano*, 1923, 9

Weinland, D. F.     "     "Reprint—Informações sobre a fauna helminthológica de Mato Grosso," *Folk Med.*, Rio de Janeiro, 1923

Weinland, D. F.     "     "Human Cestodes," *Cambridge*, 1859, 10

# THE BRACKISH-WATER FAUNA OF MADRAS.

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Received October 1, 1937

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### *Introduction*

The study of the brackish-water fauna of India dates from the time of Stoliczka, and ever since he published his interesting paper in 1869, numerous isolated accounts of the fauna of the Gangetic Delta, dealing with particular groups or species of animals, have appeared from time to time. Faunistic and ecological studies pertaining to definite brackish-water regions were not, however, made until Annandale commenced, in 1907, his series of contributions on the fauna of the brackish ponds of Port Canning in Lower Bengal. This was followed by the biological survey of the Chilka Lake, under the leadership of Annandale and Kemp (1915). The various groups of animals collected from this lake were worked out by specialists in different parts of the world, these reports (1915-1922) have formed an excellent ground-work for further studies on Indian brackish-water fauna. The results of similar surveys of Taléh Sap in Siam, Tai Hu in China, and

Lake Biwa in Japan, were published in the succeeding years, in Annandale's *Zoological Results of a Tour in the Far East* (1918-1925). In recent years, considerable data have accumulated on the euryhaline fauna of the Gangetic delta mainly by the contributions of Annandale, Kemp, Sewell, and other officers of the Zoological Survey of India. An investigation of the brackish-water areas near the city of Madras was thought desirable as no attempt has been made to advance our knowledge of the brackish-water animals of South India beyond the references in the Chilka Lake reports. Apart from this aspect, the local animals which could be examined first-hand are so little known in many of the educational centres in India that both the student and the teacher are often handicapped owing to the paucity of familiar local species illustrating several biological phenomena, hence this work was planned with the hope that it may benefit College students in general. Comparatively little is known of the bionomics of the brackish-water types of animals, and since the place chosen was suited for close and constant investigation, an intensive ecological study was made extending over a period of three years. As will be seen from the following account, a fairly rich and specialized fauna exists at Adyar, showing interesting peculiarities both in mode of life and life-history.

#### *Material and Methods*

The account of the fauna given here is based upon a number of collections from the Adyar backwater, Adyar River, and the brackish-water localities of the Cooum, made within the course of three years, dating from November 1933. A few collections made previous to this have also been utilized, but these do not include any species that we have not taken subsequently. During the early part of the survey, i.e., for about a year, the Adyar backwater and the river were visited about thrice a month on an average, and regular shore and plankton collections were made. The backwater and the river were less frequently visited during the next year, but special attention was devoted to the upper reaches of the river and the pools of brackish-water near the Boat Club. Observations were also made on the fauna of the small islands in the river near the Elphinstone Bridge, and the brackish tracts of the Cooum. Field observations during the third year have confirmed the previous years' results.

The shallow nature of the backwater and the river was of considerable advantage in that no elaborate equipment was necessary for conducting this study. Shore collections were made by us regularly during our visits. For examining the fauna of the mud, the most useful instrument was a large shovel with which mud could be levered up without causing much

disturbance to the organisms. Mud taken in this manner from the backwater, salt pools, river, etc., was carefully examined in the laboratory and the organisms picked up. The tow-net was used to collect the free-swimming invertebrates, small fish and larvae. The collection also includes specimens obtained by dredging from the deeper parts of the river.

For the collection of fish, prawns and the larger species of swimming crabs, we have entirely depended upon the implements used by the fishermen of the locality. These were closely meshed nets of various sizes and shapes. Fishing is usually done in the mornings. Catches obtained by the fishermen from the river and the backwater were regularly purchased until representatives of most of the species were obtained. Afterwards, their catches were only examined on the field.

The hydrographical readings given were kindly taken by Dr M. K. Subramaniam, based on analyses of water-samples from the Adyar River near the bar. We have taken a number of temperature records on subsequent occasions, which have clearly illustrated the wide range of variation that is likely to occur from place to place, and also the extent of diurnal variation.

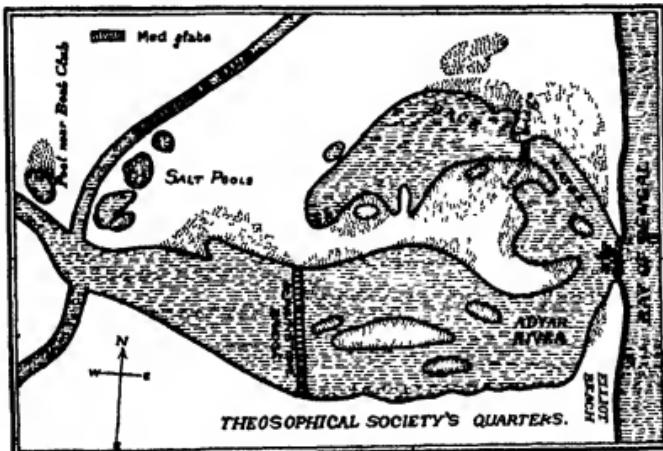
The species have been worked out by us, and in many cases the identifications have been checked with the help of the collections of the Madras Government Museum. Dr S. L. Hora has kindly identified for us four species of Gobies, Dr H. S. Rao, three Gastropods, and Dr F. H. Gravely, one spider. We wish to thank all of them for their valuable help.

#### *Description of the Area*

The brackish-waters of the city of Madras (Lat 13° 4', Long 80° 17' E) include the backwater at Adyar, and the mouth of the Adyar River which is in close association with the former. To this may be added those portions of the Cooum River and the Buckingham Canal, which are either in communication with the sea or with the Adyar backwater or river. The present study is mainly based on animals collected from the backwater and the river-mouth where the fauna is fairly rich and characteristic. A number of collections have also been made from the bar where the River Cooum opens into the sea. This account is also applicable to the other brackish-water regions of the Cooum and the Buckingham Canal as the macroscopic fauna of these localities consists only of a few of the resistant species that are observed at Adyar.

The Chingleput or the Adyar River opens into the sea five miles south of Fort St. George. The northern side of the river leads into the backwater which is a shallow sheet of water about one square mile in area, lying between

San Thome and Adyar. On the east, the backwater closely borders the sand bank that separates it from the seashore, while its northern and western portions extend into shallow mud-flats, which are covered by water only during high tide, and are more or less permanently dry during the hot months of the year, March to July. There is no sharp separation between the backwater and the river-mouth proper as the former is something like a bay on the northern bank of the river, from the faunistic



TEXT-FIG. 1

point of view, the portion of the river from the Elphinstone Bridge up to the bar, nearly a mile in length and about quarter to half a mile in width, along with the backwater forms a typical brackish-water zone. As the river is very shallow lower down the bridge, a few island formations occur in the middle of its course.

The peculiarity of the place is brought about by the fact that the Adyar River is not in communication with the sea throughout the year. As the source of the water-supply for the river is the comparatively low rainfall of the Chingleput District (about 49 inches per annum), it is full only during the North-East Monsoon period (October to December) when there is good flow of fresh water into the sea, and the river-mouth is fairly deep and broad during the flood season. This condition exists only for a

period of about two months. As the strength of the current gradually weakens, tidal effects are markedly felt and the rapid flow of water is very much inhibited. The presence of the backwater is of much significance in that it allows a good admixture of sea water and freshwater. These conditions change with the advent of summer. The river becomes gradually reduced to a languid line of water with practically no flow. By about the end of January, the fringes of the backwater get dried up, leaving some of the mud-flats exposed. The immediate effect is an increased flow of water from the sea into the river and the backwater, but this is soon retarded by the action of breakers, which raise up a sand bank at the mouth of the river on a line with the shore. Thus the connection of the river and the backwater with the sea is intermittent during February, but it is completely cut off by the end of March. The sand bank, or the *bar* as it is called, widens out in latter months, reaching a maximum width of about three-quarters of a furlong. The bar is prominent until the next rainy season when the river-current is again strong enough to force its way into the sea. The times of opening and closing of the bar may be considered as approximately occurring between October and November, and between February and April respectively.

The *Cooum River* is only a tidal creek (similar to the Matla River of Lower Bengal) which has a tortuous course in the city of Madras. It is practically dry during the summer months. At the monsoon time, a good amount of rain water collects in the river. A temporary connection is now established with the sea, about half a mile south of Fort St. George, where the river crosses the Buckingham Canal. Soon after the monsoon, however, a bar is raised, but this is quite low and allows the inflow of small quantities of sea-water during high tide. The portion of the Buckingham Canal with which we are concerned in this paper is that between the Adyar and the Cooum, from the point where it cuts the former near the Adyar Boat Club four miles up the river, to the point where it crosses the Cooum near the Napier Bridge to the south of Fort St. George.

Geologically, the whole locality is built by the sea straggling over a large area of the sandy soil characteristic of the greater part of the Coromandel Coast. The soil of the adjoining areas consists of a mixture of sand and red loam. The nature of the soil is such that in the wet weather, many temporary pools of water are formed, this is enhanced by the presence of tidal creeks like the Cooum. In the formation of the backwater, the sand-laden current of the coast coming from the south and grazing the shore in a northerly direction towards the head of the Bay of Bengal, must have played a prominent part since it has caused the sea to recede, and has directed the water and silt of the sluggish river in the direction of the

backwater. A number of marine and estuarine shells in a sub-fossil or fossil state have been recorded from the Adyar locality by Oldham (1893).

The bottom of the river as well as of the backwater is of a muddy consistency, but the sandy element in the soil becomes pronounced as we approach the sea. Excepting on the eastern border, the whole of the backwater-edge is muddy, composed of thick brown mud mixed with black clay, the latter predominating in its northern sides where the bed is almost completely composed of fine, loose clay. In the river-bed, the clay is less marked, but the soil is composed of sand and mud mixed in more or less equal proportions. The boulders of the Elphinstone Bridge and the small bridge near the Adyar Cemetery, and the stony embankment near the Theosophical Society's Quarters on the southern bank of the river are the only hard substrata in this brackish-water tract, giving shelter to typical rock-dwelling species.

*The Depth*—The chief difference between the river and the backwater lies in their respective depths. Nowhere in the backwater is the maximum depth more than 4 feet during low tide, except during floods. Two distinct zones may be observed in the backwater: a *middle zone* that never dries up even in the hottest months and extends from the bar up to the small bridge in a curve, and a *marginal zone* on the fringes of the former, consisting of mud-flats, which are completely submerged only during monsoon times and are normally left exposed during low tide. The relative depths of the two zones are such that when the marginal zone is just exposed, the water in the middle zone will be two to four feet deep. In contrast with the backwater, the river is considerably deeper, even during the dry weather, a channel, nearly six feet deep, is noticeable near its southern bank up to about a mile from the sea. This extends right up the river and is navigable for small country crafts. A similar deep channel is seen on the northern bank almost up to the bridge but beyond this up to the Boat Club the river merges into a series of mud-flats and small pools of brackish-water. These pools are brackish although many of them have no direct communication with the river. Their brackish nature is the result of percolation of water through the raised banks. One such pool near the Boat Club is fairly big and will be specially referred to later. Higher up the river, the pools contain only fresh water and are inhabited mainly by freshwater organisms.

*The Tides*—The tides are felt only in the months during which the bar is open, their influence is felt up to a distance of about three to four miles up the river. The maximum tidal effect is a little after the monsoon, from about January to the end of February, when the river-current is feeble.

The maximum tidal range is about three feet. As has been mentioned above, the initial drying up of the backwater during the early summer months is followed by an increased flow of water from the sea into the river and backwater until a bar is formed.

*The Vegetation*—The vegetation is rather poor in the river as may be expected from the depth and the flow of water, but the backwater is in sharp contrast with this, its shallow stagnant situation being particularly suited to the luxuriant growth of algae. Numerous forms of algae inhabit the middle zone of the backwater, chief among them are *Euleromorpha* and *Chatomorpha*. Algal beds are best developed during the summer months of the year when they occur almost everywhere and give shelter to numerous organisms. The bottom of the middle zone of the backwater and the shallow portions of the river is closely covered with *Polamgetons* while marsh plants like *Avicennia* and *Sueda* are quite common in the mud-flats and the islands in the river. As we proceed higher up, freshwater forms like *Spirogyra*, *Ceratophyllum* and others may be found in large numbers.

#### *The Environment*

The salinity, temperature, pH, chlorine and excess base values are given in the accompanying table, from a set of readings taken in 1933. These readings refer to the conditions near the bar. As may be expected from the nature of the locality, the salinity of the water in the backwater and the river is liable to great variation, being not only different in different times of the year, but also varying in different places at the same time. The salinity is highest from January to March when the rains have practically ceased but the bar remains open—a period during which the maximum

TABLE I

*Hydrographical Readings Averages for 1933  
Adyar River Water, near Bar*

	Bar open						Bar closed				
	Nov	Dec.	Jan.	Feb	Mar	Apr	May	June	July	Aug.	Sept.
Temperature °C	25.4	25.0	26.2	27.7	28.2	29.0	28.8	28.6	28.5	28.2	28.2
pH	8.50	8.48	8.55	8.46	8.46	8.03	9.07	9.15	8.82	8.70	
Excess Base	30.15	20.73	23.20	17.90	25.16	24.90	24.52	26.5	23.50	23.30	
Salinity	28.10	28.17	30.44	29.92	27.20	19.74	16.91	17.10	18.20	19.88	
Chlorine	15.84	15.59	16.85	16.66	9.66	10.92	9.85	9.51	10.12	10.72	

flow of sea-water into the backwater takes place. At the bar the salinity of the water was 28.17, 30.44, and 29.92 per mille in the months of January, February and March respectively. The corresponding figures for April, May and June were 17.29, 19.74, and 16.91 respectively. The greatest variations in salinity occur when the bar is closed since the occasional showers during June-July bring down considerable quantities of freshwater, effecting a notable reduction in salinity. On the other hand, intense heat of the summer results in an increase in salinity of the backwater, especially of the pools in the marginal zone, since the volume of water is very much lowered owing to drought.

In the absence of any large river opening into the sea in the vicinity of the city of Madras, the salinity of the coastal waters here is much higher than the records for the upper limits of the Bay of Bengal. According to Sewell's charts, the maxima and minima are 34.50 (June-August) and 32.00 (September-November) per mille respectively (Sewell, 1929). This is in sharp contrast to what occurs at the head of the Bay, where the surface salinity may be so low as 21.00 per mille, owing to the large volume of freshwater brought down by the Ganges and the Brahmaputra. The change from marine to brackish-water conditions is gradual in the Gangetic Delta, while it is rather abrupt at Adyar.

*Temperature*—The maximum and minimum records of temperature of the water are 33.80 and 25.00 degrees Centigrade respectively. The temperature is the lowest during the rainy season when the bar is open, and is highest during the summer months May and June. From January to March, the temperature of the water is slightly higher than that of the air, while in the succeeding months of April, May and June it is below the air temperature. As in the case of salinity, the temperature is also subject to a great amount of fluctuation. In the backwater, there is often difference between the readings for the middle and marginal zones, and the surrounding pools. The deeper central zone has a more uniform temperature than the other regions. The exposed mud-flats with numerous pools show the greatest amount of variation as they are quickly heated up by sunlight. The following readings of surface temperature taken at 12 Noon on a bright sunny day in the month of October (1-10-1934) are illustrative—

Temperature of the sea-water	27° 70 C
Temperature of the water at bar	28°.00 C
Middle zone	29° 20 C
Marginal zone	32°.00 C
Temperature of an isolated pool.	. 33°.80 C

The maximum difference noted between morning (8 A.M.) and afternoon (2-3 P.M.) temperatures is 3.5° during the month of June

The pH values normally vary between 8 and 9. As will be seen from the table, the highest values are observed during the months June and July.

*List of Animals Collected from the Brackish-waters of Madras.*

The following list does not include the Protozoa and the Rotifera since we have not made any special study of them. Only the commonest species of the Copepods obtained are listed. The other groups are fairly exhaustive. Those species which have been recorded by other authors, but which we have not been able to obtain, are mentioned separately. The latest nomenclature has been followed as far as possible.

*Phylum Porifera*

*Spongilla* sp.

*Phylum Cælenterata*

Class *Hydrozoa*

*Dicyclozome filamentata* (Annandale) Medusæ only

*Campanulina ceylonensis* (Browne) Medusæ only  
(*Phoritis ceylonensis* of certain authors)

*Campanularia (Clytia) noliformis* McCrady Hydroid only.

*Laomedea (Obelia) spinulosa* Bale Hydroid only

Class *Scyphozoa*

*Acromitus flagellatus* (Haeckel) The species is probably the same as *A. rabanchatu* Annandale (*vide* Rao, H. S., 1931)

Class *Anthozoa*

Order *Actiniaria*

Sub-tribe *Athenaria*

*Phyllocates gangeticus* Annandale

*Phyllocarpopsis ramunni* Panikkar

*Stephensonactis ornata* Panikkar

*Pelocetes exul* Annandale.

Sub-tribe *Boloceroidaria*

*Boloceractis gopala* Panikkar

*Bunodeopsis* sp. (not *B. strumosa*)

Sub-tribe *Acontaria*

*Asplasiomorpha* sp.

*Phylum Platyhelminthes*

Order *Acala*

*Convoluta* sp.?

*Phylum Nemertea**Order Heteronemertines*

A species belonging to this order is common in the pools of brackish-water all through the year

*Phylum Nemathelminthes**Class Nematoda**Family Oncholaimidae*

*Oncholaimus* sp Common free living Nematode. The species is different from *O. indicus* and *O. chikensis*

*Phylum Annelida**Order Polychaeta**Family Hesionidae*

*Ancistrosyllis constricta* Southern

*Family Nereidae*

*Lycastis indica* Southern

*Nereis glandicincta* Southern

*Nereis chikensis* Southern

*Family Eunicidae*

*Diopatra variabilis* Southern (Fauvel, 1932, considers the species as being synonymous with *D. neapolitana*)

*Marphysa gravelyi* Southern

*Lumbriconereis polydesma* Southern

*Lumbriconereis* sp

*Family Nephthydidae*

*Nephthys polybranchia* Southern

*Family Aricidae*

*Scoloplos* sp

*Scolecolepis* sp Probably *S. indica* Fauvel

*Family Spionidae*

*Polydora ciliata* Johnston

*Polydora kempfi* Southern

-

*Prionospio cirrifera* Wieren

*Family Capitellidae.*

*Heteromastus similis* Southern

*Capitella* sp

*Family Sabellidae*

*Laonome indica* Southern

*Potamilla leptocheta* Southern

*Family Serpulidae*

*Hydroides norvegica* Gunnerus.

Order *Oligochaeta*Family *Megascolecidae**Pontodrilus bermudensis* BeddardPhylum *Polypzoa*Order *Ctenostomata*Family *Vesicularidae**Bowerbankia caudata* HincksPhylum *Arthropoda*Class *Crustacea*Order *Copepoda*

Only the commonest species are noted

*Acartia southwelli* Sewell*Pseudodiaptomus annandalei* SewellOrder *Cirripedia*Division *Operculata* Family *Balanidae**Balanus amphitrite* DarwinOrder *Schizopoda*Mysidae Family *Mysidae**Rhopalophthalmus eggregius* Hansen*Mesopodopsis orientalis* (Tatcral)Order *Amphipoda*Amphipoda *Genus* *Gammarina**Grandidierella megnea* (Giles)*Grandidierella gilesi* Chilton*Paracalliope fluvialis* (G. M. Thomson)Order *Tanaidacea**Apseudes* sp. not *A. chilensis*Order *Isopoda*Isopoda *Genus*Family *Cirolanidae**Cirolana* sp. probably *C. pleonastica* StebbingFamily *Sphaeromidae**Sphaeroma vastator* Spence Bate (S. *terebrans* of certain authors)Family *Cymothoidae*Cymothoa *indica* Schiodte and Meinert. The species is parasitic in the mouth and gill chambers of *Etroplus maculatus*, *E. suratensis* and *Glossogobius giuris* (Panikkar and Aiyar, 1937)Family *Ligidae*.*Ligia exotica* Roux.Order *Decapoda*

*Decapoda Natantia*Tribe *Caridea*.Family *Palæmonidae**Periclimenes indica* Kemp*Periclimenes demans* Kemp*Leander* sp.*Palæmonetes hornelli* Kemp*Palæmon lamarres* Milne-Edwards.Family *Alpheidae**Alpheus malabaricus* Fabricius*Alpheus paludicola* Kemp*Alpheus* sp. *A rapax*?Tribe *Penaeidae*Family *Penaeidae**Penaeus carinatus* Dana*Penaeus indicus* Milne-Edwards*Penaeopsis monoceros* (Fabricius)*Decapoda Reptantia*Tribe *Oxystomatida*Family *Calappidæ**Matuta vitor* (Fabricius)Tribe *Brachygnatha*Family *Ocypodidae**Ocypoda cordimana* Desmarest*Ocypoda macrocerata* Milne-Edwards*Uca (Gelasimus) annulipes* (Latrelle)*Uca (Gelasimus) triangularis* (A Milne-Edwards) The species is recorded by Henderson. We have not obtained it so far.*Metaplaax distincta* (Milne-Edwards)Family *Grapsidae**Varuna litterata* (Fabricius)*Sesarma tetragonum* (Fabricius)*Sesarma quadratum* (Fabricius)*Grapsus maculatus* Catesby (Syn *G. grapsus*)*Melasesarma rousseauxii* H Milne-EdwardsFamily *Portunidae**Scylla serrata* (Forskal)*Neptunus pelagicus* (Linnæus)*Neptunus sanguinolentus* (Herbst).

Tribe *Paguridea*Family *Paguridae*

*Chibanaeus olivaceous* Henderson  
*Chibanaeus padavensis* De Man  
*Diogenes* sp *D. avarus*?

Phylum *Mollusca*Class *Lamellibranchiata*Family *Ostreidae*

*Ostrea arakanensis* Sowerby The species is synonymous with *O. madrasensis* and *O. virginica* of Preston and other authors (vide Waukworth, 1931)

Family *Mytilidae*

*Modiolus striatula* Hanley  
*Modiolus undulatus* (Dunker)

Family *Arcidae*

*Area granosa* (Linnaeus)

Family *Veneridae*

*Meretrix casta* Chemnitz

Family *Cuspidariidae*

*Cuspidaria cochinensis* Preston

Class *Gastropoda*Family *Hydrobiidae*

*Stenothyra blanfordiana* Nevill  
*Amnicola (Alocinma) stenothyroides* Dohrn

Family *Cerithiidae*

*Potamides cingulatus* (Gmelin)

*Melania tuberculatus* (Muller)

Family *Nasidae*

*Pygmæonassa orissensis* (Preston)

Family *Aplysiidae*

*Aplysia* sp

Family *Hermidae*

*Stiliger gopala* Rao

Phylum *Chordata*Class *Pisces*Order *Teleostei*Sub-Order *Malacopterygii*Family *Elopidae*

*Elops indicus* Swainson

*Megalops cyprinoides* (Broussonet)

Family *Channidae**Chanos chanos* (Forskal)Family *Clupeidae**Engraulis purara* (Hamilton Buchanan)*Stolephorus commersonii* Lacepede*Clupeoides Isle* (Cuv et Val)Sub-Order *Ostariophysi*Family *Siluridae**Plotosus canus* Ham Buch*Macrones vitulus* (Bloch)*Macrones gutto* (Ham Buch)*Macrones keletius* Cuv et ValFamily *Cyprinidae**Barbus sophore* (Ham Buch)*Barbus dorsalis* (Jordan)*Barbus amphibius* (Cuv et Val) The species is not represented in our collections, but is mentioned by Raj (1916)Sub-Order *Haplochthys*Family *Cyprinodontidae**Panchax parvus* Raj*Aplocheilus melastigma* McClellandSub-Order *Percesoces*Family *Scombriscidae**Hemiramphus limbatus* Cuv et ValFamily *Mugilidae**Mugil cephalus* Linn*Mugil* spFamily *Sphyracidae**Sphyrana jello* Cuv et ValSub-Order *Acanthopterygii*Family *Serranidae**Serranus sexfasciatus* Cuv et Val*Chanda (Ambassis) ambassis* (Lacepede)*Chanda (Ambassis) myops* Gunther*Lutjanus johnii* (Bloch)*Therapon jarbua* (Forskal)*Therapon puta* Cuvier*Therapon quadrilineatus* (Bloch)Family *Sillaginidae**Sillago sihama* (Forskal)

Family *Squamipinnes**Scatophagus argus* (Bloch)Family *Mullidae**Upeneus sulphureus* Cuv et ValFamily *Gobiidae*Sub-family *Gobinae**Gobius polynema* (Bleeker)*Gobius melanosticta* Day (Not obtained by us, but mentioned by Day.)*Gobius criniger* Cuv et Val*Glossogobius giuris* (Hamilton Buchanan)*Ctenogobius (Oligolepis ?) acutipinnis* (Cuv et Val)*Ctenogobius meggitii* Hora and Mukerji.*Acentrogobius neills* (Day)*Acentrogobius viridis punctatus* (Day)*Oxyurichthys tentacularis* (Cuv et Val)*Apocryptichthys* sp Not *A. cantoris* Day*Boleophthalmus boddaerti* (Pallas)Sub-family *Eleotrinae**Eleotris fusca* (Bl and Schn)Sub-family *Periophthalminae**Periophthalmus kelsebrentri* (Pallas)*Periophthalmus pearsei* EggertSub-Order *Zeorhomboi*Family *Bothidae**Pseudorhombus arnisi* (Hamilton Buchanan)*Pseudorhombus javanicus* (Bleeker)*Scombriformes*Family *Carangidae**Caranx cirratus* (Bloch)*Jugulares*Family *Blennidae**Petroscirtes bhattacharyae* Chaudhuri*Scleroparei*Family *Platycephalidae**Platycephalus scaber* (Linn)*Platycephalus insidiator* (Forskal)*Chromides*Family *Cichlidae**Eretroplus maculatus* (Bloch)*Eretroplus suratensis* (Bloch)Sub-Order *Plecognathi*

*Sclerodermi*Family *Triacanthidae**Triacanthus brevirostris* Temm and Schleg.*Gymnodontes*Family *Tetraodontidae*,*Tetraodon patoca* Hamilton Buchanan*Tetraodon inermis* Temm and SchlegClass *Reptilia*Order *Ophidia**Natrix piscator* (Schneider)*Cerberus rynchops* (Schneider)*Regional Distribution and Range of Fauna*

Apart from the regional survey of the fauna of Adyar that follows, the range of distribution of many of the species may be indicated here, especially of those that show great restriction or variation in occurrence. Species that have a wide range of distribution from the bar (even seashore in some instances) up to the inner reaches of the river and the backwater where the water is almost fresh during low tide are —

*Acromitus flagellatus**Lycastis indica**Heteromastus similis**Capitella* sp*Modiolus striatula**Scylla serrata**Mesopodopsis orientalis**Tetraodon patoca*

These species have been obtained from collections made from the river near the Teachers' College and the Engineering College. *Acromitus flagellatus* has been observed only during high tide, probably it comes in with the rising tide and retreats with the tidal flow, without having the necessity to remain in freshwater. Others have been collected irrespective of tidal influence, *Scylla serrata*, *Mesopodopsis orientalis*, *Lycastis indica* and the Capitellids can remain in water that is nearly fresh. Large numbers of young ones of Tetrodons and colonies of Modiolas were collected from about two furlongs east of the Sandapet Bridge, the latter in clusters of hundreds from the stony banks of the stream. This was in the month of January, 1935, an abnormal year as the rainfall was rather scanty, and the bar was about to close even though the rainy season was only just over. Only Modiolas, Mesopodopsis and the Capitellids appear to be able to breed

here in the uppermost reaches of the brackish-water zone among the animals listed previously

Coming down the river for about a mile and a half from the above locality where the water is almost fresh, the marine element in the fauna becomes more marked, and near the Adyar Boat Club the conditions probably range from oligohaline to mesohaline environments. The fauna is not rich in the river proper, but the adjoining pools that occur on either side harbour a rich fauna. The organisms found in these pools will be considered separately. The 'marine' species that are common in this zone of the river are —

- Phytocetes gangeticus*
- Lycastis indica*
- Lumbriconeress polydesma*
- Lumbriconeress sp*
- Marphysa grisea*
- Penaeopsis monoceros*
- Grandidierella magna*

The typical brackish-water tract that harbours a rich fauna commences from about half a mile lower down, and consists of that part of the river near the Elphinstone Bridge and the backwater. The animals that are found quite close to the bar include only true marine species and are obviously forms that cannot survive great decrease in the salinity of the environment.

One usually finds the following animals at or near the bar —

- Glycera sp*
- Ocypoda cordimana*
- Ocypoda macrocera*
- Matula vincta*
- Neptunus sanguinolentus*
- Neptunus pelagicus*
- Aplysia sp*
- Plaicephalus insidiator*

*Chthamalus padavensis* is also found but its occurrence in this region is not due to its lack of adaptational power to less saline environments, but only to a preference to a sandy bottom. The invertebrate animals that are common in the seashore just opposite the mouth of the river and the vicinity are —

- Cavernularia malabarica*
- Sphenopus marsupialis*

*Onuphis eremita*  
*Glycera* sp  
*Donax* sp  
*Leiodesmus vittatus*  
*Littorina* sp  
*Oliva gibbosa*  
*Albunea seminista*  
*Emerita (Hippa) astatica*  
*Philyra scabriscula*  
*Ocypoda platytarsis*  
*Ocypoda macrocera*

Excepting *Ocypoda macrocera*, none of these species has invaded the brackish-water

The distribution of many of the species is considerably influenced by the nature of the substratum. Species like *Potamides cingulatus*, *Phycocetes gangelius*, *Phyloceratopsis ramunni*, *Stephensonactis ornata*, *Seylla serrata*, *Uca annulipes*, *Sesarma quadratum*, *Sesarma tetragonum* and a few others are not found in completely sandy areas. They show a definite preference to localities where the bottom consists of sand and mud mixed more or less in equal proportions. However, *Sesarma tetragonum*, *S quadratum* and *Uca annulipes* are more frequently found in wet mud-banks that are not submerged as all of them are of almost terrestrial habits. The majority of Polychaetes are also found in shallow localities of a mixture of sand and mud. *Neptunus pelagicus*, *N sanguinolentus*, *Matula vincta*, *Clibanarius padavensis*, *Ocypoda cordimana*, *O macrocera* and *Platyccephalus insidiator* are species almost invariably restricted to sandy regions. The south-western fringe of the backwater is characterised by soft black clay that emits hydrogen sulphide and the fauna here is rather sparse. There are practically no animals where the clay is thick, but in other regions where the sub-stratum is loose *Lumbriconereis* sp., *Marphysa gravelyi* and *Prionospio cirrifera* are often common among the Polychaetes, and *Stenothyra blanfordiana* and *Pygmæonassa orissensis* among the Molluscs. The Buckingham Canal, the Cooum, and the brackish-water ditches adjoining them have a poor macroscopic fauna as the water in them is much polluted. The animals that are collected usually from these localities are *Sesarma quadratum* and *Lycastis indica* near the water edge, *Marphysa gravelyi*, *Melanoides tuberculatus* and *Lumbriconereis* sp. in the mud. Small ditches of water near the Cooum bar harbour large numbers of *Potamides cingulatus*. *Metasesarma rousseauini* is often found crawling amidst the stones near the Napier Bridge.

*Bionomical Classification of the Fauna*

The backwater and the river may be divided according to the environmental conditions and the habits of the different species into the following groups

*I Fauna of the Algal Beds* —The backwater and the adjoining pools harbour a rich algal flora consisting of species of *Enteromorpha*, *Chætomorpha*, *Gracilaria* and other forms, while the bottom is often covered with dense growths of *Potamogeton* sp. The number of animals inhabiting the algae and the leaves of *Potamogeton* is very great and they are here collectively designated as the algal-bed fauna. As the maximum algal growth is during the summer when the bar is closed and the water still, the animals inhabiting the algae have corresponding maxima periods in summer. The following forms have usually been obtained from the algal beds: —

- Boloceractus gopalai*
- Bunedeopsis* sp
- Planarian* (*Convoluta* sp ?)
- Oncholaimus* sp
- Bowerbankia caudata*
- Nereis glandicincta*
- Modiolus undulatus*
- Stenothyra blanfordiana*
- Amnicola (Alocinma) stenothyroides*
- Cuspidaria cochinchensis*
- Stiliger gopalai*
- Grandisserella magna*
- Grandisserella gilesi*
- Paracalliope fluviatilis*

Of these the species of Amphipods are obtained in large numbers all through the year. The Molluscs are usually common only during the summer and a few succeeding months. The *Planarian* has been noted only in the month of August, and that for a short period. None of the other species shows any particular dominance during other periods. The algae amongst which the animals live form the food for many of them.

*II Fauna of Rocky Substrata* —The animals grouped under this head include (1) attached forms like *Balanus* and *Laomedea* which are found on the boulders of bridges, stones on the banks, etc., (2) species that occur in water where the bottom is stony, and (3) forms that live on stones on the water-edge and are aquatic, amphibious or terrestrial. The following are the common species: —

*Spongilla* sp  
*Campanularia noliformis*  
*Laomedea spinulosa*  
*Aiptasiomorpha* sp  
*Nereis chilensis*  
*Ancistrosyllis constricta*  
*Hydroids norvegica*.  
*Medsolus striatula*  
*Ostrea arakanensis*  
*Balanus amphitrite*  
*Sesarma quadratum*  
*Clibanarius olivaceous*  
*Grapsus maculatus*  
*Liguexotica*  
*Cirolana* sp

Of these the Sabellid and Hydroids are common only when the bar is open. Goboid fishes like *Acentrogobius viridipunctatus*, *Ctenogobius meggitti* and *Glossogobius giuris*, and the Cichlid fish *Etroplus maculatus* and *E. suratensis* are often collected from localities with a stony bottom. Their eggs are found attached to the stones, empty shells, etc. The snake *Natrix pectoralis* is fairly common on the southern banks of the river.

*III The Free-Swimming Fauna*—The following animals constitute the free-swimming element of the fauna —

*Dicycloeryne filamentata* (Meduse)  
*Campanulina ceylonensis* ( .. )  
*Acromitus flagellatus*  
*Mesopodopsis orientalis*  
*Rhopalophthalmus egregius*  
*Periclimenes indica*  
*Periclimenes demans*  
*Leander* sps  
*Palamon Lamarres*  
*Penaeus carinatus*  
*Penaeus indicus*  
*Penaeopsis monoceros*  
*Copepods*  
*Fish*

*IV Fauna of the Marginal Zone*—The animals that inhabit the marginal zone of the backwater include numerous species, the majority of them are inter-tidal forms found in the mud-flats or the water edge. Most

of the species are either amphibious or terrestrial, or the species are capable of surviving temporary exposure to air. They are highly resistant and are endowed with great powers of adaptability. The following species may be noted —

*Phycocetes gangeticus*  
*Clibanarius olivaceus*  
*Clibanarius padavensis*  
*Sesarma quadratum*  
*Sesarma tetragonum*  
*Melasesarma rousseauxii*  
*Metaplexa distincta*  
*Varuna littoralis*  
*Uca annulipes*  
*Stenothyra blanfordiana*  
*Amnicola stenothyrodes*  
*Potamides cingulatus*  
*Periophthalmus pectoralis*  
*Periophthalmus kælreutri*  
*Boleophthalmus boddarti*

The species of *Clibanarius*, *Sesarma quadratum*, *Uca annulipes*, *Potamides cingulatus*, the Hydrobiid Molluscs, and *Boleophthalmus boddarti* are the commonest species found near the edges of the Adyar backwaters, especially when the mud-flats are exposed. The marginal species found near the mouth of the Cooum are *Potamides cingulatus*, *Melania tuberculatus*, *Melasesarma rousseauxii*, *Ocypoda cordimana*, *Sesarma quadratum* and *Uca annulipes*.

*V Fauna of the Middle Zone* — Passing on to the middle zone of the backwater, the following species are commonly met with —

*Pelocates exul*  
*Marphysa gravelyi*  
*Diopatra variabilis*  
*Lumbriconereis* sp  
*Heteromastus* sp  
*Meretrix casta*  
*Pygmæonassa orissensis*  
*Matuta vincta*  
*Scylla serrata*

In addition, most of the free swimming species of Crustaceans, and fishes are also obtained from the middle zone. The anemones *Phytoceteopsis ramunculus*, *Stephensonactis ornata* and *Phycocetes gangeticus* are found between

the marginal and middle zones of the backwater. The last mentioned species has also been obtained from exposed mud-flats with very little water.

*VI The Mud-Burrowing Fauna*—Most of the species listed above under the fauna of the marginal and middle zones are either mud-burrowing in habits or are capable of temporarily remaining in the mud. The following species are typical burrowers.—

(a) Under water—

- Phytocetes gangeticus*
- Phytoceteopsis ramunni*
- Stephensonactis ornata*
- Pelocates exul*
- Marphysa gravelyi*
- Diopatra variabilis* (also tube-dwelling)
- Lumbriconerets* sp
- Prionospio* sp
- Capitella* sp
- Heteromastus* sp

(b) Outside water—

- Sesarma quadratum*
- Sesarma tetragonum*
- Melasesarma rousseauxii*
- Varuna littoralis*
- Uca annulipes*
- Ocypoda cordimana*
- Ocypoda macrocera*
- Pontodrilus bermudensis*

Species that can temporarily remain under the mud in the water for varying periods are—

- Apsenades* sp
- Meretrix casta*
- Scylla serrata*
- Alpheus malabaricus*
- Alpheus paludicola*
- Penaeus indicus* (Young ones)
- Penaeus carinatus* (Young ones)
- Penaeopsis monoceros*
- Leander* sp
- Periophthalmus pearsi*
- Boleophthalmus boddaerti*
- Platycephalus scaber*

The following species creep about near the water-edge :—

- Clibanarius olivaceous*
- Clibanarius padavensis*
- Potamides cingulatus*
- Stenothyra blanfordiana*
- Amnicola stenothyronides*

*VII Species Capable of Aerial Respiration*—Many of the animals inhabiting the mud-flats are either amphibious or terrestrial in habits. Though they belong to groups of animals that are primarily aquatic, they are capable of varying degrees of aerial respiration. The crabs and the Gobiod fishes are the most remarkable among them as they show a series of adaptations designed for aerial respiration, and some of the Crustaceans are thoroughly terrestrial and can respire only outside water. The following species mostly remain outside water.—

*Crustacea*—

- Grandidierella magna*
- Grandidierella gilesi*
- Paraceliope fluviatilis*
- Ligia exotica*
- Uca annulipes*
- Ocypoda cordimana*
- Ocypoda macrocera*
- Metaplaax distincta*
- Melasesarma rousseauxii*
- Sesarma quadratum*
- Sesarma tetragonum*
- Grapsus maculatus*
- Varuna littoralis*
- Clibanarius olivaceous*
- Clibanarius padavensis*

Among these species, *Ocypoda cordimana*, *O. macrocera*, *Grapsus maculatus*, and *Ligia exotica* are almost completely terrestrial.

*Fishes*—

- Periophthalmodon pearsei*
- Periophthalmodon kaelreuteri*
- Boleophthalmus boddarti*

*Other Invertebrates*—

- Lycasias indica*
- Capitellids*

*Potamides cingulatus*  
*Stenothyra blanfordiana.*  
*Amniscola stenothyroides*

In addition, the following species can survive exposure to air for different periods varying according to the species —

*Phycocetes gangeticus*  
*Scylla serrata*  
*Neptunus sanguinolentus*  
*Palamon lamarres*  
*Penaeopsis monoceros*  
*Oxyurichthys tentacularis*  
*Ctenogobius (Oligolepis ?) acutispinus*  
*Platycephalus scaber*

**VIII Fauna of Isolated Brackish-water Pools** — The pools of brackish-water in association with the backwater and the river are of three kinds, they are —(1) Small pools on the fringes of the backwater, harbouring a rich algal flora during the summer, and at other times having the same environmental conditions as the backwater, (2) Salt pools that occur on either side of the river up to an extent of about three miles from the river mouth, and (3) Pools in which the water is only very slightly brackish or completely fresh, occurring higher up the river and containing freshwater organisms in addition to a few brackish-water species that enjoy a wide range of distribution (compare above)

The first type of pools contains the following species, excluding the alga-inhabiting forms which have been given separately

*Phycocetes gangeticus*  
*Phycoceteopsis ramunni*  
*Campanulina ceylonensis*  
*Clibanarius olivaceous*  
*Paracalliope fluvialis*  
*Grandisiderella megae*  
*Potamides cingulatus*

The temperature fluctuations are rather high in these pools as has already been pointed out.

The pools belonging to the second category are numerous, but a large and typical one is that found near the Adyar Boat Club, the fauna of which during the different seasons was studied by us in detail. This pool is situated in somewhat marshy soil just adjoining the Adyar River on its west and the Buckingham Canal to the south, and it is about half a furlong

long. The pool gets filled up by the November rains, but the level of the water gradually falls in the succeeding months. The depth of water is about 2-2½ ft. during November-December period (this is about the maximum depth), but by February, the depth is only a few inches. The pools are almost without water by about March. Owing to percolation of water from the river, the water in the pool remains brackish all through the year and the fauna is what is characteristic of the brackish-water. From November to March, the following species are obtained from these pools —

- Acromitus flagellatus*
- Phyllocetes gangeticus*
- Lumbriconereis polydesma*
- Lumbriconereis* sp
- Lycaster indica*
- Murphysa gravelyi*
- Heteromastus similis*
- Prionospio curvifera*?
- Mesopodopsis orientalis*
- Periclimenes indica*
- Palaeomon lamarrei* (Stray individuals only)
- Paracalliope fluviatilis*
- Sesarma quadratum*
- Sesarma tetragonum*
- Aplocheilus melastigma*
- Acentrogobius neilli*
- Acentrogobius viridis punctatus*

By April, the conditions of the pool change very much since most of the aquatic species perish with the drought. The interesting point about this pool is that unlike the pools in the mud-flats that completely dry up during the summer, the bottom remains slightly moist even in the hottest months of the year as a result of percolation of water from the river. Contrary to what happens in the mud-flats, the muddy substratum does not become cracked up in the usual manner. During this period, the pool has a certain amount of resemblance to a salt marsh, and as is usual with the salt marsh fauna, typical terrestrial species are met with in large numbers. Numerous insects, among which the one frequently encountered is a Formicid, and a spider belonging to the genus *Olios* (Family Sparassidae) and probably to the widely distributed Oriental species *Olios lamarchi* (Latreille), are quite common in the marshy mud until about the month of October, when again the pool gets filled up by the rains, thus giving place to the other set of animals. Since the soil is wet in the middle of the pool even

during the summer, species like *Lumbriconereis* spp., *Phytocetes gangeticus*, and the Capitellids do not actually perish during drought, but remain alive and inactive in the wet mud.

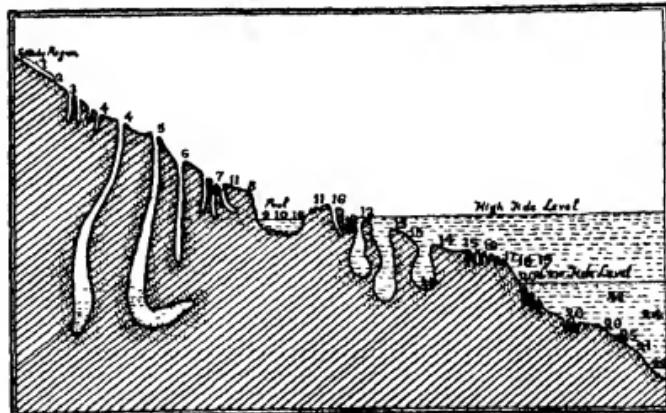
The pools belonging to the third category are much smaller in area and poorer in fauna than those described before. The following species are noted from them —

- Lycastis indica*
- Lumbriconereis* sp
- Stenothyra blanfordiana*
- Amnicola stenothyrodes*
- Indoplanorbis exustus*
- Melania tuberculatus*
- Mosquito larvae (Culex group)
- Chironomid larvae

In addition to these species, tadpoles and adults of the common frog *Rana cyanophlyctes* Schneider have also been met with in these pools during the rainy weather.

#### Vertical Distribution of Animals

I. Inhabitants of the Mud-flats — The vertical distribution of the inhabitants of the mud-flats is represented in an imaginary section passing



TEXT FIG. 2

through a mud-flat. The different levels at which the burrowing species occur are indicated in the figure. All the species marked do not occur in any one particular locality. The groups of animals that show a distinct zonation in the inter-tidal zone are the crabs. The Ocypods, *O. cordimana* and *O. macrocera*, are the species that occur farthest from the water-level near the bar, in other places the corresponding species is *Sesarma tetragonum*, which inhabits deep burrows. The habits of the Ocypods and those of *Sesarma* are by no means similar, for, while the former mostly remain outside the burrows, returning into them only for safety, the latter rarely come out. *Gelasimus (Uca) annulipes* and *Sesarma quadratum* inhabit burrows above water edge. In all the burrowing crabs, the depths of the burrows are such that they run on a line with the low tide level so that there will be a little water in them even during low tide. Among the Polychaetes, *Lycastis indica* and the Capitellids are often observed above the water-level. *Chibanarius padavensis*, *C. olivaceous* and *Scylla serrata* are inter-tidal in habits, but the last mentioned is a swimming form and hence cannot be said to have any restricted occurrence.

**II. Plankton.**—The following notes on the plankton of the Adyar backwaters are based upon a number of collections made between November 1933 and October 1936. An ordinary muslin tow-net was used for making collections, which, on the average, were made once a fortnight. During the August January period when the planktonic organisms are particularly noticeable, tow-netting was done almost every week during the years 1934 and 1935. On the other hand, the number of collections made during the summer months is rather low, and often, the tow-net water yielded nothing. Planktonic study not being our main objective, the observations made here are intended only for pointing out some of the more important features of the plankton. We have not paid attention to the study of the Protozoa, of the Copepods, only the commonest species have been noted. One great difficulty experienced was in tow-netting small pools of brackish-water, these pools have been eliminated from the study, but several collections have been made from the large salt pool near the Boat Club. There is not the least doubt that variations take place from year to year in the constituent elements of the plankton in the different months depending upon the changes in the rainy season and their influence on the time of opening and closing of the bar. An approximate idea of the nature of the plankton during the different months may be gathered from the following summary of our records.—

November	Bar just open	
	<i>Acromitus flagellatus</i>	Ephyrae Common
	<i>Campanulina ceylonensis</i>	Rare
	<i>Rhopalophtalmus egremus</i>	Swarms in 1933
	<i>Mesopodopsis orientalis</i>	Swarms
	<i>Penaeus carinatus</i>	Post-larval stages Common
	<i>Penaeus indicus</i>	Post-larval stages Common
	<i>Clupeid larvæ</i>	Common
	<i>Therapon jarbua</i>	Young ones
	<i>Acentrogobius neilli</i>	Larval and Post-larval forms
	<i>Aplocheilus melastigma</i>	Young and adults
December	Bar open	
	<i>Acromitus flagellatus</i>	Ephyrae Common
	<i>Dicyclocoryne filamentata</i>	Rare
	<i>Penaeus carinatus</i>	Post-larval and young ones Plenty
	<i>Penaeus indicus</i>	Post-larval and young ones Plenty
	<i>Mesopodopsis orientalis</i>	Common
	<i>Pecilimenes indica</i>	Rare
	<i>Cirripede nauplius</i> (Probably of <i>Balanus amphitrite</i> )	Common
	<i>Pseudodiaptomus annandalei</i>	Common
	<i>Ocypode megalopæ</i> ( <i>Ocypoda cordimana</i> ?)	
	<i>Therapon jarbua</i>	Young ones
	<i>Therapon quadrilineatus</i>	Young ones
	<i>Clupeid larvæ</i> (Engraulis and <i>Stolephorus</i> )	
	Young mullets	
	<i>Aplocheilus melastigma</i>	Post-larval and young ones
January	Bar open	
	<i>Acromitus flagellatus</i>	Young ones Plenty
	<i>Campanulina ceylonensis</i>	Rare
	<i>Penaeus carinatus</i>	Post-larval and young ones Common
	<i>Penaeus indicus</i>	Post-larval and young ones Common
	<i>Pecilimenes indica</i>	Rare
	<i>Pseudodiaptomus annandalei</i>	Common
	Post-larval Goboids	Common
	<i>Aplocheilus melastigma</i>	Young and adult Common
	Young mullets	Rare
February	Bar open, but very narrow	
	<i>Campanulina ceylonensis</i>	Rare
	<i>Penaeus indicus</i>	Young ones Common
	<i>Penaeus carinatus</i>	Young ones Common

Young mullets	Rare
<i>Periclimenes indica</i>	Rare
<i>Veliger larva</i>	Rare
<i>Mesopodopsis orientalis</i>	
March	Bar almost closed, but sea-water may get in during high tide
<i>Aplochelus melastigma</i>	Young ones Common
<i>Acartia southwells</i>	Common
Penaeids	Young ones Rare
<i>Veliger larva</i>	Common
April	Bar closed
<i>Campanulina ceylonensis</i>	Rare
<i>Acartia southwells</i>	Plenty
<i>Pseudodiaptomus annandalei</i>	Rare
<i>Veliger larva</i>	Common
<i>Mesopodopsis orientalis</i>	Rare
May	Bar closed
<i>Periclimenes indica</i>	Rare
<i>Pseudodiaptomus annandalei</i>	Rare
<i>Veliger larva</i>	Plenty
June	Bar closed
<i>Campanulina ceylonensis</i>	Common
<i>Veliger larva</i>	Common
<i>Periclimenes demans</i>	Rare
<i>Periclimenes indica</i>	Common
<i>Mesopodopsis orientalis</i>	Common
<i>Palamoneutes hornelli</i> ?	Rare
July	Bar closed
<i>Campanulina ceylonensis</i>	Common
<i>Acalous planarian</i> ( <i>Convoluta</i> sp?)	Rare
<i>Acartia southwells</i>	Common
<i>Periclimenes indica</i>	Common
<i>Mesopodopsis orientalis</i>	Common
<i>Pseudodiaptomus annandalei</i>	Rare
<i>Leander</i> sp	Young ones Common
August	Bar closed
<i>Campanulina ceylonensis</i>	Common
<i>Acalous planarian</i>	Large numbers
<i>Periclimenes indica</i>	Common
<i>Mesopodopsis orientalis</i>	Common
<i>Aplochelus melastigma</i>	Young ones
<i>Acentrogobius neilli</i>	Young ones

*September* Bar closed

- Acromitus flagellatus* Ephytæ
- Campanulina ceylonensis* Swarms
- Copepod nauplius* Large numbers
- Pseudodiaptomus annandalei* Common
- Acaria southwellsi* Rare
- Acarous planarian* Rare
- Penaeopsis monoceros* Very young ones Common
- Mesopodopsis orientalis* Common

*October* Bar usually closed, but may open during this month

- Acromitus flagellatus* Ephytæ Common
- Campanulina ceylonensis* Common
- Copepods Numerous (Several species)
- Mesopodopsis orientalis* Common
- Acentrogobius neilli* Young ones Common
- Acentrogobius viridipunctatus* Young ones Common
- Periclimenes indica* Rare
- Penaeopsis monoceros* Young ones Common

#### General Conclusions on the Fauna

The list of animals given previously will clearly illustrate the predominantly marine character of the fauna of Adyar and the adjoining brackish-waters of Madras. Of about 92 species of Invertebrates noted, more than eighty species belong to groups of animals that are typically marine. The freshwater species which have more or less acclimatised to the brackish-water conditions are thus comparatively few here, this constitutes the most important difference between the fauna of an estuarine brackish-water tract and that of a salt marsh. In the latter, the animals which are of typical freshwater habitat, and terrestrial species that have secondarily taken to an aquatic mode of life are quite numerous and show varying degrees of adjustment to saline environments. In the Adyar fauna, the noteworthy groups of freshwater animals which have secondarily taken to brackish-water life are the members of the family Hydrobiidae among the Mollusca and some of the Palaeomidae among the Crustacea.

Among the 64 species of fishes, only seven freshwater forms are noted. The others include several casual or seasonal migrants from the sea, while the majority of them are more or less permanent inhabitants of the back-water. The true brackish-water species have been caught when the bar is open and when it is closed, and in most of them, the young as well as the adult fish have been met with. Marine species like *Stolephorus commersoni*,

*Sphyraena jello*, *Serranus sexfasciatus*, *Lutjanus* sp., *Therapon quadrilineatus*, *Therapon puta*, *Upeneus sulphureus*, *Scatophagus argus*, *Caranx cirratus*, *Pseudorhombus javanicus*, *Parsus* and *Triacanthus brevirostris* have been obtained only during the months when the bar is open and the salinity high. The freshwater species are the two Chichids, *Etioplus maculatus* and *E. suratensis*, and *Macrones vitellatus*, *Panchax parvus*, *Aplocheilus melastigma*, *Barbus sophore* and *Barbus dorsalis*. All of these species breed in brackish-water.

#### Breeding Habits

Many species inhabiting the Adyar backwaters are capable of breeding all through the year, but a period of intense reproductive activity is seen in many of the inhabitants. This is determined by the extent to which the environmental conditions are suitable for the breeding of any particular animal. Thus most fish cannot breed when the backwater and the river are partially dried up, nor can the Gastropods with their densely packed gelatinous egg masses breed under flooded conditions. Ripe sex cells or embryos have been observed in the following animals during all months of the year —

- Pelocretes exul*
- Phytocetes gangeticus*
- Phytocateopsis ramunni*
- Stephensonacis ornata*
- Marphysa grisea*
- Lycastis indica*
- Lumbiconereis* sp.
- Diopatra variabilis*
- Mesopodopsis orientalis*
- Grandidierella magna*
- Grandidierella gilesii*
- Paracalliope fluviatilis*
- Clibanarius padavensis*
- Clibanarius olivaceous*
- Potamides cingulatus*
- Stenothyra blanfordiana*
- Amnicola stenothyroides*
- Stiliger gopala*
- Meretrix casta*
- Cymothoa indica*
- Cirolana* sp.
- Aplocheilus melastigma*.

*Panchax parvus.*  
*Etroplus maculatus*  
*Etroplus suratensis*  
*Acentrogobius neilli*  
*Acentrogobius viridipunctatus*

The groups of animals that show great intensity in reproductive activity during particular seasons are the crabs, Molluscs and the fishes. Among the Brachyura, only the Grapsoid and Ocypod crabs breed in the brackish-waters of Madras. Ovigerous females belonging to different species have been collected from December to March. The Pagurids are perennial spawners. Among the Caridea, ovigerous females of *Perulimenes indica* have been obtained only from April to July. Very young ones of *Alpheus malabaricus* are very common in the backwater during January and February, and this species also appears to breed under brackish-water conditions. *Penaeus indicus* and *P. carinatus* do not attain sexual maturity in the backwater, but their young ones are noticed in fairly large numbers during the months when the bar is open (compare plankton records)\*. In contrast with this is another Penaeid, *Penaeopsis monoceros*, which appears to breed in the brackish-water as judged by the fact that numerous young specimens 15 to 20 mm long are obtained in tow-net collections taken in September and October, about six months after the bar is closed.

Most of the fish show an intense reproductive activity soon after the monsoon in November. Large numbers of young ones of *Acentrogobius neilli*, *A. viridipunctatus*, *Gobius* sps and *Mugil* sps have been obtained in tow-nets during November and December. Along with them are found larval and post-larval stages of Clupeid fish which are brought into the backwater from the sea. Young ones of *Aplocheilus melastigma* are obtained during almost all the months of the year, especially a day or two after any shower. Rains have a remarkable influence over the breeding in most fishes. Species like *Acentrogobius neilli*, *A. viridipunctatus*, *Etroplus maculatus*, *E. suratensis*, *Panchax parvus*, *Aplocheilus melastigma*, etc., spawn intensively even during the summer months if there be occasional heavy rains.

The optimum season for breeding of Molluscs is the summer. During the months March, April, May, June and July, the egg cases of many Gastropods are common in the backwater. Large numbers of young ones of *Potamides cingulatus* and *Meretrix casta* are noticed in July and August. *Stenothyra blanfordiana*, *Amnicola stenothyrodes* and *Stiliger gopala* exhibit

\* For habits of brackish-water prawns, *vide Panikkar, 1937 b.*

enormous powers of reproduction, though they breed all through the year, they have periodical phases of intensive propagation determined by the monsoon, salinity and other environmental conditions

The subject of breeding in the common brackish-water species of Madras is dealt with in detail in another study by us<sup>†</sup>. The summary of our observations is given below—

1 Breeding of animals of the brackish-waters of Madras is not particularly confined to any definite part of the year, actively reproducing species are met with during all seasons

2 The following types of breeding are noted—

- (a) Continuous breeding throughout the year occurring more or less uniformly and irrespective of seasons
- (b) Continuous breeding with a marked season of higher activity during one part of the year than the remainder
- (c) Breeding season confined to some definite part of the year
- (d) Discontinuous breeding occurring all through the year, spawning often taking place irregularly, mostly determined by the rains

#### *Discussion*

*General Considerations*—The interesting feature of the fauna of the brackish-water is the intimate association of animals of the sea, backwater and freshwater for life in a common habitat. The animals of marine origin constitute the largest number of species as most of them are either present in the sea or are represented there by closely related species. The number of marine animals that have invaded the brackish-water is greater in the tropics than in temperate regions. Annandale (1922) mentions that the aquatic fauna of Europe is easily divisible into the freshwater and marine faunas, and that with the exception of *Paleomonetes varians* in brackish-water in North Europe, and *Mysis relicta* and a few other species of estuarine Molluscs in different parts of Europe, the separation of marine and freshwater faunas is a constant feature. Recent studies of the brackish-water fauna of Europe have shown, however, that this statement of Annandale is not altogether correct. The contributions of Redeke (1933) on the fauna of the Zuider Sea, of Remane (1934) on the Baltic Sea, of Sick (1933) on the brackish ditches of North Germany, and of Brandt, Wundsch (1933), Lundbeck (1932), Sergeant (1934) and others on the different brackish tracts of Europe have shown that a fairly large number of characteristic marine and euryhaline species exist in Europe inhabiting waters of low

<sup>†</sup> "Observations on Breeding in Brackish-water Animals of Madras" By N. Kesava Panikkar and R. Gopala Aiyar (Unpublished).

salinity. The conditions in England are also more or less similar as shown by the studies of Allen and Todd (1900 and 1902), Percival (1929), Stammer (1928), Ellin (1932), Robson (1925) and Fraser (1932) on the estuarine fauna, of Nicol (1936) on the fauna of the brackish lochs of Scotland, and of Lambert (1930) and Nicol (1935) on the animal life of the salt marshes adjoining sea coasts or estuaries. It is clear from all these papers that a more or less distinct brackish-water fauna is present in the temperate regions as well. It must, however, be admitted that as compared to the tropics, the brackish-water species are fewer in numbers and less in variety in the land-locked seas, estuaries, backwaters and salt marshes of the temperate regions. As pointed out by several observers, the penetration of a large number of marine animals into waters of low salinities is a characteristic feature of the tropics, and the numbers of species inhabiting brackish-water are so many and representative of almost all the major marine invertebrate phyla except the Echinodermata. An interesting fact regarding the tropical brackish-water fauna is the occurrence of numerous adaptations in the mode of life of animals of marine origin, which serve to facilitate their life in these peculiar environs.

*Acclimatisation of Marine Animals to Fresh and Brackish-Waters*—It is almost universally accepted that the present freshwater fauna has been derived to a large extent from ancient marine animals. Many of them have penetrated into freshwater through media of progressively decreasing salinity as found at the meeting places of large volumes of freshwater and sea water like the openings of big rivers or backwaters. Most of the modern brackish-water organisms are forms that migrated from the sea into the brackish-water at a very recent geological epoch, often, we find along with them, numerous marine species capable of surviving in the brackish-water. We find in every tropical backwater, tidal creek, or estuary, an active and aggressive attempt on the part of many marine species to secure a permanent foot-hold either in the semi-aquatic mud-flats and mangrove swamps or in the salt marshes with peculiar environmental conditions of fluctuating salinity, temperature and hydrogen-ion concentration. Many explanations have been suggested to explain this peculiar phenomenon of the tropics, but as pointed out by Annandale (1922) the factors contributing to this must necessarily be varied, and it is perhaps necessary to study the known instances individually before any definite view is taken. The conditions necessary for the successful colonisation of the brackish-water are many and may now be considered.

Sollas (1883 and 1905) in his classical discussion on the origin of freshwater faunas, emphasized the current strength of rivers as the chief obstacle

to the progress of marine animals into freshwater. He pointed out that an animal must either be fixed or strong enough to withstand the current of streams if it should establish itself in fresh-water. This must be true of the larvae as well as the adults, the epiplanktonic elutated larvae of marine Invertebrates being extremely unsuitable for life in a flowing stream. Hence, only those animals as have got rid of the free-swimming larval stages, by an abbreviation of the life-history, were successful in establishing in the freshwater. The absence of free eggs and larvae from the freshwater plankton is explained by this theory, and the larger sizes of eggs of freshwater animals as compared with their marine relatives is also explained as an attempt at further acquisition of yolk to cope with the curtailment of life-history. While explaining some of the important differences between freshwater and marine animal life, this theory does not explain the difference between the temperate and tropical conditions. This was attempted much earlier by von Martens (1868) who emphasized that the freshwater environment with its alternative liability to periods of freezing and desiccation, is more severe than that of the sea. The difference between the cold and hot seasons is very great in the cold countries, while a more even temperature conditions prevail in the tropics, hence acclimatisation was easier here than in the cold countries where extreme temperature conditions prevail. In addition to these two explanations, Needham (1930) made the interesting suggestion that there is a third factor limiting the penetration of marine animals into freshwater, viz., that of the inorganic deficiency of the freshwater from the point of view of the larval development of marine animals. Based on the experiments of Ponchet and Chabry (1889), Herbst (1897), Rapkine (1927), Ranzi (1930) and his observations on the phosphate contents of developing eggs, he pointed out that the developing planktonic larvae of marine Invertebrates depend upon the inorganic contents of sea-water for certain items of nutrient. Proper development and metamorphosis would be impossible without the supply of calcium, phosphates, etc., inorganic materials not available in freshwater. Penetration into freshwater is possible only for those marine animals that have so perfected their development as to be independent of the environment for their inorganic requirements.

The three theories summarised above overlooked another important aspect of the problem—the question of osmotic regulation of marine animals on which attention has been focussed in recent years by the work of Schleiper (1929, 1929 a, 1930 and 1935), Dakin (1908, 1908 a, 1931 and 1935), Schwabe (1933), Adolph (1925, 1926 and 1930), and others. The most important physiological difference between freshwater and marine organisms is that in the osmotic pressure relative to the external medium. The body fluids

of marine invertebrates have almost the same osmotic pressure as that of the sea-water in which those animals live (Schleiper, 1930), in many instances, this pressure is slightly higher than that of the surrounding medium as shown by recent investigations (Dakin, 1935). These animals allow a free interchange of body fluids with the sea-water, and changes in salinity of the external medium hence affect the osmotic concentration of the body fluids. All freshwater and numerous brackish-water animals, as also the marine fishes, have developed a power of regulation of osmotic pressure that keeps their body fluids in a permanent state of hypertonicity irrespective of the low concentration of the medium that bathes the animal. When the salt content of the environment is lowered, the higher osmotic pressure of the body fluids of marine invertebrates is not maintained owing to the passage of water into the interior and loss of salts by diffusion. This would go on until a state of isotonicity is reached, a condition in which the animal may not survive unless a greater concentration of the body fluid is maintained by some regulating mechanism. Thus the colonisation of brackish-water and freshwater is made possible only with the development of osmoregulatory powers as poikilosmoticity would be fatal in hypotonic media.

Exact data are not yet available regarding the energy requirements for the maintenance of osmoregulatory powers in aquatic animals. In several cases, however, there is a relationship between the salt content of the external medium and the oxygen consumption. As shown by Schleiper (1931), Beadle (1931), Lowenstein (1935) and others, there is a rise in the amount of oxygen consumed when marine animals are transferred to media of lower concentrations, the rise gradually goes on until it reaches a maximum and then remains constant at a level above the normal. The deleterious effects of oxygen tension combined with reduction in salinity of the environment have been clearly demonstrated by Schwabe (1933) for the crab *Carcinus maenas*. This increase in oxygen requirements has a great bearing on the problem of survival in the brackish-water. Integumental regulation plays an important rôle in the maintenance of the steady state. Pantin (1931) has observed that the presence of calcium in the water is of considerable advantage to the estuarine Platyhelminth *Gunda silvæ* in overcoming the adverse effects of salinity fluctuations. All these point to the conclusion that the factors involved in the entry of marine animals into brackish-water and freshwater are many and cannot easily be reduced to any single theory.

In the light of what has been said above, the relative advantage of the tropics for the inward migration of marine organisms may be explained in

being due to various reasons. We consider that the temperature factor is of the greatest importance, for, in addition to the relative evenness as suggested by von Martens, the differences between the air, freshwater and ocean temperatures are comparatively small in the tropics. Not only was this advantageous in the successful colonisation of brackish and freshwater, but also in the assumption of amphibious and terrestrial modes of life by several littoral marine animals (cf. Pearse, 1929 and 1936). The greater rainfall of the tropics and the presence of large rivers bringing down an immense amount of fresh water into the sea, have considerably altered the coastal salinities of the tropics and this has endowed the tropical marine animals with great powers of adjustment as they are under the constant influence of salinity fluctuations. Sewell (1934) shows that this lowering of salinity at the head of the Bay of Bengal where the Ganges and Brahmaputra open, affects even up to a depth of about fifty fathoms. The great change in the coastal salinity as a consequence of the monsoon in the coasts of Indo-China is also mentioned by Pearse (1922 a). The effects of this prolonged acclimatisation of marine animals to frequent changes in salinity would be a gradual change in their physiology from stenohalism to euryhalism, and from poikilosity to varying grades of homioosmotic behaviour, depending upon the species concerned and the exact conditions of the environment. Only those species of marine invertebrates which have thus responded to the influence of the environment by a gradual assumption of independence over the osmotic concentration of the environment have succeeded in colonising brackish-waters. The inorganic materials washed down into the sea by the intense rainfall and the numerous large rivers, support a rich Diatom flora which increases the food resources of the coasts and thereby attract numerous marine animals (Sewell, 1934). From the description of the area on which this study is based, it will be clear that the coastal salinity is not lowered to any appreciable extent, there being no large river in the vicinity of the city of Madras, and the rainfall being comparatively poor. The conditions here would seem to favour the temperature factor more than anything else, since the brackish-water fauna is as representative and predominantly marine as that of the Gangetic Delta or of the Chilka Lake.

*Integumental Adaptations*—The changes attendant upon the migration of marine animals into brackish and fresh waters are many, but from the point of view of their morphology, the important among them are the integumental adaptation and the strengthening of the respiratory system by the development of accessory structures to cope with increased oxygen requirements. Most brackish-water animals have well-developed

mucus secreting devices which effectively guard against sudden changes in salinity by preventing the passage in or out of water for a certain period. Paul Bert (1871 and 1886) made the interesting discovery that eels, which were carelessly handled so that the mucus was completely removed, were no longer able to withstand sudden changes from fresh-water to sea-water and *vice versa* (Dakin, 1935). This has been corroborated by the interesting series of experiments of Duval (1925). The power of mucus secretion is widely prevalent amongst aquatic animals, but it is especially well developed in euryhaline and brackish-water species. Mucus-cells in the body-wall are remarkably developed in the brackish-water *Hakampatids* of Adyar, but they are comparatively few and insignificant in a related marine species (Panikkar, 1936, 1937 and 1937 c). The Polychaetes, *Lycastis indica*, *Diopatra variabilis* and certain others occurring here have also got mucous glands on the body-wall which are remarkably developed, while there is not a single Gastropod of Adyar that is not capable of copious mucus secretion. Many species remain enveloped in mucus whenever environmental conditions are adverse, similar to what Duerden (1906) observed in certain corals. The prominence and wide-spread occurrence of such external secretory structures in brackish-water species, and the way they are brought into action when animals are transferred to media of varying salinities, would indicate that they are of great advantage to these animals. In the case of species that are under the constant danger of being left exposed for fairly long periods, the presence of mucus would also undoubtedly prevent desiccation.

*Respiration*—Several observers have shown that the oxygen content of shallow brackish-waters and salt marshes is low as the temperature is high in small volumes of water. Unfortunately, we have not made studies on the oxygen content of the water, but having observed the high thermal limits to which the pools of brackish-water get heated up, and the occurrence of decaying matter in the river and backwater, there is reason to believe that the oxygen present in the water is low. This raises another problem of survival of the brackish-water organisms. The predominance of the amphibious element in the Adyar fauna was emphasized in an earlier section, it was shown that the species inhabiting the marginal zone or the actual water edge are many and varied, and larger in communities than those that inhabit the deep water and rely completely on aquatic respiration. The animals which have developed extensive means of aerial respiration are the crabs and fishes. The proportion of oxygen to the total volume is much higher in air than in water while the oxygen content of the air is about 20%, water under normal conditions of temperature holds only

5 to 10 c.c. of oxygen per litre. Winterstein (1921) has indicated that air is more favourable for oxygen absorption than water which, in turn, is better suited than air for carbon dioxide liberation (cf. Carter, 1931). In general, the respiratory changes that we find in brackish-water animals are caused by the growth of organs of aerial respiration, and not the increased development of gills for aquatic respiration. According to Carter and Beadle (1931) the accessory respiratory organs usually developed in fishes are chiefly organs of oxygen absorption, while the original gills discharge the function of carbon dioxide liberation. These additional structures are internal in those animals since they still live in an aquatic medium, in shallow water or at the water edge. The changes that have taken place in animals which became slowly adapted to respire in air are (1) the development of additional structures like arborescent folds, etc., (2) devices to maintain the dampness of the epithelium, and (3) devices to prevent evaporation of water from the respiratory surface. Amongst the Adyar fishes, organs for aerial respiration are present in the form of vascularised walls of gill chambers, gill septa, etc., in *Periophthalmus kuhli*, *P. pearsei* and *Boleophthalmus boddarti*. In the last species, the body is covered by numerous minute papillæ which have a respiratory function (Harms, 1929). The Amphipods of Adyar, though amphibious in habits, do not seem to have any special structural modification in their respiratory organs. The Isopod *Ligia exotica*, though depending completely on aerial respiration, is not found far away from the water edge. The thinly chitinized endopodites of pleopods in the aquatic Isopods function as lung in the case of terrestrial species (cf. Verhoef, 1919). Bepler (1909) finds that they are provided with glands at their bases, the secretion of which keeps them moist. The air-breathing capacities of several Decapods are well known, among the crabs, we find a series of adaptations intended for this purpose. Structural adaptations are absent in Macrourous species, though some forms, like *Penaeopsis monoceros* are capable of remaining outside water in a healthy condition for fairly long periods. The Anomura of Adyar are all amphibious and respire partly in air. The gills are normal in most Decapods except in completely terrestrial genera. The Grapsidae and the Ocypodidae, which are either amphibious or are completely terrestrial, show a common modification directed to retaining water in the gill chamber (Carter, 1931). The abnormally developed hairs near the opening of the gill chamber in *Sesarma*, *Cardisoma* and *Uca* (*Gelasimus*) (Ortmann, 1901), the spongy structures on the walls of the gill chamber in *Gecarcinus* and *Ocypoda* (Winterstein, 1921), and the peculiar fold at the floor of the gill chamber in *Uca* (Jobert, 1876) are all designed for this purpose. Enlargement of

the gill chamber is seen in many brackish-water species like *Gecarcinus* (Calman, 1911), *Uca* and *Cardiosoma* and an upper lung chamber and a lower gill chamber are distinct in *Gecarcinus* (Calman, 1911, Carter, 1931) Vascularised epithelial folds that function as accessory organs are met with in the gill chambers of *Gelasimus* (*Uca*) (Jobert, 1876), *Cardiosoma* and *Ocypoda* (Zimmer, 1926-27), and lastly, special apertures that assist the entrance of air are found in the chambers (usually the posterior part) of *Ocypoda*, many *Catametopa*, and *Grapsus* (Muller, 1863, Ortmann, 1901) All these modifications have not much affected the gills proper, and as a rule, the gill chambers are provided with air in the case of air-breathing forms, and not water Carter (1931) suggests that the need for adaptations designed to retain water in the gill chamber is probably caused by the necessity to keep the vascular folds damp

*Exposure and Desiccation*—We have found from field observations as well as experiments with several species, that many of the brackish-water Invertebrates of Adyar can survive exposure to air for a longer period than they can remain in freshwater Exception should, however, be made of the species that have a wide range of regional distribution from the sea to almost freshwater This fact explains the predominance of the amphibious element in the fauna of the brackish-waters of the type described here In many terrestrial species life under water is impossible even though these species may be found only in the damp zone just above the water edge Many of the amphibious forms like *Clibanarius padavensis* and *Clibanarius olivaceus* cannot thrive under water for long periods The result is that these species have to keep moving to and fro so as to remain at the water edge when changes in level take place by tidal action It would appear from numerous examples that in brackish-water animals, the tendency to become terrestrial is more marked than that to become freshwater inhabitants Our observations are in agreement with the suggestion made by Pearse (1929) that the number of animals which have reached the land from the littoral marine and from brackish-water is much higher than the number of species that have become fluviatile

The brackish-water areas of the Indian coasts are subject to periodical desiccation in the summer, and the problem of survival during the period of drought is another important factor that the brackish-water animals have to cope with It is the amphibious and the attached animals which are affected by drought For the first group, which consists mostly of burrowers, it is difficult to leave their original burrows and migrate downwards, but this is actually done by several crabs like *Uca annulipes* and *Sesarma quadratum* Species of *Clibanarius*, *Ocypoda cordimana* and *Scylla*

*serrata* exhibit what may be called migratory movements in response to seasonal changes. During summer, these species are practically absent from the fringes of the backwater, but congregate in large numbers on the southern bank of the river. Most of the attached forms like *Companularia noliformis*, *Laomedea spinulosa*, *Balanus amphitrite*, *Hydrodes norvegica* and *Aiptasiomorpha* sp. perish during the summer, but fresh individuals settle down every year when the bar is open. The periodic inactivity or aestivating habit of the two Grapeoid crabs *Varuna littoralis* and *Sesarma tetragonum* has been described by Hora (1933), of these, the former is extremely rare at Adyar during the summer, but *S. tetragonum* which occurs in large numbers in the banks of the backwater and the Cooum, behaves in the same manner as in the Gangetic Delta described by Hora. We may add to this list another species, *Sesarma quadratum* which also retires into the burrows during the summer and remains more or less inactive. It may also be mentioned here that none of the burrowing crabs has been observed to breed during the summer months. The Gastropod *Potamides cingulatus* is capable of surviving exposure to air and sunlight for long periods, and specimens have on several occasions been collected far away from the water in an inactive state but which become active again on removal to water. The anemone *Phylloctetes gangeticus* remains alive in the mud outside water for weeks together, the anemones do not perish so long as the mud remains a little moist (Panikkar, 1937). Many sedentary brackish-water organisms are known to have evolved peculiar methods of asexual reproduction, reminding one of the gemmule formation of the freshwater sponges, whereby the survival of the species is ensured, the well-known instances are Hydroid *Annulella gemmata* (Ritclie, 1915), the sponge *Laxosuberites lacustris* (Annandale, 1915 a), and the Polyzoan *Loxosomatoides laris* (Annandale, 1915 b and 1922). None of these species has so far been observed here, and perhaps the only instance of active asexual reproduction is that of a sea anemone *Boloceractis gopala* where there is a peculiar method of tentacular regeneration (Panikkar, 1937 a). This cannot, however, be considered as having any bearing on the habitat as similar features have been noticed also in a related marine genus.

*Abbreviation of Life-History*—The disadvantages of the free-swimming ciliated larvae in the brackish and fresh-waters have already been pointed out. The changes in reproductive habits consequent upon marine animals taking to a brackish-water mode of life are interesting as they involve curtailment of life-history in several of the species, especially the invertebrates. This is very well illustrated by the life-histories of certain brackish-water Polychaetes *Marpissa grisea* (Aiyar, 1931) and *Diopatra variabilis*

(Krishnan, 1936) have been studied in detail from Adyar, in both the species an abbreviated development has been observed by the above authors. In the first, the eggs are laid in large gelatinous egg cases anchored to the worm-burrows and harbouring thousands of eggs in various stages of development, the jelly serving the three-fold purpose of preventing the larvae from being washed into the sea, of protecting them from drought when they are exposed during low tide, and of catering nutrient to the large number of rapidly developing neotochaetes. The richly yolk-laden gigantic eggs of *Diopatra variabilis*\* metamorphose into the young worms even in the tube itself, without the intervention of a free swimming larva. The development of this species is characterised both by precocity and by increased provision of food yolk in the eggs as compared to the other species of the same genus, and the developmental differences of very closely allied species could only be explained on the basis of the differences in environmental conditions. Gigantic eggs are also found in *Lumbrineres* sp., also a genus in which Fewkes (1883-85) observed an abbreviated development. The collection of eggs in gelatinous egg-masses is a feature common to most of the Gastropods noticed at Adyar and to certain other species of Polychaetes like *Scoloplos* sp. Among the Gastropods there is no doubt a free-swimming stage in the life-history, but this phase is often extremely short. Interesting details of an extremely short larval stage and rapid metamorphosis have been observed by Rao in the Nudibranch (Ascoglossa) *Stiliger gopala*.† Most of the species of fish that breed at Adyar have demersal eggs, and though specimens of Clupeid and other groups of fish are commonly caught in the brackish-water as the list of animals collected by us would show, none of them breed in the brackish-water.‡

*High Rate of Reproduction*—Apart from the modifications in development and the asexual reproductive devices, there is another aspect which has not received the proper attention it deserves. This is the high intensity of reproductive activity of many of the species. As in most tropical animals (cf. Orton, 1920) many species found at Adyar would appear to be able to breed all through the year, though a well-marked season for breeding is noticeable in many of them in as much as a high rate of reproduction is noticeable during this period. As pointed out before, this season is determined by the optimum environmental conditions for any particular species.

\* Fauvel (1931) considers the species as being synonymous with *Diopatra neapolitana*.

† "Habits, Structure and Early Development of a New Species of *Stiliger*," by K. V. Rao (1937), unpublished.

‡ *Engraulis* *purpureus* is a possible exception, *vide* Raj, 1916.

The high rate of sexual propagation accounts for the high intensity of population of animal communities in certain biotopes. The animal communities also often prove the correctness of Thienemann's rule. Most of the species attain sexual maturity at a very small size, well-developed sex-cells are observed in very small individuals of *Potamides cingulatus*, *Sesarma quadratum*, *Acentrogobius neilli*, *Clibanarius olivaceus*, *Metasesarma rousseauxii* and *Meretrix casta*. Rao (*op. cit.*) finds that in *Stiliger gopalas*, sexual maturity is attained within a very short time after metamorphosis and several generations of these Gastropods are produced with singular rapidity. In the Anemone *Phytocetes gangeticus* where the early development up to the Edwardsia stage is rapid, the post-Edwardsia stages are rather prolonged and the gonads develop even in the larval Edwardsia stages with the incomplete quota of nucleotenes. This instance of probable nucteny is further confirmed by the presence of blastula-like stages in the coelenteron of post-larval individuals. It is probable that the brackish-water habitat may, in some measure, be responsible for this capacity for juvenile reproduction (Panikkar, 1937). Almost every species of invertebrate from Adyar investigated so far, has shown a definite tendency towards early sexual maturity and rapid development, and other reproductive changes like the assumption of hermaphroditism have been observed in the Polychaete *Lycasis sindica* (Aiyar, 1938).

*Classification of Faunistic Elements*—A convenient basis of classification of animals living in an environment of variable features as the brackish-water is necessary to estimate the exact ecological characters of the different species. Based on the chlorine content of the water, Redeke (1922 and 1931) suggested a division of the brackish-water species into oligohaline, mesohaline and polyhaline groups, the first representing the least saline region (salinity 0.2-1.9 per cent, Cl, 0.1-1.0 gm per litre) adjoining the freshwater, the second mesohaline region forming the middle zone (salinity 1.9-18.6 per cent, Cl, 1.0-10.0 gm per litre), and the last polyhaline region merging into the sea-water (salinity 18.6-31.8 per cent, Cl, 10.0-17.0 gm per litre). He pointed out that these differences in the chlorine content correspond exactly to the differences in the fauna, irrespective of the type of brackish-water investigated. However much this classification may be satisfactory when applied to large volumes of brackish-water like the Baltic Sea, the Black Sea, the Chilka Lake, etc., this division ceases to have any value for small bodies of water as suggested by Redeke himself, and as conclusively shown by Nicol (1936) for salt marsh pools. At Adyar, the grouping of animals according to Redeke's method is not at all applicable since every sudden outbreak of rain in the locality would, especially

when the bar is closed, result in quick changes from polyhaline to oligohaline or almost lacustrine conditions, and the most interesting feature of the fauna is the singular capacity of several species to thrive in environments liable to profound changes in salinity and temperature. Many of the burrowing invertebrates adjust to salinity changes by vertical migrations (cf. Panikkar, 1936). So far as our observations go, we have not found strict demarcation of animal communities exclusively on salinity basis. At Adyar, the distribution of species is controlled by a multiplicity of factors as the substratum, depth, current of water, vegetation, etc.

*Fauna of Adyar and of other Regions Compared*—Comparing the fauna of the brackish-waters of Madras with that of the Gangetic Delta and the Chilka Lake, it is clear that though the area is small and without the facilities for admixture of salt and freshwaters on a large scale, the fauna is as representative and almost as rich as in the two other places, especially in regard to true brackish-water species. The main difference lies in the fact that in the Chilka Lake and the Gangetic Delta, there are many species from the sea, which at Madras are exclusively marine in habits and are not found in the brackish-water. The species of *Phlyra*, *Pleurobrachia bengalensis*, *Membranopora* sp., *Akyonium mytilis*, *Thalassema* sp., *Squilla scorpio*, *Squilla interrupta* and *Lucifer hansenii*, to mention but a few examples, have not so far been observed at Adyar, though they are common in the Madras Coast, and in the brackish-waters of the Chilka Lake and the Gangetic Delta. The reason for this is that owing to the absence of a proper gradation of salinities, the small size of the area involved, and the pollution of water as the backwater is situated in the city, only the more resistant of the euryhaline species seem to have secured a foot-hold in the Madras brackish-waters. Also, we do not find here a rich fauna as found in the Gangetic Delta represented by recent marine forms that no longer occur in stenohaline or euryhaline conditions, but only in fresh or slightly brackish-water. The general features of the fauna described here are in agreement with those of the fauna of the Gangetic Delta and the Chilka Lake, but the constituent elements of the fauna present distinctive features in the occurrence of several characteristic species amongst the invertebrates, especially the Anemones and the Polychates. The proper evaluation of the fauna would, however, be possible only after exhaustive studies have been made of the fauna of the numerous other brackish-water tracts of India.

#### *Summary*

The fauna of the brackish-waters of the city of Madras has been studied in detail with special reference to its relation with the habitat

The environmental conditions are described in detail. About ninety-two species of Invertebrates and fifty-six species of Vertebrates have been observed in the brackish-water. The fauna is predominantly marine and includes representatives of all major marine Invertebrate phyla except the Echinodermata. The freshwater species include a few Crustaceans, Molluscs and fish. The amphibious element in the fauna is well marked, and a number of mud-burrowing species are noted. Vertical and regional distribution of the organisms is described and the groups into which they may be classified according to their habits are indicated. An account of the breeding in brackish-water species is given. The general problems concerning the biology of animal life in the brackish-water are discussed in detail with special reference to the fauna described.

#### BIBLIOGRAPHY

(This list is not exhaustive, and includes only the most important publications from the point of view of (1) Indian brackish-water Fauna, and (2) the biology of brackish-water animals in general. Many references not directly cited in the text have also been included for the sake of completeness within the above limits.)

Adolph, E. F. Some Physiological distinction between freshwater and marine organisms." *Biol. Bull. Woods Hole*, 1925, 48

— "The regulation of body volume in freshwater organisms." *Journ. Exper. Zool.* 1926, 43

— "Living Water," *Quart. Rev. Biol.* 1930, 5

Aiyar, R. Gopala "An account of the development and the breeding habits of a Polychaete worm of the genus *Marpheus*." *Journ. Ind. Soc. Zool. Landau*, 1931, 37

— "Observations on the development of *Lepturogobius nebulosus*," *Zool. Anz.* 1934, Bd. 111

— "Hermaphroditism in *Lycodes indica* Southern." *Curr. Sci.* 1935a, 3

Alcock, A. "Materials for a Circumlogeal fauna of India." *Journ. Anat. Soc. Bengal*, 1895-1900, 64-69

Alexander, W. B., Southgate, B. A., and Bassindale, R. "The salinity of the water retained in the muddy foreshore of an estuary." *Journ. Mar. Biol. Assoc.* 1932-33, 18

Allen, F. J., and Todd, R. I. "Fauna of the Fye estuary," *ibid.* 1900-02, 6

— "Fauna of the Salcombe estuary," *ibid.*

— "Fauna of the brackish ponds of Port Canning, Lower Bengal," *Rec. Ind. Mus.*, 1907, 1

— "The distribution and origin of the fauna of the Jordan system with special reference to that of Lake Tiberias," *Journ. Anat. Soc. Bengal*, 1915, 11

— "Fauna of the Chilka Lake. Polyzoa of the lake and brackish-waters of the Gangetic Delta," *Mem. Ind. Mus.*, 1915a, 5



Beadle, L. C. "Osmotic regulation in *Gunda ulva*" *ibid.*, 1934, 11  
 Bepler, H. 1909 Quoted by Carter, 1931  
 Bert, Paul "Sur les phenomenes et les causes de la mort des animaux d'eau douce qui l'on plonge dans l'eau de mer," *C R Acad Sci Paris*, 1871, T 73  
 \_\_\_\_\_ "Animaux d'eau douce dans l'eau du mer, animaux d'eau de mer dans l'eau dessalée, animaux d'eau sursalée," *C R Soc Biol Paris*, 1885, T 37 Quoted by Dakin, 1935

Bhattacharya, D. R. "Fauna of the Chilka Lake. Stages in the life-history of *Gobius*, *Pterosurtes* and *Hemiramphus*," *Mem Ind Mus* 1916, 5

Borradaile, L. A. "Fauna and Geography of Maldiva and Laccadive Archipelagoes," *Crustacea* 1930, 1  
 1897. Quoted by Dakin, 1935

Bottazzi, F. "Die Fauna der Ost-see, insbesonders die der Kieler Bucht," *Arch Deutsch Zool Ges.* 1897, 10 Quoted by Remane, 1934

Bruce, J. R. "A pH method of determining the carbon dioxide exchanges of marine, brackish-water and freshwater organisms," *Brit Journ Fisher Biol.* 1924, 2  
 \_\_\_\_\_ "Physical factors of the sandy beach I. Tidal, edaphic and climatic," *Journ Mar Biol Assoc.* 1928, 15  
 "Ibid. Part II Chemical changes, etc," *ibid.*, 1928

Brues, C. T. "Occurrence of the marine crab *Callinectes ornatus* in fresh and brackish-water," *Amer Nat* 1917, 51  
*Animal Life in Deserts*, London 1923

Buxton, P. A. *The Life of the Crustacea* London 1911

Calman, W. T. "Respiration of aquatic animals" *Biol Rev Camb.* 1931, 6

Carter, G. S. and Beadle, L. C. "Fauna of the Swamps of the Paraguayan Chaco in relation to its environment," *Journ Trop Soc Zool London* 1931, 37

Chaudhuri, B. L. "Fauna of the Chilka Lake. Fish, Pts 1-3," *Mem Ind Mus*, 1922, 5

Chilton, Chas. "Fauna of the Chilka Lake. Amphipoda," *ibid.*, 1921, 5  
 "Ibid. Some terrestrial Isopoda from the shore of the Lake," *ibid.*  
 "Ibid. Tanaidacea and Isopoda," *ibid.*, 1923

\_\_\_\_\_ "Zoological results of a tour in the Far East. Tanaidacea and Isopoda of Taleh Sap," *Rec Ind Mus* 1926, 28

Cowles, R. P. "Habits, reactions and associations of *Ocytoda arenaria*," *Corn Inst Wash* 103, *Pap Trop Lab.*, 1908, 2

Dakin, W. J. "The osmotic concentration of the blood of fishes taken from sea-water of naturally varying concentrations," *Biochem Journ*, 1908, 3

Dakin, W. J. "Variations in the osmotic concentrations of blood and coelomic fluids of aquatic animals caused by changes in the external medium," *Phil. Trans. Roy. Soc. London*, 1908.

"Notes on the biology of fish eggs and larvae," *Intern. Rev. Hydrobiol.* 1911, 4.

"Oxygen requirements of certain animals and its bearing on food supply," *Brit. Journ. Phys. Biol.* 1925, 2.

"The osmotic concentrations of the blood of *Callorhynchus milii* and *Epinephelus (Neoceratodus) fosteri*," *Proc. Zool. Soc. Lond.* 1931.

"The aquatic animal and its environment (Presidential Address)," *Proc. 1ann. Soc. New South Wales*, 1935, 60.

Dakin, W. J. and Edmonds, E. "The Regulation of the salt contents of aquatic animals and the problem of permeability of the bounding membranes of aquatic invertebrates," *Int. Journ. Exper. Biol.*, 1931, 8. Quoted by Dakin, 1935.

Duerden, J. E. "The Role of mucus in corals," *Quart. Journ. Mar. Sci.*, 1906, 49.

Duval 1925 Quoted by Dakin, 1935.

Eggert, Brunn "Beitrag zur Kenntnis der Biologie und Systematik der Periophthalmine," *Zool. Jahrb. Abt. Syst.* 1935.

Ekmann, Sven "Geographie des Meeres," Leipzig, 1935.

Ellis, A. E. "Habits of the Hydrobiidae in the Adur estuary," *Proc. Mal. Soc. Lond.*, 1932.

Ellis, W. G. "Calcium and resistance of *Nereis* to brackish-water," *Nature*, 1933, 132, 748.

Fauvel, P. "Annelida Polychaeta of the Indian Museum," *Mem. Ind. Mus.*, 1932, 12.

Fewkes, J. W. "Development of certain worm larvae," *Bull. Mus. Comp. Zool. Harvard*, 1883-85, 11.

Fischer, P. E. "Nouvelles observations sur l'ordre euryhalinum 'des spèces littorales,'" *Bull. Inst. Océan. Monaco*, 1933.

Flattely, F. W. and Walton, C. L. *The Biology of the Seashore* N.Y., 1922.

Florentin, M. R. "Faimé des mares salées de Lorraine," *Ann. Sci. Nat. Zool.* (8), 1899, 10.

Garrey, W. C. "The osmotic pressure of sea-water and the blood of marine animals," *Biol. Bull.*, 1933.

Gessner, F. 1932 and 1933 Quoted by Nicol (1935) and Remane (1934).

Gravely, F. H. "A note on the marine invertebrate fauna of Chittagong, Orissa," *Rec. Ind. Mus.*, 1919, 16.

Gueylard, F. 1924 Quoted by Dakin, 1935.

Gurney, R. "Notes on *Leander longirostris* and other British Prawns," *Proc. Zool. Soc. London*, 1923.

Hall, F. G. "Vital limit of excitation of certain marine animals," *Biol. Bull.* 1922, 16.

Harms, J. W. "Die realisation von genen und die consecutive Adaptation, Pt 1," *Zeit Wiss Zool.*, Bd. 133, 1929

\_\_\_\_\_, Pt 2," *Akad Verlagsbuch Heine*, 1934

\_\_\_\_\_, Pts 3 and 4," *Zeit Wiss Zool.*, 1932 and 1935, Bd. 140 and 146

\_\_\_\_\_, "Die Plastizität der Tiere," *Rev. Suiss Zool.*, 1935, Bd. 42

Harvey, H. W. "Biological Physics and Chemistry of Sea Water," *Camb Comp Physiol Ser.*, 1928

Henderson, J. R. "A contribution to Indian Carcinology," *Trans. Linn. Soc. London Zool.*, 1894, 5

\_\_\_\_\_, "Hermit crabs from the Chilka Lake," *Rec. Ind. Mus.*, 1915, 11

Herbant 1897 Quoted by Needham, 1930

Hesse, R. "Türgéographie auf Ökologischer Grundlage," Jena, 1924

Hora, S. L. "Fauna of the Chilka Lake Fish, Pt 5," *Mem. Ind. Mus.*, 1922, 5

\_\_\_\_\_, "Animals in brackish-water at Uttarbhag, Lower Bengal," *Curr. Sci.*, 1933, 1

\_\_\_\_\_, "A Note on the Bionomics of two estuarine crabs," *Proc. Zool. Soc. London*, 1933

Hornell, J. "A revision of the Indian species of *Mereatrix*," *Rec. Ind. Mus.*, 1917, 13

\_\_\_\_\_, "The Common Molluscs of South India," *Medras Fisheries Bull.*, 1922, 14

Huntsman, A. G. "The vertical distribution of certain intertidal animals," *Trans. Roy. Soc. Canada*, 1918, 2

Jobert 1876 Quoted by Carter, 1931

Jones, S. "Observations on the breeding habits and development of certain brackish-water fishes of Adyar, Madras," *Proc. Ind. Acad. Sci. (B)*, 1937, 5

Kemp, Stanley "Fauna of the Chilka Lake Crustacea Decapoda," *Mem. Ind. Mus.*, 1915, 5

\_\_\_\_\_, "Ibid., Stomatopoda," *ibid.*, 1915

\_\_\_\_\_, "Ibid., Cumaceae," *ibid.*, 1915

\_\_\_\_\_, "Notes on the fauna of the Matlah River in the Gangetic Delta," *Rec. Ind. Mus.*, 1917, 13

\_\_\_\_\_, "Notes on the Crustacea Decapoda in the Indian Museum, Pt 10 The Hymenosomatidae," *ibid.*

\_\_\_\_\_, "Ibid., Pt 11 *Leander styliferus* M. Edw and related forms," *ibid.*

\_\_\_\_\_, "Ibid., Pt 12 Scopumerinae," *ibid.*, 1919, 16

\_\_\_\_\_, "Ibid., Pt 15 Pontoniinae," *ibid.*, 1922, 24

Keys, A. "The mechanism of adaptation to varying salinity in the common eel and the general problem of osmotic regulation in fishes," *Proc. Roy. Soc. Ser. B.*, 1933, 112

Krishnan, G  
"The development of *Diplopia variabilis* Southern," *Zeit Wiss Zool*, 1936, Bd 147

Kunkel, B. W  
"The selective action of certain environmental conditions on the hermit crab, *Clibanarius tricolor* Giles," *Carn Inst Wash*, 1934, Publ 415

Lambert, F. J  
"Animal life in the marsh ditches of the Thames Estuary," *Proc Zool Soc London*, 1930

Lenz, J  
1933 (in *Verb Intern Verein Immunolog*), quoted by Remane, 1934

Lloyd, R. E., and Annandale, N  
"On the Hydrozoan *Campanulina ceylonensis* (Browne)," *Rec Ind Mus* 1915, 12

Lowenstein, O  
"The respiratory rate of *Gammareus cherreensis* in relation with the changes in salinity," *Journ Exper Biol*, 1935, 12

Lull, R. S  
"Organic Evolution, N.Y., 1917

Lundbeck, J  
Quoted by Nicol, 1935

Macallum, A. B  
"The Paleochemistry of body fluids and tissues," *Physiol Rev*, 1926, 6

Mergneria, R  
"The osmotic changes in some marine animals," *Proc Roy Soc*, (B), 1931, 107

Martens, P. von  
"On the occurrence of Marine Animal forms in freshwater," *Ann Nat Hist London* 1858, 1, 3rd Ser

Mayer, A. G  
"Effect of temperature on tropical marine animals," *Carn Inst Wash*, 1933, *Pop Tort Lab*, 1914, 6

Muller, F. R  
1863, quoted by Carter, 1931

Needham, J  
"Penetration of marine organisms into freshwater," *Biol Zent*, 1930, Bd 50

Nicol, E. A. T  
"Ecology of a salt marsh," *Journ Mar Biol Assoc*, 1935, 20

Oldham, J  
"The brackish-water lochs of North Uist," *Proc Roy Soc Edin*, 1936, 56

Ortmann, A. E.  
"Geology of India, 1893

Orton, J. H  
"H. G. Bronn's Klassen und Ordnungen der Arthropoden," *Crustacea*, 1901, Bd 5

Pankkar, N. Kesava  
"Sea temperature, breeding and distribution in marine animals," *Journ Mar Biol Assoc*, 1920, 12

\_\_\_\_\_  
"The structure, bionomics and systematic position of two new brackish-water Actinaria from Madras," *Proc Zool Soc London*, 1936, Pt 1

\_\_\_\_\_  
"A study of the Actinian *Phycocatetes gangeticus* Annandale, with an account of the post-larval development and the occurrence of neoteny in the Anemone," *Zool Jahrb Abt Anat u Oest*, 1937, Bd 62

\_\_\_\_\_  
"The morphology and systematic relationships of a new Boloceroidarian from brackish-water, together with an account of its asexual reproduction," *Proc Ind Acad. Sci*, Ser B, 1937, 5, No. 2



Remane, A "Die brackwasser fauna," *Verh Deutsch Zool Ges. Zool Ans. Supplement* 1934

Ritchie, J "The Hydroids of the Indian Museum II A new and remarkable brackish-water Hydroid, *Annulella gemmata* *Rec Ind Mus* 1915, 11

Robson, G C "The animal life of estuaries," *J Quiclett Micros Club*, London, 1925, 15, quoted by Pearse, 1936, Nicol, 1935, etc

Saunders, J T "The Hydrogen Ion concentration of natural waters I. The relation of pH to carbon dioxide," *Journ Exper Biol* 1926, 4

Saville, L P "The effects of salinity changes on body-weight and survival of *Nereis virens*, *Biol Bull* (2), 1935, 69

Schleiper, C "Über die Einwirkung niedriger Salzkonzentrationen auf marine organismen *Zeit Tiers Physiol* 1929, Bd 9

— "Die osmoregulation der salzwasserkrabben," *Verh Deutsch Zool Ges* 1929a

— "Die osmoregulation wasserlebender Tiere" *Biol Rev Cambridge* 1930, 5

— "Neuere Ergebnisse und Probleme aus dem gebiet der osmoregulation wasserlebender Tiere," *Biol Rev Cambridge*, 1935, 10

Schwabe, E "Über die osmoregulation verschiedener krebse," *Zeit Tiers Physiol* 1933 Bd 19

Semper, Karl "Inimal Life Int Sci Ser. London, 1885

Sergestale 1914 Quoted by Remane, 1914

Sewell, R B S "Notes on plankton from the Chilka Lake *Rec Ind Mus* 1913, 9

— "A preliminary account of some new species of Copepoda," *ibid*, 1919, 16

— "Fauna of the Chilka Lake Crustacea Copepoda," *Mem Ind Mus* 1923, 5

— "Hydrographical and oceanographical Researches in Indian Waters," *Mem Asiat Soc Bengal*, 1929, 9

— "Fauna of the Salt Lakes, Calcutta," *Rec Ind Mus*, 1934, 35

Sewell, R B S and Annandale, N "Fauna of the Chilka Lake The Hydrography and Invertebrate fauna of the Rambha Bay in an abnormal year," *Mem Ind Mus*, 1921, 5

Sick, F "Die Fauna der Meeresstrandtümpele des Bottsandes (Keiler Bucht)," *Arch Naturg* 1933

Smith, H W "Water regulation and its evolution in fishes," *Quart Rev Biol*, 1932, 7

Sollas, W J "On the origin of freshwater faunas," *Trans Roy Dublin Soc* Ser 2, 1883, 3

Southern, R *The Origin of Earth* London, 1905

— "Fauna of the Chilka Lake Polychaeta," *Mem Ind Mus*, 1921, 5

Stammer	1928 Quoted by Remane, 1934, and others
Stephenson, J	"Fauna of the Chilka Lake Oligochaeta," <i>Mem Ind Mus</i> , 1915 and 1917, 5
Stoliczka, F	"On <i>Sagittaria schilleriana</i> n. sp. and <i>Membranipora bengalensis</i> n. sp." <i>Journ Anat Soc Bengal</i> , 1869, 38
Subramaniam, M. K	"Oogenesis of <i>Clebunarites olivaceous</i> Henderson, with special reference to a seasonal variation in the cytoplasmic inclusions," <i>Journ Roy Micros Soc</i> , 1935, 55
Sumner, F. B	"The Physiological effects upon fishes of changes in density and salinity of water," <i>Bull U.S. Bureau of Fisheries</i> 1906, 25
Tattersal, W. M	"Fauna of the Chilka Lake. The Mysidacea of the Lake, with description of a species from the Coast of Orissa," <i>Mem Ind Mus</i> 1915, 5
-----	"Zoological results of a tour in the Far East. Mysidacea, Tanagridae and Isopoda," <i>Mem Asiat Soc Bengal</i> , 1921, 8
-----	"Indian Mysidacea," <i>Rec Ind Mus</i> 1922, 24
Verhoef, Karl	1919 Quoted by Carter, 1931
Vernon, H. M	"The relation between animal and vegetable life," <i>Brit Zool Stat Neapel</i> 1889, 13
Winckworth, R	"Mollusca from Pulicat Lake," <i>Proc Mal Soc London</i> , 1931, 19
Winterstein, H	1921 Quoted by Carter, 1931 Carter and Beadle, 1931
Wundsch	1933 ( <i>Verh Int Ver Limn</i> ) Quoted by Remane, 1934
Zimmer, C	"Crustacea," Kukenthal and Krumbach's <i>Handbuch der Zoologie</i> , 1926-27, Bd 3, Pt 1 Quoted by Carter, 1931

## EXPLANATION OF FIGURES

## PLATE XVIII.

Photographs of the Adyar backwater and the river, taken during March 1935. The first is a portion of the backwater between the Adyar cemetery and the sand bank, showing the marginal and middle zones. The second photograph shows the Adyar river after the Elphinstone Bridge. Much of the river-bed is exposed owing to drought.

## PLATE XIX

- (1) Northern bank of the river above the Elphinstone Bridge
- (2) Inner reaches of the river showing pools of fresh or slightly brackish-water

## PLATE XX.

- (1) The river after Elphinstone Bridge to show the deep channel on the southern side
- (2) Part of the Adyar backwater near the small bridge close to the cemetery

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and R. Gopala Aiyar



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and R. Gopala Aiyar*

*Proc. Ind. Acad. Sci., B, vol. VI, Pt. XX*



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2



## TEXT-FIG. 1

Map of the Adyar river and backwater to illustrate the different regions surveyed in this study

## FIG. 2

Imaginary section through a mud-flat on the side of the backwater to show the approximate zonation of animals, especially the burrowing species. Numbers 1-26 represent the following species --

1	<i>Pontodrilus bermudensis</i>	14	<i>Cibularia oblonga</i>
2	<i>Oxypoda cordimana</i>	15	<i>Potamides cingulatus</i>
3	<i>Oxypoda macrocera</i>	16	<i>Phytocatoptris saundersi</i>
4	<i>Sesarma tetragonum</i>	17	<i>Pygmaeomysa oryzensis</i>
5	<i>Larvula littoralis</i>	18	<i>Cibularia radiatus</i>
6	<i>Metaplaes disticta</i>	19	<i>Syllis serrata</i>
7	<i>Uca annulipes</i>	20	<i>Stephensonactis ornata</i>
8	Zone of <i>Lyngbya medusa</i>	21	<i>Pilocerat escul</i>
9	<i>Lumbriconereus</i> sp.	22	<i>Menidia casta</i>
10	<i>Phytocatoptris gangetica</i>	23	<i>Niphargus pelagicus</i>
11	<i>Sesarma quadratum</i>	24	<i>Neptunus sanguinolentus</i>
12	<i>Boleophthalmus boddarti</i>	25	<i>Marphysa sanguinea</i>
13	<i>Periophthalmodon pearsoni</i> and <i>P. kuhlii</i>	26	<i>Diopatra torquata</i>

1725-37—Printed at The Bangalore Press, Bangalore City, by G. Srinivas Rao, Superintendent,  
and Published by The Indian Academy of Sciences, Bangalore.

## THE MYXOPHYCEAE OF THE UNITED PROVINCES, INDIA.—III.\*

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Received June 25, 1937.

[Communicated by Prof. Y. Bharadwaja, M.Sc., Ph.D. (Lond.), F.I.S.]

The present communication is the third<sup>1,2</sup> of the series, 'The Myxophyceae of the United Provinces, India'. It deals with algae (hitherto unrecorded with the exception of *Anabaena ambigua* Rao) collected mainly from Benares and its environs from 1934 onwards. A few forms collected from Chakta, Chunar, Saranath, Ramnagar and Mussoorie have also been included. In all there are one hundred and five forms, representing twenty-five genera, and out of these, two species, sixteen varieties and twenty-seven forms are new. The habitat of these plants is varied, thirty-five have been collected on soil, seven from moist bricks, stones and rocks, eleven on walls and other elevated places, four on the bark of trees, five from the stagnant water of crop-fields and the rest from tanks, ponds, puddles and other aquatic situations.

The major part of the Benares Myxophyceae are sub-aerial in habitat. After the rains, one comes across very frequently on semi-clayey soil, during July and August, a very thick, semi-transparent and ash-brown mucilaginous scum extending over large areas and making the ground slimy. This is mostly comprised of *Aphanothecia pallida*. It may be intermingled with *Chroococcus montanus* forma and var. *hyalina*, *Aphanocapsa Grevillei*, *Microcoleus chthonoplastes* and others. On clayey soil, however, the stratum formed is deep blue-green and slimy, but closely adhering to the surface. This is commonly comprised of *Glaucapsa stegophila* var. *crassa*, *Cylindrospermum muscicola*, *Microcoleus chthonoplastes* and *M. socialis*. During July and August, *Cylindrospermum muscicola* is noticed very frequently to form by itself blue-green strata on moist soil, and as its spores ripen the strata turn blackish-brown. In the month of September, conspicuous blue-green erect tufts of a form of *Symploca muralis* appear on shady soil. Frequently

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<sup>1</sup> Bharadwaja, Yajnavalkya, "The Myxophyceae of the United Provinces, India—I," *Proc. Ind. Acad. Sci.*, 1935, 2, No. 1, Sec. B.

<sup>2</sup> Bhashyakarla Rao, C., "The Myxophyceae of the United Provinces, India—II," *Proc. Ind. Acad. Sci.*, 1936, 3, No. 2, Sec. B.

during August and September, circular, woolly, shiny and brown patches are seen scattered on the surface of lawns. These expand and finally coalesce to produce a brownish green stratum which for the most part consists of *Microcoleus tenera*, *Scytonema javanicum*, with which, however, *Microcoleus chthonoplastes* and a form of *Anabaena variabilis* may be intermingled. *Oscillatoria terebriformis* is the most common species of *Oscillatoria* in Benares and it occurs not only on all exposed muddy places, but also in stagnant ponds, puddles and dirty drains. *Oscillatoria formosa* is another form commonly met with in Benares, especially during rains, forming a crimson-green layer on the soil of shallow puddles and ditches. Among the other common blue-green algae, occurring on moist soil, mention may be made of *Oscillatoria sancti*, *Phormidium Retzii* and *Lyngbya ceylanica*. On places which are constantly wet, because of their being near drains or other water sources, *Phormidium subtruncatum* is very common. This alga has also been found growing in very delicate blue-green films spread on the sides of the aspirator bottles kept out of use in the laboratory. *Scytonema ocellatum*, *Lyngbya truncula*, *L. aestuaria* var. *arbuscula* and *L. arboricola* are seen after rains in the form of expanded bluish-green woolly layers on the bark of *Mangifera indica*, *Eugenia Jambolana*, *Bassia latifolia* and *Famadihns indica*. As the season becomes drier, the strata formed by these algae become thin and papery, and may ultimately peel off the trunk of the tree.

In Benares, there is a comparative paucity of the real aquatic forms. This is due to the fact that there are no large and permanent sheets of stagnant water. The characteristic aquatic habitats of these algae are a large number of pools, puddles and ditches formed by the stagnant rain-water in low-lying areas either in waste-places or amidst crop-fields or on the sides of the River Ganges, which flows by the side of the town. Some of the common planktonic forms are *Microcystis flos-aquae*, *Chroococcus turgidus*, *C. minutus*, *Merismopedia tenüssima* and *Spirulina platensis* var. *crassa*. *Oscillatoria princeps* is rather frequent near the edges of pools, puddles and ditches as black masses of densely crowded hair-like filaments. Species of *Anabaena*, such as *A. aphanizomenoides* var. *ellipsospora*, *A. Iyengari* var. *tenuis*, and *A. unispora* var. *crassa*, occur in a large number of rain-water pools. They are sterile during July and August, but form spores in September and October. *Glaetrichia natans* occurs as large blue-green globules attached to aquatic angiosperms. *Glaetrichia intermedia* var. *kanwense* is abundant in several rain-water pools, either in a free state or adhering to *Chara*, and other aquatic plants. *Aulosira fertilissima* var. *tenuis* and *Phormidium mucosum* var. *arvense* are found in abundance during rains in stagnant water of crop-fields. *Lyngbya conervoides* grows in thick

fibrous and deep blue-green masses in the shade of closely spreading leaves of *Nelumbium* in the ornamental water reservoirs

The Benares Myxophyceæ grow luxuriantly during the rainy months of July, August and September and to some extent in October and November. But they are quite scarce during May and June, when it is very hot and dry. Some of the forms available during these hot months are *Microcystis flos-aqua*, *Merismopedia tenissima* and *Spirulina major*, occurring in tanks with constant water supply, and also *Oscillatoria terebriformis* and *Phormidium subtruncatum* inhabiting places near water.

SYSTEMATIC ENUMERATION OF THE SPECIES OBSERVED

I CHROOCOCCALES

*Chroococcaceæ*

Genus *Microcystis* Kützing

1 *Microcystis aeruginosa* Kützing Geitler, in Rabenhorst's *Kryptogamenflora von Europa*, Band XIV, Cyanophyceæ, 1930-32, p 136, Fig 59 d, Frémy "Les Mixophyceæ de Madagascar," *Annales de Cryptogamie exotique*, t. III, fasc. IV, 1930, Pl IV, Fig 5, Crow, "The taxonomy and variation of the genus *Microcystis* in Ceylon," *New Phytologist*, 1923, Vol 22, No 2, Pl I, Fig a, Tilden, *Minnesota Algae*, Vol I, 1910, Pl II, Figs 21 and 22

Var *elongata* var nov (Fig 1, A-C)

Colonies spherical or elongated, solid or broken through to form a net, broken parts of the colony looking like filaments, sheath indistinct, stained violet with methylene blue, cells pale blue-green, more or less spherical, with gas-vacuoles

Lat cell, 2-2.8  $\mu$

Habitat —In an ornamental water reservoir of a private garden, along with *Oscillatoria Annae*, *Lynghya limnetica* and *Oedogonium* sp

The form agrees with the type in having spherical or elongated colonies, which are solid or broken through with an indistinct sheath, and spherical cells with gas-vacuoles, but differs in the cells being smaller and the broken portions of the colonies looking like filaments

Genus *Aphanocapsa* Naegele

2 *Aphanocapsa Koordersi* Ström Geitler, *op. cit.* 1930-32, p 155, Fig 68

Colonies without definite shape. Cells laxly crowded, circular to sub-globose with pale blue-green homogeneous contents

Lat cell, 2-2.3-2  $\mu$

Habitat:—In the fountain tank, Women's Hostel, Benares Hindu University, along with *Synechocystis aquatilis* and *Oedogonium* sp (Miss P. R. Parukutty Amma)

3 *Aphanocapsa Grevillei* (Hass) Rabenh Geitler, *op. cit.*, 1930-32, p 158, Fig 71, Frémy, "Les Myxophycées de l'Afrique équatoriale française," *Arch d Bot*, iii (1929), Mem 2, 1930, p 23, Fig 25, Tilden, *op. cit.*, 1910, Pl II, Fig 7, West, *Algae*, I, 1916, p 3, Fig 2, A

Lat cell, 3.2-4.8  $\mu$

Habitat.—On the sides of the steps of a tank along with other algae

The plant-mass occurs as small yellowish patches distributed irregularly

4 *Aphanocapsa pulchra* (Kütz) Rabenh Geitler, *op. cit.*, 1930-32, p 156, Fig 69g, Frémy, *op. cit.*, 1930, p 23, Fig 22

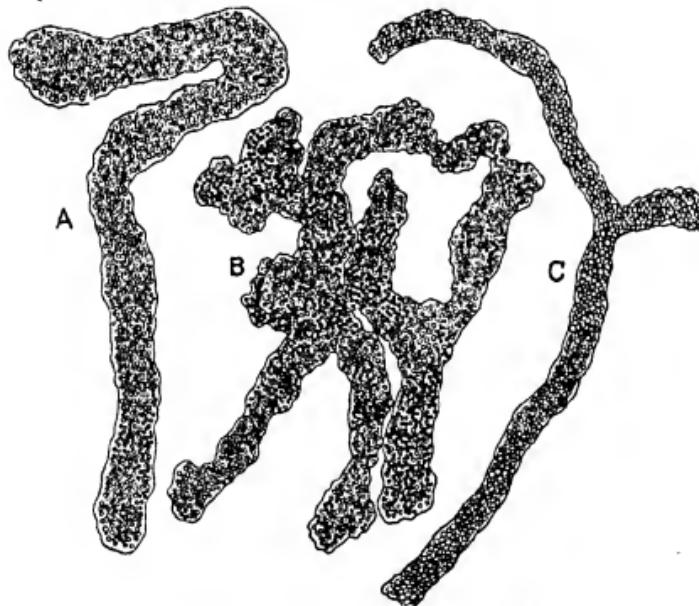


FIG 1. A-C—*Microcystis aeruginosa* Kütz, var. *elongata* var. nov.

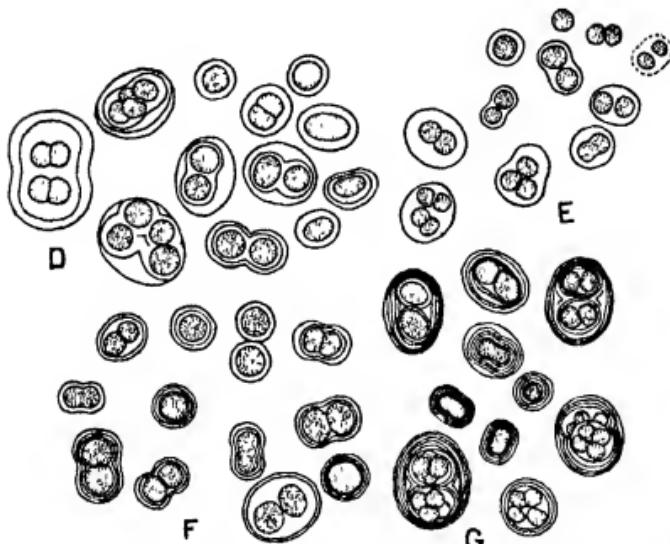


FIG. 1 *D—Glaoocapsa stegophila* (Jitzig) Rabenh var. *erecta* var. nov., *E—Chroococcus minimus* (Keissl) Lemm., *F & G—Chroococcus montanus* Hansg. forma and var. *hyalina* var. nov. respectively

*A-C & G*  $\times 425$ , *D & F*  $\times 875$ , *E*  $\times 1,475$

Lat cell, 3-4.5  $\mu$

Habitat —Planktonic in a tank, Cantonment area

Genus *Aphanothecæ* Nageli

*5 Aphanothecæ bullosa* (Menegh) Rabenh Frémv. op. cit., 1930, p. 33,  
Fig. 33

Lat cell, 4.9-5.4  $\mu$ , long cell, 6.6-13.2  $\mu$

Habitat —Free-floating in a rain-water pool along with *Lyngbya spiralis* Geitler

The stratum is a deep blue-green fragile mass

Genus *Glaoocapsa* Kützing

*6 Glaoocapsa punctata* Nag Frémv. "Les Myxophyceæ de Madagascar,"  
*Annales de Cryptogamie exotique*, t. iii, fasc. IV, 1930, Pl. V, Fig. 19,  
Geitler, op. cit., 1930-32, p. 189

Lat cell, 1.6-3  $\mu$ , long cell, 3.2  $\mu$ , lat cell cum vag, 3.5-4  $\mu$ , long cell cum vag, 4.2-6  $\mu$

Habitat —In a cemented water-reservoir of a private garden, along with *Nostoc spongiforme* var *tenuis*, *Tolyphothrix lanata* forma, *T. distorta* var *samoensis* and *Oedogonium* sp

*T. Glaucoptila stegophila* (Itzigs) Rabenh Geitler, *op cit*, 1930-32, p 197, Fig 91b, Tilden, *op cit*, 1910, Pl I, Fig 24

Var *crassa* var nov (Fig 1, D)

Thallus soft, yellowish-brown, cells spherical, sub-spherical or elongated, commonly single or in colonies of 2-4, sheath golden yellow, sometimes striated

Lat cell, 4.6-4  $\mu$ , long cell, 6.4-9.6  $\mu$ , lat colon cum vag, 8.15  $\mu$ , long colon cum vag 9.6-19.2  $\mu$ , crass vag, upto 3.2  $\mu$

Habitat —On moist soil, singly or along with *Microcoleus chthonoplastes*, *M. sociatus*, *Cylindrospermum muscicola* and others

The variety differs from the type in having bigger cells and colonies

#### Genus *Chroococcus* Naegeli

8 *Chroococcus turgidus* (Kutz) Nag Geitler, *op cit*, 1930-32, p 228, Fig 109b, Frémv, *op cit*, 1930, p 41, Fig 40, Tilden, *op cit*, 1910, Pl I, Fig 3, West, *op cit*, 1916, p 41, Fig 25b

Lat cell, 11.5-16.5  $\mu$ , long cell, 16.5  $\mu$ , lat cell cum vag, 11-19.2  $\mu$ , long cell cum vag 19.5  $\mu$ , lat colon cum vag, 16.5-21.2  $\mu$ , long colon cum vag, 21.4-23.1  $\mu$

Habitat —Planktonic in the tank of the Benares Electric Light and Power Supply Co, Ltd, along with *Chroococcus minutus*, *Merismopedia tenuissima* and *Spirulina major*

The cells are yellowish-green, mostly 2-4 in each colony

9 *Chroococcus minutus* (Kutz) Nag Geitler, *op cit*, 1930-32, p 234, Fig 113c, Frémv, *op cit*, 1930, p 41, Fig 42

Lat cell, 5.6-6  $\mu$ ; long cell, 3.3-6  $\mu$ , lat cell cum vag, 6.0-10  $\mu$ , long cell cum vag, 6.3-13  $\mu$ , lat colon cum vag, 6.6-10  $\mu$ , long colon cum vag, 6.6-15.5  $\mu$

Habitat —Planktonic in the tank of the Benares Electric Light and Power Supply Co, Ltd, along with *Chroococcus turgidus*, *Merismopedia tenuissima* and *Spirulina major*

Cells are mostly in colonies of two and rarely four.

10 *Chroococcus minimus* (Keissl) Lemm. Lemmernann, *Kryptogamenflora d. Mark Brandenburg*, iii, Algen I, 1910, p. 59, Geitler, *op. cit.*, 1930-32, p. 233 (Fig. 1, E)

Lat cell, 1.6-2.5(-3)  $\mu$ , long cell, 2.3-2  $\mu$ , lat cell cum vag, 3-4.5  $\mu$ , long cell cum vag, 3.2-5  $\mu$ , lat colon cum vag, 4.8-6.4  $\mu$ , long colon cum vag, 4.8-8.4 (-9.6)  $\mu$

Habitat —In an ornamental tank of a private garden, on the sides of a cemented water reservoir, by the side of the road leading to Allahabad

The sheath in the form collected from the ornamental tank is sometimes dissolved or very indistinct

11 *Chroococcus varius* A. Braun Geitler, *op. cit.*, 1930-32, p. 235, Fig. 114 a

Lat cell, 2.4  $\mu$ , long cell, 2.5-4  $\mu$ , lat cell cum vag, 3.5  $\mu$ , long cell cum vag, 3.5-5  $\mu$ , lat colon cum vag, 4.2-8.4  $\mu$ , long colon cum vag, 4.5-11.6  $\mu$

Habitat —On the cemented platform of the Physical Laboratory, College of Science, Benares Hindu University

12 *Chroococcus montanus* Haussg. Geitler, *op. cit.*, 1930-32, p. 236  
Forma (Fig. 1, F)

Lat cell, 5.6-6.4  $\mu$ , long cell, 4.8-8  $\mu$ , lat cell cum vag, 6.4-9.6 (-12.8)  $\mu$ , long cell cum vag, 8-11.2  $\mu$ , lat colon cum vag, 8.8-16  $\mu$ , long colon cum vag, 11.2-14.4  $\mu$

Habitat —On moist soil, along with *Microcoleus chthonoplastes* and others

The form differs from the type in the sheath being hyaline

Var. *hyalina* var. nov (Fig. 1, G)

Stratum thick, mucilaginous and blue-green. Cells spherical or sub-spherical or elongated, single or 2-4 or occasionally upto 8 in spherical or ellipsoidal colonies. Colonies separate. Sheath thick, hyaline and lamellated

Lat cell, 4.8-7.5  $\mu$ , long cell, 8-16.5  $\mu$ , lat cell cum vag, 13.2-23.1  $\mu$ , long cell cum vag, 13.2-30  $\mu$ , lat colon cum vag, 19.8-26  $\mu$ , long colon cum vag, 19-36  $\mu$

Habitat —On moist soil along with other algae

The variety agrees with the type in the mucilaginous stratum, the presence of 2-4 or occasionally more cells in each colony, and the sheath enveloping the colonies being stratified, but it differs from the same in the stratum being blue-green and the colonies, which are on the average bigger, possessing a hyaline and thick sheath

13. *Chroococcus pallidus* Nüg. Geitler, *op. cit.*, 1930-32, p. 238, Fig 116 b, Frémy, *op. cit.*, 1930, p. 41, Fig. 48

Lat cell, 4.8-6.6  $\mu$ , long cell, 4.8-7  $\mu$ , lat cell cum vag, 6.6-10  $\mu$ , long cell cum vag, 6.6-11.6  $\mu$ , lat colon cum vag, 8.4-10.5  $\mu$ , long colon cum vag, 10.5-13.2  $\mu$

Habitat —On wet soil along with *Cylindrospermum muscicola*, *Microcoleus chthonoplastes* and others

The cell-contents are deep blue-green

Genus *Merismopedia* Meyen

14. *Merismopedia minima* G. Beck Geitler, *op. cit.*, 1930-32, p. 263, Bhashyakarla Rao, "The Cyanophyceae of the United Provinces, India—II," *Proceedings of the Indian Academy of Sciences*, 1936, Vol III, Sec B, p. 166, Fig. 1, B

Forma

Lat cell, 0.2-0.4  $\mu$

Habitat —Planktonic in the fountain tank, College of Science, Benares Hindu University

The form differs from the type in possessing smaller cells and colonies consisting of upto 320 cells

15. *Merismopedia tenuissima* Lemm. Geitler, *op. cit.*, 1930-32, p. 264, Fig 129 b, Frémy, "Les Cyanophycées des Côtes d'Europe," *Mémoires de la Société Nationale des Sciences Naturelles et Mathématiques de Cherbourg, tome XL*, 1934, Pl. I, Fig. 1, Geitler, in Pascher's *Süßwasserflora Deutschlands, Österreichs und der Schweiz*, Heft 12, Cyanophyceae, 1925, p. 107, Fig. 123 a

Lat cell, 1.6-2  $\mu$

Habitat —Planktonic in the tank of the Benares Electric Light and Power Supply Co., Ltd., along with *Chroococcus turgidus*, *C. minutus* and *Spirulina major*.

The colonies are generally of 16-48 cells

Genus *Synechocystis* Sauvageau

16. *Synechocystis aquatilis* Sauvageau Tilden, *op. cit.*, 1910, Pl. I, Fig. 10 (Fig. 2, A)

Lat cell, 5.6-6.5  $\mu$

Habitat —In the fountain tank, Women's Hostel, Benares Hindu University, along with *Aphanocapsa Koordersii* and *Oedogonium* sp (Miss P. R. Parukutty Amma), in a temple tank, Benares City

Genus *Dactylococcopsis* Hansg

17. *Dactylococcopsis raphidoides* Hansg Gentler, *op. cit.*, 1930-32, p. 281, Fig. 137, Printz, "Sub-aerial Alge from South Africa," *Norske Videnskabers selaskabs skrifter*, 1920, No. 1, Pl. 11, Figs. 328-36, Borge, *Beitrage zur algenflora von Schweden*, "Arkiv for Botanik", Band 18, No. 10, Pl. I, Fig. 1

*Forma.*

Lat cell, 1 6-2  $\mu$ , long cell, 20-35 (- 40)  $\mu$

Habitat —In a pond on the University grounds, along with *Calothrix* *marciana* var. *intermedia*, *Anabaena fertilissima* sp. nov., *A. Iyengari* var. *tenuis*, *Oscillatoria animalis*, *Lyngbya Digeuli* forma and sterile filaments of *Spirogyra* and *Oedogonium*

The form differs from the type in the cells being longer and usually falsiformed. It also differs from forma *falciformis* Printz in having much longer cells

II CHAMOSIPHONALES

*Chamosiphonaceæ*

Genus *Chamosiphon* A. Braun et Grunow

18. *Chamosiphon siderophilus* Starmach Gentler, *op. cit.*, 1930-32, p. 431, Fig. 251

Var. *glabra* var. nov (Fig. 2, B)

Sporangia distributed on the host either singly or in dense clusters, usually club-shaped, rarely cylindrical, bent or straight, pale blue-green with homogeneous contents. Sheath thin, hyaline and smooth. Exospore one

Lat sporang., 2.4-3.2 (- 4)  $\mu$ , long sporang., 4-14.4  $\mu$ , lat. sporang. cum vag., 3.2-3.5  $\mu$ , long sporang. cum vag., 4.8-17.6  $\mu$

Habitat —On *Lyngbya gracilis* in the reserve water tank of the Benares Water Works

The variety agrees with the type in the possession of club-shaped or cylindrical sporangia with one exospore and a foot formed by the enveloping sheath, but differs in the sporangia possessing much shorter range of dimensions and the sheath being always smooth, hyaline and without any calcareous impregnation

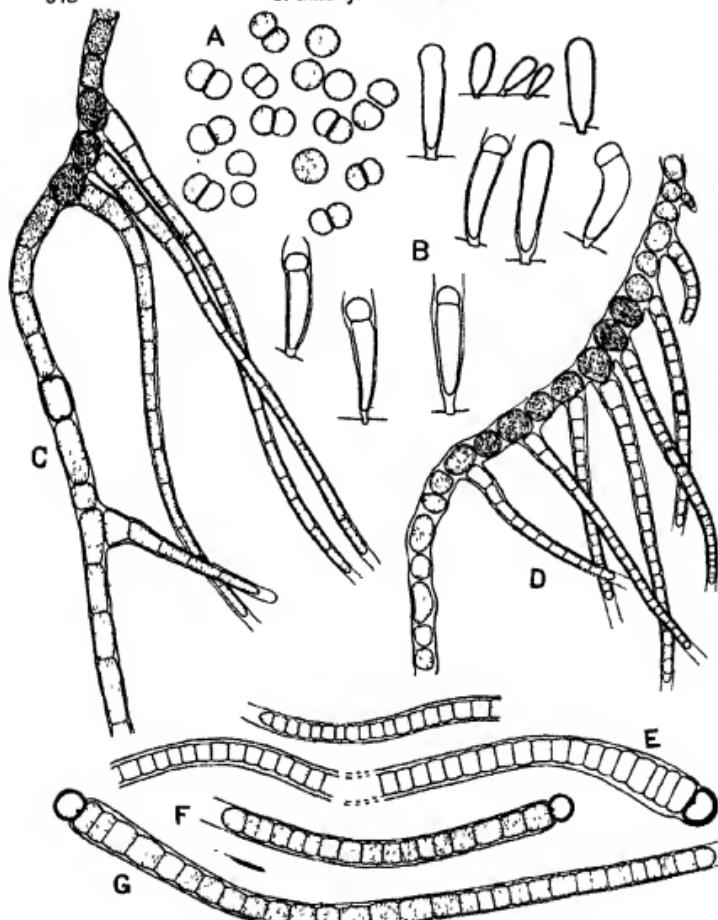


FIG 2 A—*Synechocystis aquatica* Sauvageau; B—*Chamopion eiderophilum* Star  
mach var. *globus* var. nov.; C & D—*Haplosporion Welwitschii* W. et G. S. West forma,  
E—Portions of the filament of *Calothrix marchica* (Lemm.) var. *crassa* var. nov.; F & G—  
*Calothrix marchica* Lemm. var. *intermedia* var. nov.

B  $\times$  1,475, all the rest  $\times$  875

III HORMOGONEALES

1 *Stigonemataceæ*

Genus *Hapalosiphon* Naegeli

19. *Hapalosiphon* *Weltwischii* W et G S West Götter, *op. cit.*, 1930-32, p 530, Fig 328 a

*Formæ* (Fig 2, C and D)

Plant-mass dark green, filaments closely entangled Sheath thin, firm and hyaline Cells spherical, quadratic or longer than broad Branches short and narrower than the main axes Heterocysts rare, long-cylindrical Spores more or less spherical or longer than broad

Main axis Lat cell, 3-7.5, rarely 9  $\mu$ , long cell, 4.5-15, rarely 16.8  $\mu$  Branches lat cell, 3-6  $\mu$ , the basal cell being 7.5  $\mu$ , long cell, 2.2-10.5  $\mu$  Lat het, 4.5-7  $\mu$ , long het, 7.5-10  $\mu$ , lat spor, 6-12  $\mu$ , long spor, 5.2-12.5  $\mu$ , cross vag, 0.2  $\mu$

Habitat —Closely adhering to the moist bricks at the edge of a rain-water pool

The form differs from the type in the terrestrial habitat and in the filaments being closely entangled

2 *Rivulariaceæ*

Genus *Calothrix* Agardh

20 *Calothrix marchica* Lemm Götter, *op. cit.*, 1930-32, p 607, Fig 382 a

Var *crassa* var nov (Fig 2, E)

Filaments in groups, irregularly bent and closely entangled Sheath thin, firm, yellowish or hyaline Trichomes with constrictions at septa, with tapering ends but without any hair-like prolongation, end-cell conical with a rounded apex, sometimes pointed Cells quadratic, as well as shorter or longer than broad Heterocysts single, basal, spherical or sub-spherical

Lat fil, 9.6-14.4  $\mu$ , long trich, upto 450  $\mu$ , lat trich, 8.4-12.8  $\mu$ , long cell, 2.3-2  $\mu$ , at top 4.8  $\mu$ , lat het, 8.2-12.5  $\mu$ , long het, upto 5  $\mu$

Habitat —On the plinth of the College building, near the outlet of a drain, along with *Calothrix linearis* forma

The variety agrees with the type in (1) the tapering trichomes without any hair-like prolongation, (2) the barrel-shaped cells, (3) the rounded or sometimes pointed end-cells and (4) the single spherical or sub-spherical basal heterocysts, but it differs in the broader, irregularly bent and closely

entangled filaments, arranged in groups, in the much broader trichomes and heterocysts, and in the yellow sheath

Var *intermedia* var nov (Fig. 2, F and G)

Filaments epiphytic on other algae, placed singly or in groups of two or three, with slight attenuation, without the formation of a terminal hair. Sheath thin, firm and hyaline. Cells quadratic, as well as shorter or longer than broad, with constrictions at the joints, end-cell rounded. Heterocysts single, basal and usually spherical.

Lat fil, 6-8  $\mu$ , long trich, upto 350  $\mu$ , lat trich, 5 8-7 8  $\mu$ , long cell, 3.2-8  $\mu$ , lat het, 5 6-7 8  $\mu$

Habitat.—In a pond on the University grounds, along with *Dactylococcopsis raphidioides* forma, *Anabaena fertilissima* sp nov, *A. Iyengari* var *tenuis*, *Oscillatoria animalis*, *Lyngbya Digeuti* forma and sterile filaments of *Spirangyra* and *Oedogonium*.

This form comes close to *Calothrix atricha* Frémy (Frémy, *op cit*, 1930, p 263, Fig 233) which Geitler (*op cit*, p 625) considers to be identical with *Calothrix marchica* Lemmermann. It resembles *Calothrix marchica* Lemm. in the free filaments, the absence of a hair-like prolongation at the ends of the trichomes, the constrictions at the joints, the thin and hyaline sheath, and in the single, spherical or sub-spherical basal heterocysts. But it differs from this form in the trichomes possessing only slight attenuation, in the rounded end-cell and in the bigger dimensions of all parts. The dimensions of this form are intermediate between those of the type and the var *crassa*.

21 *Calothrix linearis* Gardner Geitler, *op cit*, 1930-32, p 622, Fig 395 d

*Forma*

Lat fil, at base 10-12  $\mu$ , at top 4-5  $\mu$ , long trich, upto 450  $\mu$ ; lat trich, at base 6.6-9.5  $\mu$ , at top 3.3-3  $\mu$ , long cell, 2.5-5  $\mu$ , lat het, 6 6-8  $\mu$ , long het, 5-7  $\mu$

Habitat.—On the plinth of the College building, near the outlet of a drain, along with *Calothrix marchica* var *crassa*.

The form differs from the type in the trichomes being broader at the base and the cells in the terminal portions of the trichomes being quadratic or shorter than broad.

22 *Calothrix brevissima* G S West "Report on the Fresh-water Algae, including Phytoplankton of the Third Tanganyika Expedition conducted by

Dr W. A. Cunningham, 1904–1905," *Journal of the Linnean Society Bot.*, 1907, 38, p 180, Pl 10, Fig 8

Lat fil, 4.8–6.6  $\mu$ , long fil, 40–85  $\mu$ , lat trich, 3.8–4.5  $\mu$ , long cell, 1.6–3.5  $\mu$ ; lat het, 3.8–4.8  $\mu$ , long het, 4.4–5  $\mu$

Habitat.—Epiphyte on *Hydrodictyon reticulatum* growing in a rain-water pool, along with *Spirogyra chuniae* forma

Genus *Glaetrichia* Agardh

23 *Glaetrichia Raciborskii* Wołoszynska Geitler, *op cit*, 1930–32, p 637, Fig 405 a and b, Geitler, *op cit*, 1925, p 233, Fig 281 b

Var *kashiense* var nov (Fig 3, A–E)

Thallus forming large irregularly-lobed blue-green masses, 2–10 cm thick. Filaments with a thick, stratified and hyaline sheath. Trichomes with constrictions at joints, ending in a long hair. Cells at the base of the trichome barrel-shaped, much shorter than broad or almost as long as broad, higher up cylindrical, in the hair long cylindrical. Heterocysts single, spherical to ellipsoidal. Spores long, ellipsoidal to cylindrical with a hyaline smooth outer wall.

Lat. trich, at base 8.4–10  $\mu$ , higher up 4.8–6.4  $\mu$ , at apex 2.4  $\mu$ , long. trich, 800–1,000  $\mu$ , long cell, at base 2.4  $\mu$ , later 3.3–10  $\mu$ , in the hair upto 15  $\mu$ , at apex upto 35  $\mu$ , lat het, 8–12.8 (–13.2)  $\mu$ ; long het, 8.4–15 (–16.5)  $\mu$ , lat spor, 11.5–16  $\mu$ , average 13.2  $\mu$ , long spor, 42–66  $\mu$

Habitat.—In a rain-water puddle amidst crop-fields

The variety agrees with the type in all respects except that the former has much bigger thallus, broader trichomes, hyaline sheath, bigger heterocysts, that may be ellipsoidal, and narrower spores of a wider range in length with a hyaline outer wall. This form is also comparable to var *Liszenfeldiana* (Wol) Geitler on account of the spherical or ellipsoidal heterocysts and cylindrical spores with a colourless outer wall, but here also it differs in possessing bigger heterocysts and narrower spores.

3 *Microchaetaceae*

Genus *Microchaete* Thuret

24 *Microchaete tenera* Thuret Frémy, *op cit*, 1930, p 281, Fig 248. Tilden, *op cit*, 1910, Pl X, Fig. 11

Lat fil, 5.7–1  $\mu$ , lat cell., 4–5  $\mu$ , long. cell., 4.8–8  $\mu$ ; lat. het., 4.5–6.4  $\mu$ , long. het, 6.4–10  $\mu$

Habitat.—On wet soil of the lawns, Benares Hindu University

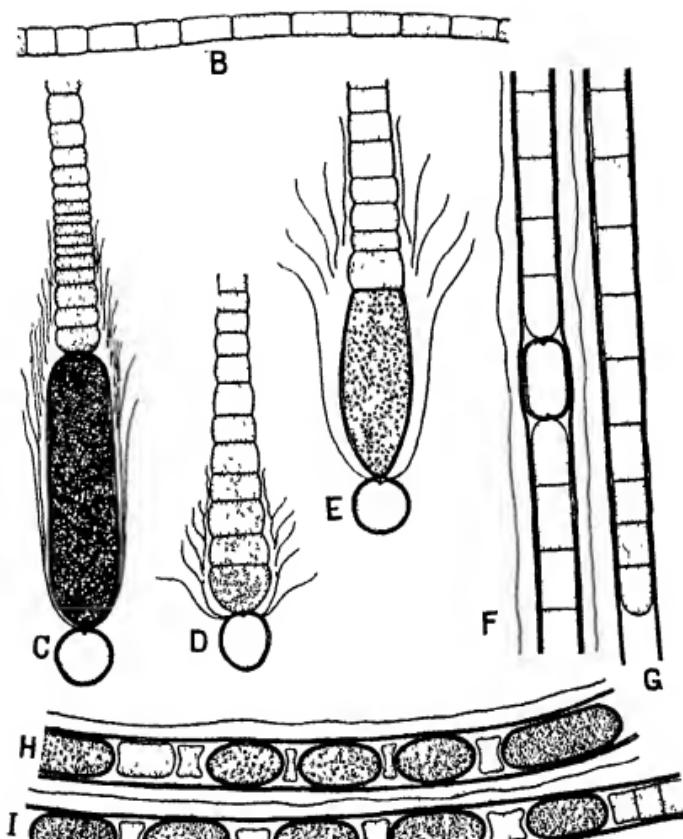


FIG 3. A-Terminal, B-middle and C-E-lower portions of the filament of *Gigastrchia Raciborskae Wołoszynska var. kashmirensis var. nov.*, F & G—vegetative and H & I—sporogenous portions of the filaments of *Aulosira fertilissima* Ghose var. *tenuis* var. nov.  
 A-E  $\times 875$ ; F-I  $\times 1,475$ .

Genus *Aulosira* Kirchner

25 *Aulosira Fritschii* Bhāradwāja Bharadwaja, "Contributions to our knowledge of the Myxophyceæ of India," *Annals of Botany*, 47, 185, 1933, p 123-131, Figs 3 and 4

Lat fil, 11.2-16.5  $\mu$ , crass vag, upto 3  $\mu$ , lat cell, 8.11-8.8  $\mu$ , long cell, 5.26  $\mu$ , lat het, 11.2-13.2  $\mu$ , long het, 11.8-10  $\mu$ , lat spor, 10-13.2  $\mu$ , long spor, 6.6-30  $\mu$

Habitat —In a stagnant rain-water puddle, by the side of the B N W Railway line, about 2 miles off Benares Cantonment Station, along with *Lyngbya conervoides*, *Oscillatoria princeps* and several others

26 *Aulosira fertilissima* Ghose Ghose, "A systematic and an ecological account of the blue-green algae of Lahore and Simla," *Jour Linn Soc Bot*, 46, 1923, Taf 31, Fig 9

Var *tenuis* var nov (Fig 3, F-J)

Plant-mass fibrous and greyish blue. Filaments free more or less straight and closely crowded together, with a thin, firm and brown sheath, that may sometimes be enveloped by distinct mucilage. Trichomes slightly tapering at the ends, rarely constricted at the septa, end-cell with a rounded apex. Cells cylindrical, sometimes quadratic. Heterocysts cylindrical, with rounded end-walls broader than the trichome and causing the filament to bulge. Spores in long chains, ellipsoidal to cylindrical, intercalated by moribund cells.

Lat fil, 5.6-6.4  $\mu$ , lat trich, 3.3-4.8  $\mu$ , long cell, 3.3-19.8  $\mu$ , lat het, 4.5-7  $\mu$ , long het, 8.2-19.8  $\mu$ , lat spor, 4.8-8.8  $\mu$ , long spor, 10.5-19.2  $\mu$ , crass vag, 0.3-0.8  $\mu$

Habitat —In the stagnant water of a paddy field

The variety agrees with the type in having (1) cylindrical or quadratic cells, (2) cylindrical heterocysts, (3) ellipsoidal spores in chains and (4) moribund cells in between the spores, but it differs from it in the fibrous stratum, the much narrower trichomes, the narrower heterocysts and the much smaller spores (that are also cylindrical) with a colourless outer wall

4 Scytonemataceæ

Genus *Tolyphothrix* Kützing

27 *Tolyphothrix nodosa* Bhāradwāja Bharadwaja, "The Taxonomy of *Scytonema* and *Tolyphothrix* including some new records and new species from India and Ceylon," *Revue Algalogique*, 1933, n 1-2, p 176, Fig 7c.

Lat fil, 5-8  $\mu$ , lat trich., 4.8-6.8  $\mu$ , at apices 4  $\mu$ , long cell., 3.2-12.8  $\mu$ ; lat het, 4.8-9.6  $\mu$ , long het, 6.4-16.8  $\mu$ , rarely upto 23  $\mu$ .

Habitat —In rain-water pools

28 *Tolypothrix lanata* Wartm Geitler, *op cit*, 1930-32, p 717, Fig 459 d

*Forma*

Lat fil, 8.2-11.2  $\mu$ , lat trich, 6.8-2  $\mu$ , long. cell, (5-) 6.4-10.4  $\mu$ , lat het, 6.6-8.2  $\mu$ , long het, 10.5-16.5  $\mu$

Habitat —In a cemented water reservoir in a private garden, along with *Nostoc spongiforme* var *tenuis*, *Tolypothrix distorta* var *samoensis*, *Glazocapsa punctata* and *Oedogonium* sp

The form differs from the type in having narrower trichomes and in the absence of heterocysts in chains

29 *Tolypothrix distorta* Kütz var *samoensis* Woile Bhāradwāja, *op cit*, 1933, p 176, Fig 7 b, Bhāradwāja, "False branching and Sheath-structure in Myxophycace, with special reference to the Scytonemataceae," *Archiv für Protistenkunde*, Band 81, Heft 2, 1933, Fig 3, E and Fig 4, G.

Lat fil, 14-19.8  $\mu$ , when old upto 22  $\mu$ , crass vag, 3.3-4  $\mu$ , when old and unhealthy upto 6  $\mu$ , lat trich, 12.2-18  $\mu$ , when old narrowed down to 10.2  $\mu$ , long cell, 3.2-12  $\mu$ , lat het, 11.8-13.2  $\mu$ , long het, 10-23  $\mu$

Habitat —In a cemented water reservoir in a private garden, along with *Nostoc spongiforme* var *tenuis*, *Tolypothrix lanata* forma, *Glazocapsa punctata* and *Oedogonium* sp

30. *Tolypothrix robusta* Gardner Geitler, *op cit*, 1930-32, p 715, Fig 457 b

*Forma*

Diam fil, 13-17  $\mu$ , when old upto 20  $\mu$ , crass vag, 2.4-5  $\mu$ , when old and unhealthy upto 6.6  $\mu$ , diam trich, 9-12  $\mu$ , when old and unhealthy narrowed down to 6.6  $\mu$ , at growing apices upto 15  $\mu$ ; long cell, 10-12  $\mu$ ; when old and unhealthy upto 30  $\mu$ , at growing apices upto 5  $\mu$ , lat het, 10-13.2  $\mu$ ; long het, 13-42  $\mu$ .

Habitat —Floating in a stagnant pond

The form differs from the type in the presence of narrower filaments and trichomes and in the heterocysts being frequently longer and found singly or in chains of 2 to 3.

31. *Tolyphothrix fragilis* (Gardner) Gentler Gentler, *op. cit.*, 1930-32, p 725, Fig 465

Lat. fil., 5.5-7  $\mu$ , lat. trich., 4.9-6  $\mu$ , long. cell., 3-5  $\mu$ , lat. het., 6.6-7  $\mu$ ; long. het., 4-10  $\mu$

Habitat.—On the white-washed wall of the temple at Kanwa, Benares, along with *Lyngbya truncata* and others

Genus *Scytonema* Agardh

32. *Scytonema coactile* Mont. Gentler, *op. cit.*, 1930-32, p 753, Fig 479 a-c

Lat. fil., 17-19.8  $\mu$ , when old upto 23  $\mu$ , crass. vag., 2-3  $\mu$ , when old upto 4.8  $\mu$ , lat. trich., 10-15  $\mu$ , when old narrowed down to 7  $\mu$ , lat. het., 13.2-15  $\mu$ , long. het., 11.8-17.3  $\mu$

Habitat.—Free-floating in a tank of a private garden

33. *Scytonema stuposum* (Kütz.) Born. Frémy, *op. cit.*, 1930, p 305, Fig 260, Tilden, *op. cit.*, 1910, Pl XII, Figs 13 and 14

Lat. fil., 10.8-23  $\mu$ , when old upto 26  $\mu$ , crass. vag., 2.4-8  $\mu$ , when old and unhealthy upto 6  $\mu$ , lat. trich., 10-14  $\mu$ , when old and unhealthy narrowed down to 8  $\mu$ , long. cell., 3.3-13.2  $\mu$ , lat. het., 13.2-15  $\mu$ , long. het., 11.8-19  $\mu$

Habitat.—On clayey soil by the side of the road leading to Saranath

The sheath in this form is yellowish-brown and striated

34. *Scytonema ocellatum* Lyngbye Frémy, *op. cit.*, 1930, p 309, Fig 263

Lat. fil., 17.6-19.2  $\mu$ , when old upto 20  $\mu$ , crass. vag., 2.4-3.2  $\mu$ , when old upto 5  $\mu$ , lat. trich., 9.6-14.4  $\mu$ , when old and unhealthy narrowed down to 8  $\mu$ , long. cell., 5-14.4  $\mu$ , when old and unhealthy upto 19  $\mu$ , lat. het., 14.4-16  $\mu$ , long. het., 11.2-16  $\mu$

Habitat.—On moist soil in shade, on the bark of *Tamarindus indica* and *Mangifera indica*

35. *Scytonema javanicum* (Kütz.) Born. Frémy, *op. cit.*, 1930, p 310, Fig 264, Ghose, "On some Myxophyceæ from Rangoon," *Journal of the Burma Research Society*, Vol XV, Part III, 1926, Pl VII, Fig 18.

Lat. fil., 13.2-16.5  $\mu$ , when old upto 18  $\mu$ , crass. vag., 0.9-1.8  $\mu$ , when old upto 3.5  $\mu$ , lat. trich., 11.8-13.2  $\mu$ , when old narrowed down to 8  $\mu$ ; long. cell., 2-11.8  $\mu$ , lat. het., 11.8-13.2  $\mu$ , long. het., 10-16.5  $\mu$

Habitat.—On shaded soil at the edge of a rain-water pool

36 *Scytonema guyanense* (Mont.) Born et Flah Frémy, *op. cit.*, 1930, p. 312, Fig. 265

Lat fil., (13.2-) 15-16.5  $\mu$ , when old upto 20  $\mu$ , crass vag., 2-3  $\mu$ , when old upto 4  $\mu$ , at apices thinned out to 1  $\mu$ , lat cell., 11-14  $\mu$ , when old narrowed down to 9  $\mu$ , long cell., 4.8-19.8  $\mu$ , lat. het., 10-14  $\mu$ , long het., 13-20  $\mu$

Habitat —On moist rocks, among liverworts and mosses, Mussoorie

37 *Scytonema Hofmanni* Ag. Frémy, *op. cit.*, 1930, p. 313, Fig. 266. Geitler, *op. cit.*, 1925, p. 268, Fig. 317

Lat fil., 6.4-8  $\mu$ , when old upto 10  $\mu$ , crass vag., 1-1.5  $\mu$ , when old upto 2.2  $\mu$ , lat cell., 4.7-6.4  $\mu$ , long cell., 4.8-9  $\mu$ , lat het., 6.4  $\mu$ , long het., 9.6-12  $\mu$

Habitat —On the cemented compound wall of a private garden

38 *Scytonema mirabile* (Dillw.) Born Geitler, *op. cit.*, 1930-32, pp. 776 and 777, Fig. 498 a-f, Frémy, *op. cit.*, 1930, p. 318, Fig. 268. Bhāradwāja, "The Taxonomy of *Scytonema* and *Tolyphothrix* including some new records and new species from India and Ceylon," *Revue Algalogique*, 1933, n. 1-2, p. 171, Fig. 5, A

Lat fil., 13.2-16.5  $\mu$ , when old upto 19  $\mu$ , crass vag., 3.3-4  $\mu$ , when old upto 6.6  $\mu$  and thinned out at apices to 1.2  $\mu$ , lat trich., 6.0-9.5  $\mu$ , when old narrowed down to 4  $\mu$ , long cell., 3.6-6  $\mu$ , lat het., 6.6-8  $\mu$ , long het., 6.6-10  $\mu$

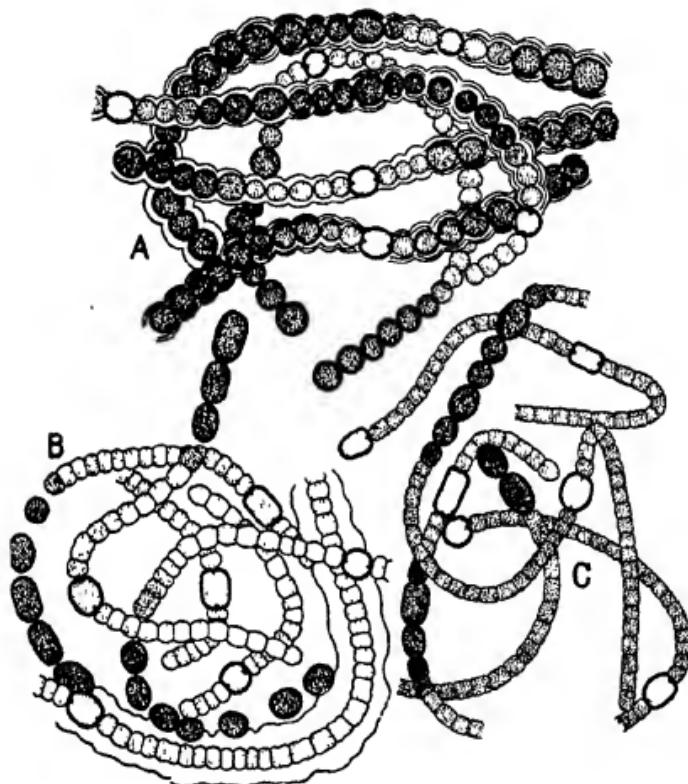
Habitat —On the moist bricks of a wall in the shade of a tree; on moist soil in a field

39 *Scytonema Bewsii* Fritsch and Rich "Contributions to our knowledge of the Freshwater Algae of Africa 4 Freshwater and Sub-aerial algae from Natal," *Trans Roy Soc S Africa*, Vol XI, 1924, p. 364, Fig. 23

Lat fil., 10-16  $\mu$ , when old upto 19  $\mu$ , crass vag., in old healthy filaments upto 4.9  $\mu$ , in old unhealthy ones upto 7  $\mu$ , lat trich., 4.8-7  $\mu$ , in old unhealthy ones narrowed down to 3.2  $\mu$ , at growing apices upto 10  $\mu$ , long cell., 11.2-23  $\mu$ , at apices 3.5-4.8  $\mu$ , lat. het., 6.6-10  $\mu$ , long het., 6.6-16.5  $\mu$

Habitat —On the mud settled down on the rocks near the dam at Latif Shah, Benares State

The stratification of the sheath in this form is mostly diverging



*Limckia* (Roth.) Born. et Flah. var. *arvensis* var. nov., B—*l*  
*Ag.* var. *varians* var. nov., C—*l*

5. *Nostocaceae*Genus *Cylindrospermum* Kuetzing

40 *Cylindrospermum muscicola* Kütz. Frémy, *op. cit.*, 1930, p. 377, Fig. 313, Tilden, *op. cit.*, 1910, Pl. X, Fig. 6, Ghose, *op. cit.*, 1926, Pl. VII, Fig. 15

Lat cell, 2.8-3  $\mu$ , long cell, 2.8-5  $\mu$ , lat het, 3.7-4.5  $\mu$ ; long het, 4.5-6 (-7.5)  $\mu$ , lat spor, 9-10.5  $\mu$ , long spor, 13.2-16.5 (-21.4)  $\mu$

Habitat —On moist soil, singly or along with *Microcoleus chthonoplastes*, *Chroococcus pallidus* and others

Another form collected a few miles off Benares possesses heterocysts, which sometimes measure as much as 10.5  $\mu$  long

Genus *Nostoc* Vaucher

41 *Nostoc paludosum* Kütz. Frémy, *op. cit.*, 1930, p. 333, Fig. 275, Tilden, *op. cit.*, 1910, Pl. 6, Fig. 38

*Forma*

Lat cell, 3-4  $\mu$ , long cell, 3.8-4.8  $\mu$ , lat het, 5.6-6.4  $\mu$ , long het, 6.6-8  $\mu$ , lat spor, 4.6-6.4  $\mu$ , long spor, 6.4-10  $\mu$

Habitat —In the stagnant water of a crop-field along with *Nostoc piscinale* formae and others

The form differs from the type in having bigger spores

42 *Nostoc Linckia* (Roth) Born et Flah. Frémy, *op. cit.*, 1930, p. 333, Fig. 276, Tilden, *op. cit.*, 1910, Pl. VII, Fig. 1, West, *Algæ*, 1916, Vol. I, p. 43, Fig. 31, A-C

Var. *arvense* var. nov (Fig. 4, A)

Plant-mass gelatinous, expanded, presenting an uneven surface, yellowish-brown to blue-green, filaments numerous, flexuous, trichomes frequently enveloped by a lamellated and yellowish-brown mucilaginous sheath that follows their contour, cells spherical or barrel-shaped, heterocysts almost spherical, usually not enveloped by any mucilage, spores in long chains, more or less spherical with a brown outer wall

Lat cell, 4.5-6  $\mu$ , long cell, 4.8-6.4  $\mu$ , lat het, 4.8-7.2  $\mu$ , long het, 4.8-6.4  $\mu$ , lat spor, 6.4-7.2  $\mu$ , long spor, 7-8  $\mu$

Habitat —On water-logged soil in a crop-field

The variety is comparable to the type on account of its possessing (1) flexuous trichomes, (2) spherical or barrel-shaped cells, (3) almost spherical heterocysts and spores with a brown outer wall; but it differs

from the same in the broader trichomes and heterocysts, and in commonly possessing a lamellated and brown mucilaginous sheath following the contour of the trichomes.

43 *Nostoc piscinale* Kütz Frémy, *op cit.* 1930, p. 334, Fig. 277

*Forma*

Lat cell, 4-6  $\mu$ , lat het, 6 4-7 2  $\mu$ , lat spor, 6-10  $\mu$

*Habitat*—In the stagnant water of a crop-field along with *Nostoc paludosum* and others.

The form differs from the type in the slightly bigger dimensions of all parts.

44. *Nostoc spongiforme* Ag. Frémy, *op cit.* 1930, p. 338, Fig. 279 a and b, Tilden, *op cit.* 1910, Pl. VII, Figs. 4 and 5

*Var. varians* var nov (Fig. 4, B)

Plant-mass thin, spreading, blue-green, trichomes loosely entangled, sometimes individually enveloped by yellowish-brown mucilage, cells barrel-shaped, end-cell conical with a rounded apex, heterocysts barrel-shaped or cylindrical with rounded or flat ends, broader than the trichomes, spores in long chains, cylindrical with rounded ends, sometimes ellipsoidal, rarely spherical, with a smooth hyaline outer wall.

Lat cell, 3-3.5  $\mu$ , long cell, 2-3 5  $\mu$ , lat het, 4 8-6 4  $\mu$ , long het, 5 6-8  $\mu$ , lat spor, 4-4.8  $\mu$ , long spor 4 8-10  $\mu$

*Habitat*—On moist soil at the edge of a rain-water pool.

The variety resembles the type in the loosely entangled filaments, the barrel-shaped cells, the barrel-shaped or cylindrical heterocysts, and the chains of ellipsoidal spores, but it differs in having a thin, blue-green and spreading stratum without any firm mucilaginous envelope, narrower cells that are never cylindrical and smaller heterocysts and spores, the latter being cylindrical or sometimes spherical with a hyaline outer wall.

45 *Nostoc ellipsosporum* Rabenh. Frémy, *op cit.* 1930, p. 339, Fig. 280, Tilden, *op cit.* 1910, Pl. VII, Figs. 9 and 10

*Var. violacea* var nov (Fig. 4, C)

Plant-mass gelatinous, irregularly expanded, dark-violet, filaments flexuous, loosely entangled, light-violet, cells almost quadratic or cylindrical, with constrictions at the joints, heterocysts almost spherical or barrel-shaped or cylindrical, with rounded or flat ends, spores ellipsoidal almost spherical or cylindrical, with a hyaline smooth outer wall.

Lat cell, 3.2-3.5  $\mu$ , long cell, 2 4-8  $\mu$ , lat het, 4-6 4  $\mu$ , long het, 4.8-8  $\mu$ , lat spor, 4.8-6 4  $\mu$ ; long spor, 5 6-15  $\mu$ .

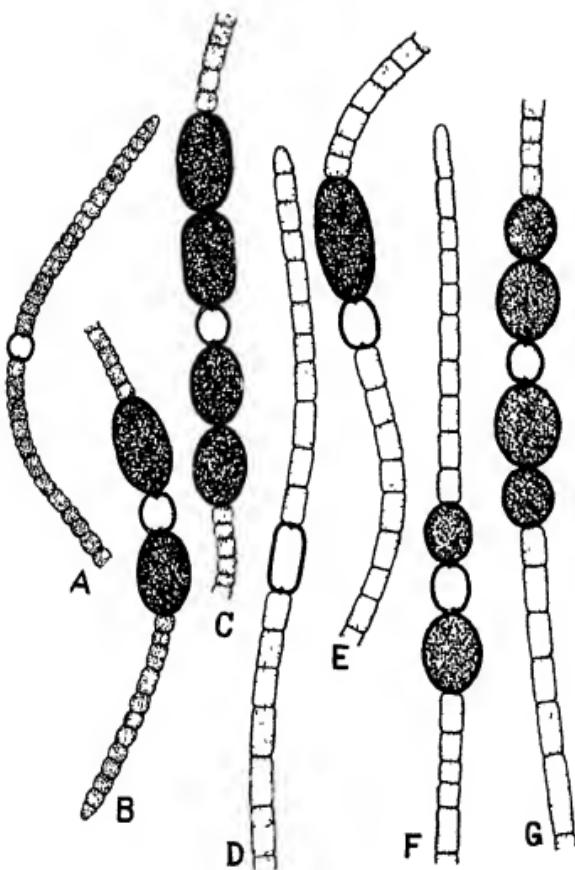


FIG. 5. *Anabana Iyengari Bharadwja* var. *tennis* var. nov.; *anispora* Garduer var. *crassa* var. nov.; F-G—*Anabana aphanizomenoides* Porti var. *elipsospora* var. nov. All  $\times 875$ .

Habitat:—On the sides of the steps of a village tank, above water-level.

The variety agrees with the type in the flexuous and laxly entangled filaments, in the cylindrical cells, in the spherical or cylindrical heterocysts, and in the long ellipsoidal or cylindrical spores with a hyaline smooth outer wall; but it differs in having a dark-violet stratum, narrower trichomes of light violet colour, comparatively shorter cells, smaller heterocysts, that are also barrel-shaped, and in the smaller dimensions of the spores possessing a hyaline outer wall.

Genus *Anabaena* Bory

46 *Anabaena ambigua* Rao Rao, "A new species of *Anabaena* (*Anabaena ambigua* sp nov)," *Proc Ind Acad Sci*, 1937, Vol V, No 3, Sec B, pp 102 and 103, Figs 1 and 2.

Long vag, 300-500 (-1,000)  $\mu$ , crass vag, 10-50  $\mu$ , long trich, 250-300  $\mu$ ; lat cell, 4.9-6.0  $\mu$ , long cell, 3.3-5  $\mu$ , lat het, 6.4-9 (-10)  $\mu$ , lat spor, 8.4-10.9  $\mu$ , long spor, 13.3-16.2  $\mu$ .

Habitat:—Free-floating or attached to grass-blades and roots of *Lemna* and *Trapa* in some ponds near the Benares Hindu University.

47 *Anabaena Iyengari* Bhāradwāja Bhāradwāja, "The Myxophyceæ of the United Provinces, India—I," *Proc Ind Acad Sci*, Vol 2, No 1, Sec B, 1935, Fig 6, H-K.

*Var tenuiss* var nov (Fig 5, A-C)

Plant-mass floccose, thin, free-floating, pale blue-green. Trichomes single, straight or irregularly curved, end-cells conical with rounded apices. Cells barrel-shaped, as long as broad or slightly shorter or longer than broad. Heterocysts more or less barrel-shaped, sometimes sub-spherical. Spores ellipsoidal or cylindrical with rounded ends, single or in pairs on either side of a heterocyst, with a smooth hyaline outer wall.

Lat cell, 3.5-4.5  $\mu$ , long cell, 3-6.4  $\mu$ , rarely 7.5  $\mu$ , lat het, 4.8-6.4  $\mu$ , long het, 5.2-9  $\mu$ , lat spor, 7.5-9.6  $\mu$ , rarely 10.5  $\mu$ , long spor, 9-19.5  $\mu$ , rarely 21  $\mu$ .

Habitat:—In a pond on the University grounds, along with *Dactylococcopsis raphidioides* forma, *Calothrix marchica* var *intermedia*, *Anabaena fertilissima* sp nov, *Oscillatoria animalis*, *Lynghya Digeuti* and sterile filaments of *Oedogonium* and *Spirogyra*.

The variety resembles the type in the barrel-shaped cells, conical end-cells with rounded apices, barrel-shaped heterocysts and ellipsoidal spores, that are on either side of a heterocyst, but differs from the same

in having narrower trichomes, smaller heterocysts and comparatively smaller spores (that are also cylindrical) with a hyaline outer wall and situated singly or in pairs on either side of a heterocyst

48 *Anabaena unispora* Gardner Geitler, *op cit.*, 1930-32, p 901, Fig 580 b

Var *crassa* var nov (Fig 5, D and E)

Plant-mass soft, mucilaginous Trichomes free, long, more or less straight, tapering at extreme ends, constricted at joints, end-cells with rounded apices Cells cylindrical, sometimes almost quadratic Heterocysts single, cylindrical or ellipsoidal, sometimes pressed from both ends Spores single, adjoining heterocysts, long ellipsoidal, sometimes pressed from both ends, with a smooth hyaline outer wall

Lat cell, 4.8-6.6  $\mu$ , long trich, 500-700  $\mu$ , long cell, 4.5-13.2  $\mu$ , lat het, 5-8.2  $\mu$ , long het, 7.4-16.5  $\mu$ , lat spor, 9.6-15  $\mu$ , long spor, 23-31.5  $\mu$

Habitat —In a rain-water pool, along with sterile filaments of *Spirogyra* and *Oedogonium*, floating on the stagnant water of a rice-field

The variety agrees with the type in possessing cylindrical cells, constrictions at septa, end-cells with rounded apices and single ellipsoidal spores on only one side of a heterocyst\*, but it differs from the same in having broader trichomes and narrower spores with a hyaline outer wall

49 *Anabaena aphanizomenoides* Forti Geitler, *op cit.*, 1930-32, p 876, Fig 556

Var *ellipospora* var nov (Fig 5, F and G)

Thallus free-floating, thin, floccose, pale blue-green Trichomes single, straight or bent, tapering at the ends, with constrictions at the joints, end-cells conical with rounded apices Cells quadratic or upto three times as long as broad Heterocysts single, intercalary, ellipsoidal, rarely spherical Spores ellipsoidal sometimes pressed from both ends, one or two on each side of a heterocyst, with a hyaline outer wall

Lat trich, 3.3-4  $\mu$ , at top 1.6  $\mu$ , long cell, 3.3-11  $\mu$ , lat het, 6-6.6  $\mu$ , long het, 6-11.6  $\mu$ , lat spor, 9.6-13.5  $\mu$ , long spor, 11.6-19.8 (-23)  $\mu$

\* Since the author of *Anabaena unispora* has not given any idea of the shape and the actual dimensions of the heterocysts, it is not possible to compare them with those of the present form. But the original figure of the type shows that the heterocysts are cylindrical as found in the form under discussion. The Benares alga, however, possesses ellipsoidal heterocysts as well

Habitat.—In a rain-water pool, along with *Aulosira Fritschii*, *Cosmarium* sp., *Closterium* sp., and several others

The variety agrees with the type in possessing quadratic and cylindrical cells, ellipsoidal heterocysts and one or two spores on each side of a heterocyst, the former with a hyaline outer wall, but it differs in the slightly narrower trichomes, in the absence of gas-vacuoles in the cells, in the longer heterocysts and in the bigger ellipsoidal spores

50 *Anabaena sphaerica* Born et Flah var *attenuata* Bhāradwāja Bhāradwāja, *op. cit.*, 1935, p 104, Fig 5, G and H

Lat trich., 3-3-5  $\mu$ , long cell, 3 3-4  $\mu$ , lat het, 4 8-6 4 (-7 5)  $\mu$ , lat spor, 10-12  $\mu$ , long spor, 9 8-14 6  $\mu$

Habitat.—On soil submerged in water at the edge of a pool

This variety, as originally reported, possessed only single spores on either side of a heterocyst, but in the present form, spores are also met with in pairs on each side of a heterocyst

51 *Anabaena fertilissima* sp nov (Fig 6, A-C)

Trichomes single, straight or bent, with almost rounded end-cells, cells barrel-shaped, heterocysts almost spherical, spores in long chains, often making the whole trichome sporogenous, adjoining the heterocysts but developed centrifugally, almost spherical, with a smooth hyaline outer wall

Long trich., upto 350  $\mu$ , lat trich., 5-5 6  $\mu$ , at apex 4  $\mu$ , long cell, 4-8-8  $\mu$ , lat het, 6 4-8 4  $\mu$ , lat spor, 4 8-8  $\mu$ , long spot, 3 2-8 8  $\mu$

Habitat.—In a pond on the University grounds, along with *Dactylococcopsis raphidioides* forma, *Calothrix marchica* var *intermedia*, *Anabaena Iyengari* var *tenuis*, *Oscillatoria animalis*, *Lyngbya Digens* and sterile filaments of *Spirogyra* and *Oedogonium*

The important feature in this alga is the formation of spores in long chains, so much so that practically the whole of the trichome becomes sporogenous. It comes close to *Anabaena sphaerica* Born et Flah on account of the barrel-shaped cells, rounded end-cells and spherical heterocysts and spores, but it differs in the spores being smaller and formed in long chains. In the presence of spherical heterocysts and spores in chains, the present alga agrees with *Anabaena gelatinicola* Ghose, but it differs from the Lahore form in the presence of narrower trichomes that are never coiled, rounded end-cells and much longer chains of smaller spores adjoining the heterocysts. The barrel-shaped cells and the chains of spores

characteristic of this alga also recall *Anabaena variabilis* Kütz., *A. aeruginosa* Gardner, *A. Iyengari* Bhāradwāja, and *A. dolosum* Bhāradwāja, but it differs from all these four species in the spores being spherical. It further differs from the former two species in the presence of spores adjoining the heterocysts and from the latter two species in possessing spherical heterocysts.

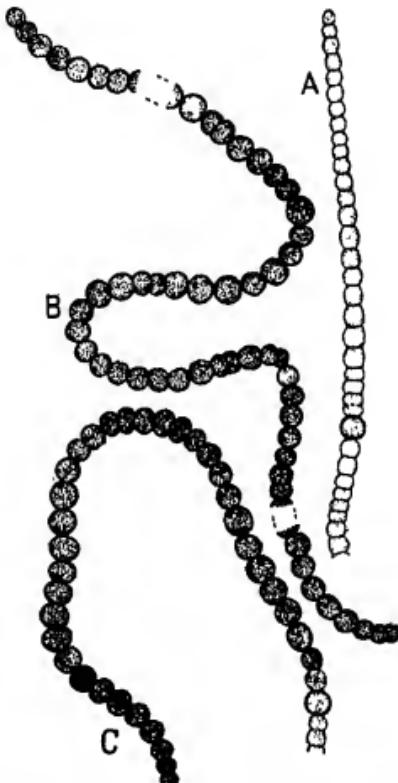


FIG. 6. A—vegetative and B & C—sporogenous portions of filaments of *fertilissima* sp. nov. All  $\times 875$ .

6. *Oscillatoriaceæ*

Genus *Spirulina* Turpin

52. *Spirulina platensis* (Nordst.) Gom. Geitler, *op. cit.*, 1930-32, p 922, Fig 590 d, Frémy, *op. cit.*, 1930, p 232, Fig 205

Var *tenuis* var nov (Fig 7, A and B)

Plant-mass greyish-brown, trichomes pale blue-green, of uniform width, in regular spirals, without constrictions at the joints, end-cells with rounded apices, cells disc-shaped

Lat trich., 5.1-6.4  $\mu$ ; long cell, 1.0-2.5 (-3.3)  $\mu$ , lat spir., 30-43  $\mu$ ; spot inter duo spir., 40-52  $\mu$

Habitat:—In a waste-water drain, in a pond, along with *Oscillatoria chalybea*, Raighat

The variety agrees with the type in the uniform width of the trichomes, in the rounded apices of the end-cells, in the spirals being regular, and in the distance between the spirals, but it differs on account of the narrower trichomes, the broader spirals, the comparatively shorter cells, and the absence of constrictions at the septa

53. *Spirulina major* Kütz. Geitler, *op. cit.*, 1930-32, p 930, Fig 595, Frémy, *op. cit.*, 1930, p 235, Fig 208, Tilden, *op. cit.*, 1910, Pl IV, Fig 46, Frémy, *op. cit.*, 1934, Pl 31, Fig 18, Carter, "A comparative study of the algal flora of two salt marshes, Part II," *Journal of Ecology*, Vol XXI, I, 1933, p 159, Fig 2, Ghose, *op. cit.*, 1926, Pl VI, Fig 3

Lat trich., 1.1-1.4  $\mu$ , lat spir., 3.2-3.5  $\mu$ , spot inter duo spir., 2.4-3.3  $\mu$

Habitat:—On moist soil along with other algae, planktonic in the tank of the Benares Electric Light and Power Supply Co., Ltd., along with *Chroococcus turgidus*, *C. minutus* and *Merismopedia tenuissima*

Genus *Oscillatoria* Vauch

54. *Oscillatoria sancta* (Kütz.) Gom. Frémy, *op. cit.*, 1930, p 210, Fig 177; Tilden, *op. cit.*, 1910, Pl IV, Fig 5, Carter, *op. cit.*, 1933, p 159, Figs 11 and 12, Geitler, *op. cit.*, 1925, p 356, Fig 418

Lat cell., 10-16  $\mu$ ; long cell, 2.5-4.8  $\mu$

Habitat:—On moist soil amidst flower pots in the Green House, University Botanical Garden, along with *Oscillatoria Ulrichii*, on moist soil along with *Oscillatoria formosa*

*Forma*

Lat cell, 9.6-11.2  $\mu$ , long cell, 2-4  $\mu$

The form has no constrictions at the joints

55 *Oscillatoria obscura* Brühl and Biswas "Algae of the Bengal Filter-beds," *Journal of the Department of Science, Calcutta University*, 1922, Vol 4, Pl II, Fig 9

Lat cell, 4.1-4.8  $\mu$ , long cell, 1-1.3  $\mu$

Habitat —On wet soil along with *Navicula* sp

56 *Oscillatoria anguina* (Bory) Gom Gentler, *op cit*, 1930-32, p 945, Fig 599 b

Lat trich, 6.6-8.2  $\mu$ , long cell, 1.5-2.5  $\mu$

Habitat —In a waste-water drain; in a rain-water puddle. Chunar

57 *Oscillatoria chalybea* Mertens Gentler, *op cit*, 1930-32, p 956, Fig 608 b

Lat cell, 7-9.6  $\mu$ , at top 6.6  $\mu$ , long cell, 3-6.6  $\mu$

Habitat —On wet soil, along with *Oscillatoria homogenea*, Mussoorie

58 *Oscillatoria tenuss* Ag Gentler, *op cit*, 1930-32, p 960, Fig 611 g, Tilden, *op cit*, 1910, Pl IV, Fig 17

Lat cell, 4.5-6.6  $\mu$ , long cell, 2.5-5  $\mu$

Habitat —On moist soil, in a road-side water course, Shivpur, Benares, along with other algae, in rain-water ditches on waste ground

59 *Oscillatoria pseudogeminata* G Schmidle Gentler, *op cit*, 1930-32, p 966, Fig 616

Lat cell, 1.6-1.8  $\mu$ , long cell, 1-2  $\mu$

Habitat —On moist soil

*Forma*

Lat trich, 1.6-2  $\mu$ , long cell, 1.8-5  $\mu$

Habitat —In stagnant water of a ditch near a well

The form possesses much longer cells as well

60 *Oscillatoria quadripunctulata* Brühl and Biswas Brühl and Biswas, *op cit*, 1922, p 5, Pl I, Fig 6, Biswas, "Road Slimes of Calcutta," *Jour Dept Sci, Cal Univ*, 1925, Vol VII, p 10, Pl II, Fig 11 a-d, Gentler, *op cit*, 1930-32, p 966

Lat trich, 1.6-1.8  $\mu$ ; long cell, 3.6-4  $\mu$

Habitat —On moist soil along with other algae; in a pond along with other algae

61. *Oscillatoria homogenea* Frémy Frémy, *op. cit.*, 1930, p. 221.  
Fig. 184

Lat. cell, 3 3-3.5  $\mu$ , long cell, 3 5-6  $\mu$

Habitat —On wet soil, along with *Oscillatoria chalybea*, Mussoorie

62 *Oscillatoria Okeni* Ag. Tilden, *op. cit.*, 1910, Pl. IV, Fig. 35,  
Gentler, *op. cit.*, 1925, p. 372, Fig. 463

Lat. cell, 5 5-6  $\mu$ , long cell, 3-4 5  $\mu$

Habitat —On moist soil along with *Oscillatoria sancta* and others,  
on water-logged soil

63 *Oscillatoria formosa* Bory Gentler, *op. cit.*, 1930-32, p. 970.  
Fig. 619 b

Lat. trich., 4 5-6 6  $\mu$ , long cell, 1 8-4 5  $\mu$

Habitat —On moist soil, along with *Oscillatoria pseudogeminata*, on  
moist rocks, along with *Phormidium subsuscum*, Mussoorie, on moist soil,  
along with *Oscillatoria sancta* and *O. princeps*, on soil, along with other  
algae, in a waste-water drain

64 *Oscillatoria claricentrosa* Gardner Gentler, *op. cit.*, 1930-32,  
p. 984, Fig. 615 c

Forma *bigranulata* form. nov (Fig. 7, C)

Lat. trich., 2-2.5  $\mu$ , long cell, 5-10  $\mu$

Habitat —In stagnant water of a drain, along with sterile filaments  
of *Spirogyra* and *Oedogonium*

The form differs from the type in the presence of two granules on either  
side of a cross-wall and in the trichomes showing a more gradual tapering

65 *Oscillatoria rubescens* D C. Gentler, *op. cit.*, 1930-32, p. 973.  
Fig. 620 a and b

Forma.

Lat. trich., 4 8-6 4  $\mu$ , long cell, 1 2-4  $\mu$

Habitat —On wet soil, Chunar

But for the presence of narrower trichomes, the Benares form agrees  
with the type in all respects

66 *Oscillatoria Lemmermanni* Woloz Gentler, *op. cit.*, 1930-32,  
p. 969, Fig. 618 i

Lat. trich., 2 2-2.8  $\mu$ , long cell, 2.5-5  $\mu$

Habitat —On moist soil along with *Phormidium fadsianum* forma

67 *Oscillatoria animalis* Ag. Gentler, *op. cit.*, 1930-32, p. 950,  
Fig. 603 e

Lat. trich., 2.5-3.8  $\mu$ ; long cell., 2.5-2  $\mu$ .

**Habitat** :—On the moist sides of the cemented pavement of a well; in a pond on the University grounds, along with *Dactylococcus raphidoides* forma, *Calothrix marchica* var. *intermedia*, *Anabaena fertilissima* sp. nov., *A. Iyengari* var. *tenuis*, *Lyngbya Digeoti* forma and sterile filaments of *Spiragyna* and *Oedogonium*, in a ditch along with *Oscillatoria sancta* and others.

68 *Oscillatoria acuminata* Gom Tilden, *op. cit.*, 1910, Pl. IV, Fig. 29.

*Forma*

Lat. cell., 3.3-3.8  $\mu$ ; long cell., 1.3-3.8  $\mu$ .

**Habitat** —On moist soil.

The form differs from the type in the smaller dimensions of the cells.

69 *Oscillatoria salina* Brühl and Biswas Geitler, *op. cit.*, 1930-32, p. 979, Fig. 624.

Lat. cell., 3.3-4.8  $\mu$ , long cell., 1.5-2  $\mu$ .

**Habitat** —In earthen water-pans in a green house, along with *Cylindrospermum indicum* and *Coccones* sp., on moist soil in a drain, Cantonment area, on moist soil, University area.

Unlike the type, the Benares alga is a fresh-water form.

*Genus Phormidium Kütz*

70 *Phormidium Bohneri* Schmidle "Beiträge zur Algenflora Afrikas," Engler's *Botanische Jahrbücher*, 1902, 30, Taf. II, Fig. 11.

*Forma*

Lat. fil., 3-3.7  $\mu$ ; lat. cell., 2.2-2.8  $\mu$ , long. cell., 1.6-2.8  $\mu$ .

**Habitat** —On the cemented surface near the waste-water outlet of a house, Ramnagar.

The form differs from the type in the presence of the broader and the closely entangled filaments with a persistent sheath.

71 *Phormidium cebennense* Gom Frémy, *op. cit.*, p. 147, Fig. 129.

Lat. trich., 1.8-2  $\mu$ ; long. cell., 1-2  $\mu$ .

**Habitat** —On the plinth of the College building near the outlet of a drain, along with *Phormidium anomala* sp. nov.

72 *Phormidium mucosum* Gardner Geitler, *op. cit.*, 1930-32, p. 1012, Fig. 646 b.

Var. *arvense* var. nov (Fig. 7, D and E).

Plant-mass thick, fragile, blue-green to greyish-blue. Filaments loosely entangled. Sheath thick, firm, hyaline, unstratified. Trichomes of uniform

width, not constricted at the joints; apical cells with rounded apices, without calyptra or cap. Cells quadratic or almost quadratic or slightly longer than broad.

Lat fil, 3.2-5.2  $\mu$ ; crass vag, upto 2  $\mu$ , lat trich, 1.8-2.5  $\mu$ , long cell, 1.5-3.6  $\mu$

Habitat:—In stagnant rain-water of a crop-field

The variety agrees with the type in all respects except that it has narrower filaments and trichomes, the latter possessing shorter cells

73 *Phormidium Retzii* (Ag.) Gom. Geitler, *op. cit.*, 1930-32, p. 1012, Fig. 647 a-d.

Lat fil, 4.5-7  $\mu$ , long cell, 4.5-8  $\mu$

Habitat—On the sides of the water storage tanks of the Benares Water Works, on moist stones near the dam, Latif Shah, Benares State

74 *Phormidium ambiguum* Gom. Geitler, *op. cit.*, 1930-32, p. 1012, Fig. 647 e, Ghose, "The Myxophyceae of Rangoon, II," *Journ. Burma Res. Soc.*, 1927, Vol. XVI, Part III, pp. 220-26, Pl. XI, Fig. 4

Forma

Lat cell, 3.4  $\mu$ , long cell, 1.5-2.8  $\mu$

Habitat:—On the sides of the cemented water reservoir, University Botanical Garden

The form has slightly narrower trichomes than those of the type

75 *Phormidium subincrassatum* Fritsch and Rich. Fritsch and Rich, "Contributions to our knowledge of the Fresh-water Algae of Africa, 7, Fresh-water algae (exclusive of Diatoms) from Oruqualand West," *Trans. Roy. Soc. S. Africa*, 1929, Vol. LXIII, Parts 1 and 2, p. 84, Fig. 27 I-L

Lat trich, 4.7-5.8  $\mu$ , crass vag, upto 0.5  $\mu$ ; long cell, 2.5-8  $\mu$

Habitat—On moist soil, along with *Microcoleus chthonoplastes* and others.

76 *Phormidium subfuscum* Kütz. Geitler, *op. cit.*, 1930-32, p. 1023, Fig. 652 d-g

Lat cell, 8.4-11.7  $\mu$ , long cell, 1.8-3  $\mu$

Habitat.—On moist soil, along with *Oscillatoria formosa*

The calyptra in this form is rounded instead of being pointed.

77. *Phormidium favosum* (Bory) Gom. Geitler, *op. cit.*, 1930-32, p. 1023, Fig. 652 a and b

Forma.

Lat. cell., 4-6  $\mu$ ; long cell, 2.4-3.5  $\mu$ .

Habitat:—On the sides of a big water reservoir.

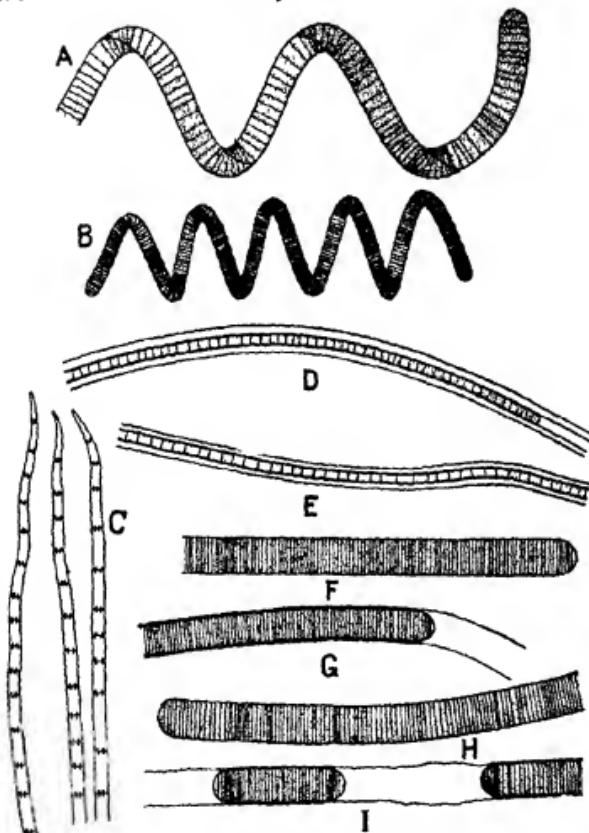


FIG 7. A—Terminal portion of the trichome and B—entire plant of *Sphaerulina platenae* (Nordal) Gom var *tenuis* var nov., C—Terminal portions of *Oscillatoria claricentroa* Gardner forms *granulata* form nov., D & E—portions of filaments of *Phormidium mucorinum* Gardner var *arvensis* var nov.; F—I—portions of filaments of *Phormidium anomalo* sp. nov. B  $\times 425$ , all the rest  $\times 675$

The form exactly resembles the type except for the presence of shorter cells

78 *Phormidium autumnale* (Ag.) Gom. Geitler, *op. cit.*, 1930-32, p. 1023, Fig. 652 *k* and *l*, Frémy, *op. cit.*, 1934, Pl. 24, Fig. 4, Carter, *op. cit.*, 1933, p. 156, Fig. 7

Lat cell, 4-4.8  $\mu$ , long cell, 2.4-4.8  $\mu$

Habitat—On the stony steps of a tank near water-level

79 *Phormidium anomala* sp. nov. (Fig. 7, *F-I*)

Thallus thick, expanded, soft, mucilaginous, deep blue-green to green. Trichomes sub-parallel, of uniform width, without constrictions at the joints. Sheath thin, colourless, unstained with chlor-zinc-iodide, persistent or dissolved. Cells disc-shaped, much broader than long, end-cells bluntly rounded, without cap or calyptra

Crass. strat., 3-6 mm., lat. trich., 8-10  $\mu$ , long cell, 0.8-1.2 (-2)  $\mu$

Habitat—On the plinth of the College building, near the outlet of a drain, along with *Phormidium cebennense*

This alga approaches *Phormidium ambiguum* Gom. and *Phormidium subincrustatum* Fritsch and Rich in the presence of an expanded stratum, the uniform width of the trichomes, the absence of constrictions at the joints, and the rounded apices for the end-cells, which are without a cap or calyptra, but it differs from both these species in having shorter and wider cells\*. It further differs from the former species in the filaments being more or less parallel, the sheath being always thin and unstratified, and remaining unstained with chlor-zinc-iodide, the absence of granules near the septa and of the gas-vacuoles in the cells. The Benares alga, while further agreeing with *Phormidium subincrustatum* in the sub-parallel trichomes and the thin, diffusible and unstratified sheath, differs from it in the thallus being thicker and without any impregnation of lime

#### Genus *Lyngbya* Agardh

80 *Lyngbya gracilis* Rabenh. Geitler, *op. cit.*, 1930-32, p. 1040, Fig. 657 *a*; Frémy, *op. cit.*, 1934, Pl. 26, Fig. 3

Lat. fil., 8-11.8  $\mu$ , crass. vag., 0.8-1.8  $\mu$ , lat. trich., 5.6-8.3  $\mu$ , long cell, 2-6.4  $\mu$ .

Habitat—In the water storage tank of the Benares Water Works, along with *Chamisiaphon siderophilus* var. *glabra*

\* Fritsch and Rich have not mentioned the length of the cells in *Phormidium subincrustatum*, but their figures show them to be shorter than broad or almost quadratic.

81. *Lyngbya spiralis* Geitler. Geitler, *op. cit.*, 1930-32, p. 1042, Fig. 659  
Lat. fil., 5-5.8  $\mu$ , crass. vag., 0.3  $\mu$ , lat. trich., 4.8-5  $\mu$ , long. cell.,  
1.5-2.5  $\mu$

Habitat.—Planktonic in a rain-water pool, along with *Aphanothecus bullosa*.

82. *Lyngbya limnetica* Lemm. Lammermann, *op. cit.*, 1910, p. 102,  
Fig. 8  
Lat. fil., 1.5-2  $\mu$ , lat. cell., 1.2-1.6  $\mu$ , long. cell., 1-2.5  $\mu$

Habitat.—In an ornamental water reservoir of a private garden, along with *Microcystis aeruginosa* var. *elongata*, *Oscillatoria Anneae* and *Oedogonium* sp.

83. *Lyngbya dendrobsa* Brühl and Biswas. "Commentationes Algologicae in alge epiphytice epiphylloide or Indian Bark Algae," *Jour. Dept. Sci., Cal. Univ.*, 1923, 5, Pl. III, Fig. 11 a-c, Geitler, *op. cit.*, 1930-32,  
p. 1051

*Forma*

Lat. fil., 9.6-13.2  $\mu$ , crass. vag., 0.8-3  $\mu$ , lat. trich., 6.6-8.8  $\mu$ ,  
long. cell., 4-8  $\mu$

Habitat.—On moist soil, spreading among *Riccia* sp. and grass blade.

The form differs from the type in possessing narrower trichomes and thicker sheath.

84. *Lyngbya corticola* Brühl and Biswas. Brühl and Biswas, *op. cit.*,  
1923, Pl. IV, Figs. 13 a-d, Geitler, *op. cit.*, 1930-32, p. 1052.

*Forma*

Lat. fil., 18.4-25.6  $\mu$ , crass. vag., 2.4-8  $\mu$ , lat. cell., 10.4-12.8  $\mu$ ,  
long. cell., 4-8  $\mu$

Habitat.—On moist soil, University area.

The form differs from the type in the sheath being very thick and stratified.

85. *Lyngbya aestuans* Liebm. var. *arbuscula* Brühl and Biswas. Brühl and Biswas, *op. cit.*, 1923, Pl. II, Fig. 9 a-c; Geitler, *op. cit.*, 1930-32, p. 1053

Lat. fil., 22.8-26.4  $\mu$ , crass. vag., 2.5-4  $\mu$ ; lat. trich., 16-18  $\mu$ ,  
long. cell., 4.5-8  $\mu$

Habitat:—On the bark of *Eugenia Jambolana* by the side of the road leading to Allahabad.

86. *Lyngbya arboricola* Brühl and Biswas. Brühl and Biswas, *op. cit.*,  
1923, Pl. III, Fig. 10 a-e, Geitler, *op. cit.*, 1930, p. 1063

Lat fil, 19-23  $\mu$ , crass vag, 2-4  $\mu$ , lat trich, 16-17  $\mu$ ,  
long cell, 4-6.4 (-9)  $\mu$

Habitat—On the bark of *Mangifera indica* along with *Lyngbya truncicola*

87 *Lyngbya truncicola* Ghose Ghose, *op. cit.*, 1923, Pl 31, Fig 6

Lat fil, 15-16.5  $\mu$ , crass vag, 0.8  $\mu$ , lat trich, 12-15  $\mu$ ,  
long cell, 1.5-4.5  $\mu$

Habitat—On the bark of *Bassia latifolia* and *Mangifera indica*, either  
singly or along with *Lyngbya arboricola*, University Botanical Garden

88 *Lyngbya rubra* Frémy Frémy, *op. cit.*, 1930, p 185, Fig 155

*Forma*

Lat fil, 6-8  $\mu$ , lat trich, 4.8-5  $\mu$ , long cell, 4-8  $\mu$

Habitat—In a stagnant pond

The form differs from the type in having slightly broader trichomes,  
shorter cells and a hyaline sheath

89 *Lyngbya ceylanica* Wille Geitler, *op. cit.*, 1930-32, p 1055,  
Fig 668 a, Ghose, "On a collection of Myxophyceæ from Mergui and  
some neighbouring islands," *Journal of the Burma Research Society*, 1927,  
Vol XVII, Part III, pp 244-51, Pl III, Fig 11

Lat fil, 12.8-19  $\mu$ , crass vag, 3.2-4  $\mu$ , lat trich, 9.6-11.2  $\mu$ ,  
long cell, 4.8-6.4  $\mu$

Habitat—On moist soil along with *Microcoleus chthonoplastes*

The sheath in the Benares form is thicker than that of the type

90 *Lyngbya lutea* (Ag) Gom Geitler, *op. cit.*, p 1058, Fig 670 a and  
b, Frémy, *op. cit.*, 1934, Pl 28, Fig 4 a-c, Carter, *op. cit.*, 1933, p 164,  
Fig 5 and 6

Lat fil, 4.8-5  $\mu$ , lat trich, 3.8-4  $\mu$ , long cell, 2.4-3.8  $\mu$

Habitat—In a cemented drain, University Botanical Garden

The sheath in this form is thin and unstratified

91 *Lyngbya Digeri* Gom. Geitler, *op. cit.*, 1930-32, p 1038, Fig 656 c

*Forma*

Lat fil, 1.5-2.3  $\mu$ , lat trich, 1.4-2.2  $\mu$ , long cell, 1.5-3.2  $\mu$

Habitat—In a pond on the University grounds, along with *Dactylo-  
coccopsis raphidiosides* forma, *Calothrix marchica* var. *intermedia*, *Anabaena  
Iyengari* var. *tenuis*, *A. fertilissima* sp nov, *Oscillatoria animalis* and  
sterile filaments of *Spirogyra* and *Oedogonium*

The form is characterised by the possession of narrower filaments.

92 *Lyngbya confluoides* Ag. Tilden, *op cit*, 1910, Pl. V, Fig. 39, Frémy, *op cit*, 1934, Pl. 28, Fig. 2; Carter, *op cit*, 1933, p. 162, Fig. 11, 1 and 2

Lat fil., 19-23.2  $\mu$ , crass vag., upto 5  $\mu$ ; lat trich., 13-19.2  $\mu$ ; long cell, 2-4  $\mu$

Habitat —In an ornamental reservoir of a private garden under the shade of the closely spreading leaves of *Nelumbium* sp.

93 *Lyngbya aeruginosa-cerulea* (Kütz) Gom. Frémy, *op cit*, 1930, p. 193, Fig. 157, Ghose, *op cit*, 1926, Pl. VI, Fig. 7

Lat fil., 6.4-8  $\mu$ , lat trich., 5.1-6.4  $\mu$ , long cell, 1.8-4.8 (-5.6)  $\mu$

Habitat —In a stagnant pond, Chunar

94 *Lyngbya pulealis* Mont. Geitler, *op cit*, 1930-32, p. 1063, Fig. 875 b. Frémy, *op cit*, 1930, p. 193, Fig. 159 a and b

Lat fil., 8-11.2  $\mu$ , crass vag., upto 0.8  $\mu$ , lat trich., 6.4-9.6, average 8  $\mu$ , long cell, 3-10.5  $\mu$

Habitat —In the stagnant water of an irrigation channel, Ramnagar

95 *Lyngbya Mariensisana* Meneghi Geitler, *op cit*, 1930-32, p. 1064, Fig. 876

*Forma*

Lat fil., 8-8.8  $\mu$ , crass vag., 0.8  $\mu$ , lat trich., 5.6-6.4  $\mu$ , long cell, 2-4.8  $\mu$

Habitat —On water plant at the edge of a rain-water pool

The form differs from the type in the sheath being always smooth and the cells being sometimes longer

96 *Lyngbya stagnina* Kütz Geitler, *op cit*, 1930-32, p. 1066, Fig. 679 b.

*Forma*

Lat fil., 10-12  $\mu$ , lat. trich., 8.5-9.5  $\mu$ , long cell, 5-10  $\mu$

Habitat —In a stagnant puddle by the side of the River Ganges along with *Cladophora* sp and *Gomphonema* sp

The form differs from the type in having narrower trichomes and longer cells\* without granules near the septa

\* From the description of the type it is clear that the cells are much shorter than broad, but Skuja's figure shows the cells to be almost quadratic or only slightly shorter than broad, just like those of the form under discussion.

97 *Lyngbya major* Menegh Tilden, *op cit.*, 1910, Pl 5, Fig 46, Geitler, *op cit.*, 1930-32, p 1066, Fig 679a, West, *op cit.*, 1916, p 42, Fig. 28 A

Lat. fil., 19.2-20.8  $\mu$ , crass vag., 3.2-4  $\mu$ , lat trich., 12.8-14.4  $\mu$ , long cell, 2-4  $\mu$

Habitat:—On moist soil in shade

Genus *Symploca* Kützing

98. *Symploca muraria* Kütz Geitler, *op cit.*, 1930-32, p 1125, Fig. 732; Frémy, *op cit.*, 1930, p 129, Fig 113a and b, West, *op cit.*, 1916, p 23, Fig 15 E.

*Forma*

Lat fil., 5.2-7.5  $\mu$ , crass vag., upto 2.5  $\mu$ , lat trich., (3.2-) 4-5  $\mu$ , long cell, 2.4-5.6  $\mu$ .

Habitat:—On moist soil, University area

The form differs from the type in having broader trichomes and sometimes longer cells

Genus *Microcoleus* Desmazières

99 *Microcoleus chthonoplastes* Thuret Geitler, *op cit.*, 1930-32, p 1134, Fig 739; Carter, *op cit.*, 1933, p 166, Figs 15-17.

Diam fil., 30-100  $\mu$ , lat vag., 15-65  $\mu$ , lat cell, 4-5  $\mu$ , long cell, 4-8  $\mu$

Habitat:—On moist soil along with other algae

100 *Microcoleus sociatus* W et G S West Frémy, *op cit.*, 1930, p 83, Fig 85a and b.

Diam fil., 30-45  $\mu$ , lat vag., 2-10  $\mu$ , lat trich., 2.4-2.8  $\mu$ , long cell, 4.8-6  $\mu$

Habitat:—On moist soil along with other algae

The writer takes this opportunity to express his great indebtedness to Professor V Bhadravaja, for his kind guidance and criticism throughout the course of this investigation

## THE PROTEINS OF GROUNDNUT (PEANUT), *ARACHIS HYPOGAEA*, LINN.

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Received August 9, 1937

(Communicated by Rao Bahadur Dr D L Sahasrabuddhe.)

INDIA produces a large variety of oilseeds in abundance. Among them, groundnut is of supreme importance as it occupies a third of the total area under cultivation of oilseeds. The total production is more than three million tons which represents nearly half the world's output of this seed.

The seed is eaten as such or after being fried. But the main use of groundnut is for its oil which is nearly 50 per cent of the seed. The cake which is available in large quantities is mainly used as cattle food or as manure. Many workers have pointed out the high nutritive value of groundnut cake for human consumption (Daniel, 1917, Wallis, 1917) Johns and Jones (1916-18) and Daniel and Menaul (1921) have isolated the proteins of the groundnut cake and analysed them. In India there has been practically no work on the nature of the proteins of oilseeds. The only solitary reference to oilseed proteins is that of Narayana and Srinivasaya (1936) on the proteins of sandal seed, which though edible is not used as a food-stuff. In this investigation, therefore, an attempt is made to get an idea of the nutritive value of groundnut cake by making a complete and detailed analysis of its proteins.

*Material*—The seeds of the two common varieties of groundnut, Local and Spanish, were employed. The oil was expressed in a local *ghans* and the cakes obtained were utilized for the isolation of proteins. Below are given the analyses of the two cakes used in this investigation.

TABLE I

	Local	Spanish
Moisture .	4.31	5.66
*Ash .	4.18	4.83
Ether Extract .	20.84 "	10.05
†Crude Protein (N × 6.25)	52.50	62.22
Crude Fibre .	2.42	3.42
Carbohydrates . .	15.75	13.82
	100.00	100.00

\* Sand . . 0.40 0.81

† Nitrogen . . 8.40 9.95.

*Experimental.*

As the preliminary experiments showed that the presence of even a small quantity of oil in the cake exerts a strong inhibiting action on the peptisation of the proteins, the cake was extracted with ether to make it completely free of oil. 75 per cent of the total nitrogen was extractable with 10 per cent sodium chloride solution at room temperature and most of it could be precipitated as a globulin.

*Isolation of the Proteins—*

*Total Globulins*—The cake was extracted with 10 per cent saline solution. Total globulins were isolated from this extract by (i) dilution and acidification with acetic acid, (ii) by dialysis and (iii) by saturation with ammonium sulphate. Three preparations were thus obtained.

*Arachin and Conarachin*—Johns and Jones (1916-18) have shown that two globulins—Arachin and Conarachin—can be obtained from the saline extract by fractional precipitation with ammonium sulphate. The saline extract of the cake was saturated with ammonium sulphate to 0.25 saturation. The precipitate of Arachin obtained was redissolved in 10 per cent saline solution and was reprecipitated from this either by dilution and acidification or by dialysis. Thus two preparations of this protein were obtained.

Conarachin was obtained by dialysing the filtrate from *Arachin* against cold distilled water to make it free from ammonium sulphate. Another preparation of the same protein was prepared by fully saturating the filtrate with ammonium sulphate. This was redissolved in 10 per cent saline solution and dialysed.

All the above preparations were washed with distilled water several times and dehydrated by washing with graded strengths of alcohol and finally by ether. The preparations were dried in vacuum and powdered to pass through a 100-mesh sieve. All the preparations were light powders, greyish white in colour. Arachin constituted the major part of the total globulins of groundnut, its yield being ten times that of Conarachin.

The table below gives the average of duplicate elementary analyses of the various preparations of the three proteins from each of the two varieties.

TABLE II. *Total Globulins*

Preparation No.	Local			Spanish		
	I	II	III	I	II	III
Method	Dialysis	Dilution	Salt Saturation	Dialysis	Dilution	Salt Saturation
Moisture .	7.60	8.20	7.18	6.80	7.56	5.41
Ash .	0.63	0.70	0.67	0.61	0.69	0.64
*Nitrogen .	17.77	17.74	17.65	17.67	17.62	17.63
*Sulphur .	0.42	0.38	0.43	0.40	0.43	0.39

*Arachin*

Preparation No.	Local		Spanish	
	I	II	I	II
Method	Dilution	Dialysis	Dilution	Dialysis
Moisture .	5.98	7.50	6.08	6.42
Ash .	0.18	0.16	0.22	0.18
*Nitrogen .	17.96	17.88	17.89	17.83
*Sulphur .	0.38	0.40	0.40	0.38

*Cowarachin*

Preparation No.	Local		Spanish	
	I	II	I	II
Method	Dialysis	Salt-Saturation	Dialysis	Salt-Saturation
Moisture .	7.92	9.08	10.02	8.82
Ash .	1.99	1.48	1.96	1.64
*Nitrogen .	16.95	16.97	16.89	16.95
*Sulphur .	0.96	0.99	1.01	0.97

\* On ash and moisture free basis.

*Analysis of Proteins.—*

All the proteins were analysed by Van Slyke's Nitrogen Distribution Method (1911) as modified by Plimmer and Rosedale (1925), Knaggs (1923), Daft (1929) and Thimman (1926). Every step such as acid hydrolysis, precipitation by phosphotungstic acid, etc., has an important bearing on the final results. It was found that 48 hours digestion with 20 per cent hydrochloric acid was sufficient to complete the hydrolysis, i.e., to yield the maximum amount of amino nitrogen. After removal of acid as completely as possible, the amount of solid calcium oxide added to neutralize the slight acidity of hydrolysate and to set free all the amide nitrogen was throughout kept at 3.5 gm per 1000 mg nitrogen. Again, the volume in which the precipitation of the bases by phosphotungstic acid was allowed to take place was 200 c.c. containing 350-360 mg of nitrogen and enough hydrochloric acid to make it exactly normal. Thus the concentrations of calcium chloride, nitrogen and acidity were controlled. The solution was first heated to boiling, to which was added a boiling solution of phosphotungstic acid. When this was done in cold, as described by Van Slyke, the precipitate could not be redissolved by heating on the water-bath.

The following tables give the average of duplicate analyses of the various preparations of the three proteins from each of the varieties.

It will be seen that the analyses of the various preparations of the three proteins from each of the two varieties of cakes agree with one another. This shows that all the preparations of each protein are equally pure. Therefore, the preparations were mixed and used for the separate estimation of arginine, cystine, tyrosine and tryptophane, as described below.

*Arginine*.—This was estimated directly in the acid hydrolysate of protein after removing ammonia by lime according to the method of Plimmer and Rosedale (1925) who have shown that the arginine value as obtained from the basic fraction of Van Slyke's analysis is lower than the true value as obtained by direct estimation in the hydrolysate.

*Cystine*.—Plimmer and Lowndes (1927) have shown that only 40 per cent of the cystine is precipitated by phosphotungstic acid along with the other bases. Hence this was estimated by the Folin and Marenzi (1929) method as modified by Remington (1930).

*Tyrosine and Tryptophane*.—These were estimated according to Folin and Marenzi's micro method (1929) in which tyrosine is separated from tryptophane by the precipitation of the latter by mercuric sulphate.

Table IV gives the percentage of these amino-acids in the two varieties.

TABLE III

*Nitrogen Distribution in Proteins (in Percentage of Total Nitrogen)  
Total Globulins*

Preparation No.	Local			Spanish		
	I	II	III	I	II	III
	Dialysis	Dilution	Salt Satura-tion	Dialysis	Dilution	Salt Satura-tion
Melanin, insol	0.50	0.48	0.58	0.71	0.67	0.60
Melanin, adsorbed by lime	0.92	0.87	1.00	0.49	0.53	0.58
Amide ..	11.47	11.62	10.87	11.87	11.26	12.03
<i>Basic:</i> —						
Arginine ..	22.86	21.98	23.08	21.84	22.17	23.03
Histidine ..	2.23	2.89	2.56	2.47	2.21	2.58
Lysine ..	4.96	5.03	4.59	5.10	4.80	5.40
Cystine ..	0.58	0.59	0.44	0.54	0.60	0.57
<i>Non-Basic:</i> —						
Amino ..	54.49	54.82	55.48	53.77	54.16	53.94
Non-Amino ..	1.92	2.06	1.63	1.93	1.96	2.03
TOTAL ..	99.88	100.34	100.23	98.52	98.38	99.76

Preparation No. .	Local		Spanish	
	I	II	I	II
	Method ..	Dialysis	Dilution	Dialysis
Melanin, insol. ..	0.39	0.29	0.56	0.34
Melanin, adsorbed by lime ..	0.47	0.73	0.29	0.53
Amide ..	11.53	11.03	12.82	10.92
<i>Basic</i> —				
Arginine ..	22.56	22.60	22.37	21.87
Histidine ..	3.13	2.85	3.03	2.89
Lysine ..	4.91	4.38	4.49	4.97
Cystine ..	0.47	0.57	0.47	0.55
<i>Non-Basic</i> —				
Amino ..	55.98	56.06	54.52	55.44
Non-Amino ..	2.07	1.61	1.79	2.29
<b>TOTAL</b> ..	<b>101.53</b>	<b>100.13</b>	<b>100.34</b>	<b>99.80</b>

*Conarachin*

Preparation No	Local		Spanish	
	I	II	I	II
Method	Dialysis	Salt Saturation	Dialysis	Salt Saturation
Melanin, insol.	0.88	1.05	1.36	1.04
Melanin, adsorbed by lime	0.97	0.69	0.98	1.22
Amide	11.93	12.16	11.14	11.12
<i>Basic</i> —				
Arginine	21.87	22.37	23.20	22.76
Histidine	3.01	3.33	3.17	2.89
Lysine	5.80	5.26	4.57	5.11
Cystine	0.93	1.01	1.04	0.98
<i>Non-Basic</i> —				
Amino	51.95	52.02	54.42	53.40
Non-Amino	0.99	1.33	1.62	2.10
TOTAL	98.29	99.21	101.50	101.62

TABLE IV  
(In per cent of Protein)

Amino Acid	Total Globulins				Albumin				Concanavin			
	Local	Spanish	Virginia	Local	Spanish	Virginia	Local	Spanish	Local	Spanish	Local	Spanish
Arginine	12.66	13.26	-	13.96	13.55	13.88	14.10	13.07	-	-	-	-
Histidine	1.67	1.58	-	1.92	1.95	1.88	1.90	1.88	1.88	1.88	1.88	1.88
Lysine	4.49	4.60	-	4.35	4.41	4.98	4.82	4.97	-	-	-	-
Cysteine	-	1.24	1.42	-	1.17	1.17	0.85*	1.57	1.57	1.57	1.57	1.57
Tyrosine	-	5.48	4.80	-	5.69	5.43	5.50†	5.09	5.23	5.23	5.23	5.23
Triptophane	0.58	0.66	-	0.66	0.60	Present	0.93	1.07	-	-	-	-

\* Method of Van Slyke (1911)

† Method of Pohl and Dene (1912)

Local and Spanish varieties—Authors

Virginia variety—Johns and Jones (1916-18)

From the above table the following points may be noted. Compared with Arachin, Conarachin is characterised by its higher contents of basic nitrogen, cystine, tryptophane and lysine. But it is, however, much poorer in tyrosine. On the whole, the proteins of the three varieties of groundnut—Local and Spanish, grown in India and Virginian grown in America—do not show any appreciable difference in their composition. Perhaps the methods of analysis are not sufficiently sensitive to show the subtle varietal differences which may possibly exist.

TABLE V

Amino Acids	1	2	3	4
	Groundnut Cake	Milk	Soyabean	Gram
Arginine .. .	13.26	4.84	5.12	11.85
Histidine .. .	1.58	2.59	1.39	1.42
Lysine .. .	4.69	5.95	2.71	7.42
Cystine .. .	1.42	1.20	†	2.02
Tyrosine .. .	4.80	4.50	1.86	2.95
Tryptophane .. .	0.66	1.50	†	0.46

1 Spanish variety, Total Globulins—Authors.

2 and 3 'Soyabean'—Piper and Morse (1923).

4 Niyogi, *et al.*, 1932.

In the above table, the total globulins of groundnut are compared with the total proteins of milk, of soyabean and of gram (*Cicer arietinum*). Groundnut protein is characterised by its higher content of tyrosine and arginine—two of the most important essential amino acids. Though slightly poor in lysine content, it is better than soya-bean protein. In its histidine content, groundnut protein is quite comparable to the proteins of soyabeans and *Cicer arietinum*.

Since milk proteins are considered to be perfect and complete, an approximate value of other proteins in animal nutrition may be obtained by comparing their compositions with that of milk proteins. The table clearly points to the conclusion that the proteins of groundnut compare more favourably with milk proteins than those of soyabeans. Thus as

revealed by chemical analysis, groundnut proteins are, in some respects, superior to soyabean proteins

Daniel and Loughlin (1918) have conducted feeding experiments with groundnut cake. According to them the proteins of groundnut are comparable to those of soyabean. Both these legumes supply the essential amino-acids in sufficient amounts for normal growth and reproduction. Groundnuts are lacking in the fat soluble accessory, but contain a considerable amount of the water-soluble vitamins. Rats fed on a mixture of groundnut meal furnishing 18 per cent of protein, 5 per cent of butter fat, lard and corn starch to which no minerals were added, grew normally, reproduced and successfully reared their young. Again, Wallis (1917) also states that groundnut contains a very complete protein (cf. Press Note of the Industries Department of the Bombay Government, 1918). Groundnut cake, milk powder and sodium bicarbonate mixed in the proportions of 94 : 5 : 1, yield a very good invalid food. Such a mixture has been successfully used for the preparation of culture media. Mixed with egg white, it can be used as a diabetic food. Wheat flour when mixed with groundnut cake in the proportion of 3 : 1 is said to make as ideal a bread as possible and can be used for the preparation of "Rotis". Johnus and Finks (1920) have made similar observations. Bread made from 25 parts groundnut flour and 75 parts wheat flour furnished adequate proteins for normal growth. Proteins of this bread were utilised twice as well as those of bread made of wheat only.

Further, groundnut is easily available in this country where it has become native for a long time and its method of cultivation is well known to the farmers. It can be grown even under dry-farming conditions in sandy soil. It can be eaten raw or cooked and can be introduced into the daily diet of the masses without much difficulty. As regards the keeping quality of groundnut cake, Sahasrabuddhe and Bhatt (1935, private communication) have shown that it can be kept for a long time if the flakes of the fresh-cake are heated to 100° C and placed in airtight containers.

On the other hand, soyabean is an exotic to this country and is only now being introduced. It cannot be digested raw and has to be cooked under pressure. Soyabean flour prepared in the ordinary way does not keep well and gets rancid easily. A well-keeping flour can be prepared by patented methods only which are not in the easy reach of poor cultivators. The bean possesses a slight bitterness for which a taste must be cultivated. Its flour when added to bread to improve the colour of the crumb has been found to destroy the Vitamin A content of the bread. Similar tests with cod-liver oil have shown that soya flour destroys nearly the whole of the

Vitamin A of the oil (*Scientific American*, February 1937, p. 104). Recently Dr W. R. Aykroyd, Director of Nutrition Research, Indian Research Fund Association, addressing the Rotary Club of Calcutta, struck a note of warning against the extended use of soyabean which is now being popularised. In his opinion, it remained to be shown by conclusive experiments whether the soyabean had any particular advantage over the common pulses (*Times of India*, December 5, 1936).

From the above it will be seen that groundnut is in no way inferior to soyabean. In some respects it has certain advantages over the latter. It is therefore advisable to investigate fully the possibilities of the easily available groundnut before advocating the use of soyabean in India.

The authors are greatly indebted to Rao Bahadur Dr. D. L. Sahasrabuddhe for his keen interest and advice during the course of this investigation.

#### REFERENCES

Daft, *Biochem. J.*, 1929, 23, 149  
 Daniel and Loughlin, *J. Biol. Chem.*, 1918, 33, 295  
 Daniel and Menaul, *ibid.*, 1921, 46, 437  
 Folin and Marenzi, *ibid.*, 1929, 83, 89, 102.  
 Johns and Jones, *ibid.*, 1910, 28, 77  
 \_\_\_\_\_, *ibid.*, 1917, 30, 33  
 \_\_\_\_\_, *ibid.*, 1918, 36, 491  
 Johns and Finks, *ibid.*, 1920, 42, 569  
 Knaggs, *Biochem. J.*, 1923, 17, 488  
 Narayana and Sreenivasayya, *J. Ind. Inst. Sci.*, 1936, 19A, 1  
 Nyogi et al., *Ind. Jour. Med. Res.*, 1932, 19, 1054  
 Piper and Morse, *Soyabean*, McGraw Hill Book Co., N.Y., 1923.  
 Plummer and Roedale, *Biochem. J.*, 1925, 19, 1004  
 Plummer and Lowndes, *ibid.*, 1927, 21, 245  
 Remington, *ibid.*, 1930, 24, 1114.  
 Thimman, *ibid.*, 1926, 20, 1190  
 Van Slyke, *J. Biol. Chem.*, 1911, 10, 15  
 Wallis, *Ind. Jour. Med. Res.*, 1917, 4, 86.

## ON SOME NEW FORMS OF BATRACHIA FROM S. INDIA.

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Received November 29, 1937

THE material which forms the subject of this paper has been collected during the excursions of the members of the Staff and students of the Department of Zoology, University of Mysore, for over six years. The primary object of these excursions has been to study the distribution of anuran fauna in the localities explored, their mode of occurrence and their general habits. We selected certain types of areas marked by distinctive physical features, and the results of our collections and of the field observations are set forth in these notes.

The Malnad areas of Kadur, Hassan and Shimoga Districts of Mysore, the Anamalai Hills of Coimbatore and Perambikulam forests of Cochin were selected for making collections, and for purposes of comparison, specimens were also taken from the plains. The chain of hills separating Mysore from Malabar and North and South Canara forms a definite landmark and well-wooded gigantic spurs, divided by innumerable water courses, extend on either side. The base of these elevated regions is clothed with dense vegetation and harbours pools formed by waterfalls. The soil is composed almost entirely of laterite, and there are places in the Malnad where rich loam and stiff clay are also encountered. During the monsoon these areas receive abundance of rain which inundates the country swelling the small mountain streams into torrents and converting the shallow puddles into dangerous pools. Perambikulam is in the heart of Cochin forests and the summit of the hills are covered by grass, being elevated above the forest belt. Below the hills are covered by bamboos and forest trees which abound in big game. Anamalai Hills are clothed by impenetrable forests and even the base of the Hills could only be approached with difficulty. On account of scanty rainfall in the lower regions, the ground is hard and water scarce. During the hot weather all these areas become parched and they seem scarcely to harbour any life. The most favourable time for collection of batrachian specimens is sometime after the rains, when cryptozoic forms will not have retreated into their burrows, but will be found lurking near about the water margins.

The distribution and occurrence of batrachian fauna in the Maidan, semi-Malnad areas and the ghats which present striking physical features, present a sharp contrast. The main point of interest is that each of these

areas is inhabited by groups which hardly extend into the adjacent territory. For instance, typical Māidan forms like *R. hexadactyla*, *R. higrina*, *R. limnocharis* and *R. breviceps* are not encountered in the ghats and the only exception is *R. cyanophlyctis* which can be taken in the thickest forest and on the tops of the highest peaks. *Bufo melanostictus* follows *R. cyanophlyctis* in its occurrence; but *Bufo fergusoni* does not extend beyond the semi-Mānad area into the ghats. Again *Rhacophorus maculatus* is strictly an inhabitant of the plains and is rarely met with in the ghats, though a few specimens of this species have been collected from the semi-Mānad areas. We can scarcely draw a line of demarcation between strictly Mānad and semi-Mānad in regard to the distribution of forms such as *Rana curtipes*, *Rana malabarica*, *R. leptodactyla*, *R. aurantiaca*, *R. dobsoni*, *R. verrucosa*, *R. rufescens*, *R. beddomii* and similarly with regard to *Rhacophorus malabaricus*, *Rh. lateralis* and *Bufo pulcher*, *Bufo microtympanum*, *B. parvulus*. When we enter the forests which clothe the sides of the hills, we discover that these forms are replaced by a bewildering wealth of batrachian life belonging to the genera *Ixalus* (*Philautus*) and *Microhyla* and *Nyctibatrachus*, and *R. cyanophlyctis* and *Bufo melanostictus*, however, freely mingling with them. Except for a few stray forms of *Ixalus* and *Microhyla* occasionally found in the semi-Mānad areas, the three genera *Ixalus* and *Microhyla* and *Nyctibatrachus* may be described as inhabitants of mountain slopes, but they rarely extend to the summits where only *R. cyanophlyctis* and *Bufo melanostictus* occur.

Among the *Engystomatidae*, the most hardy creature is *Microhyla ornata*, which in point of distribution follows *R. cyanophlyctis* and *Bufo melanostictus*, though it may not be found on the summit of the hills. *Cacopus* (*Euphlyctis*) *systoma*, *M. rubra*, *Callula* (*Ramanella*) *variegata*, *C. (R.) triangularis* and *C. (R.) obscura* are strictly confined to the plains. But the semi-Mānad zone in Mysore harbours a race of *Callula* (*Ramanella*) whose adaptive modifications to the peculiarities of the environment affected by them, differ so strikingly from their congeners in the plains, that one would be justified in creating a separate genus for the reception of these forms. They are all as a rule short-legged, the tibio-tarsal articulation hardly extending beyond the middle of the body, and they have no web, and all possess the power of climbing the smooth surfaces of the plantain trees. Our collection of *Engystomatid* forms from the plain districts is sufficiently rich and varied and a comparison of the semi-Mānad specimens with them indicates the lines along which divergence has proceeded. It is to be noted that none of these *Engystomatidae* are met with in the thickly wooded places on the slopes of hills, and their favourite haunt is the cordamom plantation and plantain gardens behind the village houses of the Mānad areas.

The hill slopes which receive an annual rainfall of over 200 inches, are cut up by mountain torrents during the monsoon, but become dry and parched during the hot weather. The Batrachians [*Ixalus (Philautus)*, *Microsaurus* and a few members of *Rhacophorus* and *Nyctibatrachus*] affecting these slopes, are scarcely met with in water, but are always found clinging to the rocks, loose stones and tree trunks in the close vicinity of waterfalls, running brooks and sheltered pools. They have large digital expansions, and excessively long hind limbs and slender bodies (the only exception being *Nyctibatrachus*), enabling them to cling to hard surfaces when their homes are flooded, or to swim against the rapid current. *R. cyanophlyctis* is a thoroughly aquatic form, but it has a habit of burrowing in sands under water and also of skimming over the surfaces of water by taking a series of short leaps, enabling the frog to escape being carried away by the floods. *B. melanostictus* is capable of burrowing or entering narrow crevices and fissures of rocks, where they abide till the disturbances of weather are over. In consequence of these peculiar habits of life, they do not seem to have developed any anatomical peculiarities different from the other members of their tribe living in the plains. Both these forms are known to traverse fairly long distances and it is this habit which has brought them to the Malnad areas and hill tracts.

The larvae of some species of *Ixalus* in our collection almost as a rule have greatly enlarged lips, which, we have noticed, they use in adhering to rocks and water weeds. They are sufficiently large to act as floats in case the tadpoles are caught in the rapids. It is well known that the tadpoles of *Nyctibatrachus* possess widely expanded lips which are suctorial. In the case of *R. cyanophlyctis* and *Bufo melanostictus*, the tadpoles have powerful caudal fins, far more powerfully developed than in the forms found in the plains and their pouting lips are provided with enlarged beaks and teeth. As a rule, they attain a larger size, far larger than tadpoles of corresponding age in the plains. In the semi Malnad belt, where a large number of species of *Rana*, *Rhacophorus* and *Bufo* occur, their larvae differ in the degree of development of fin, size of tail and in the mouth parts, which, while affording diagnostic characters, are nevertheless features due mainly to adaptive modifications. I have had occasion to study the larval forms of anura taken from all these areas, and I am of opinion that, while those coming from similar situations present fairly uniform characteristics capable of being used for taxonomic purposes, on the other hand, specimens of tadpoles belonging to the same species, but taken from situations differing widely in regard to the condition of the country and rainfall, show corresponding differences in external morphology. I shall refer below to a few instances of such divergent modifications and point out how the mouth parts alone, such as the beaks

and horny denticles, are not affected by the change of environment and how they can be used for the purpose of relating the tadpoles to their parents on the basis of these characteristics. Appropriate comments on the tadpoles will be made under the respective heads.

In the descriptive part, I have followed the nomenclature introduced by Parker, Boulenger and Malcolm Smith.

Recently, a number of emended schemes of classification of the group *Anura* have been put forward, based upon osteological characters. The family engystomatidae under the changed nomenclature is treated as Microhylidae by H. W. Parker<sup>1</sup> and the names of the Indian genera formerly included in the group have also undergone alteration. Thus *Cacopus* becomes *Uperodon*—*Callula* is now recognised as *Ramanella*, while *Microhyla* and *Melanobatrachus* are permitted to retain their names unaffected. In S. India, the family Microhylidae is chiefly represented by the following four genera (1) *Uperodon*, (2) *Ramanella*, (3) *Microhyla* and (4) *Melanobatrachus*. As the first two names are not quite familiar yet and as it is not widely understood what they replace, I have used the names still in common use in the introduction and in the systematic description. I have adopted the new nomenclature. Similarly with regard to *Ixalus*.

In 1920, Boulenger<sup>2</sup> revised the genus *Rana*, making use of several characters neglected for a long time and he pointed out that the recognition of their value for taxonomic purposes might lead to a better understanding of their inter-relationships. He has divided the genus *Rana* into 9 sub-genera of which species belonging to sub-genera *Rana*, *Tomopterna*, *Discodèles* and *Hylorana* are represented in South India.

In 1930, Dr. Malcolm Smith<sup>3</sup> pointed out that, as the name *Ixalus* was preoccupied, this genus should in accordance with the accepted terms of nomenclature be known as *Philautus* and he does not favour the fusion of this genus with *Rhacophorus* whose generic position is still left intact. He retains the name *Rhacophorus* in preference to *Polypedates*.

In the following descriptive accounts, I have used the names suggested by these authors.

<sup>1</sup> "Frogs of the Family Microhylidae," H. W. Parker, B.A., *British Mus. Nat. Hist.*, 1934.

<sup>2</sup> "A Monograph of the South Asian, Papuan, Melanesian and Australian Frogs of the Genus *Rana*," G. A. Boulenger, *Rec. Ind. Mus.*, Vol. XX, June 1920.

<sup>3</sup> "The Reptilia and Amphibia of Malaya Peninsula," Malcolm A. Smith, *Bull. Raffles Museum, Singapore*, 1930.

## Genus RANA

## Sub-Genus TOMOPTERNA

*Rana parambikulamana* n. sp.

Vomerine teeth in short oblique series, commencing from the inner angle of the choanae which are transverse, oval and directed inwards. The hinder margin of the vomerine teeth, extending far beyond the posterior border of the choanae. Tongue without a median papilla. The lower jaw with three tooth-like conical prominences.

Head convex, distinctly longer than broad, snout pointed, projecting beyond the lower jaw, longer than the eye, canthus rostralis obtuse, loreal region slightly oblique, concave, nostril nearer to eye than to the tip of the snout, distance between the nostrils greater than the interorbital width which is  $2/3$  that of the upper eye-lid, tympanum indistinct,  $2/5$  the diameter of the eye, equal to its distance from the eye.

Fingers rather short, obtuse, first nearly equal to the 2nd, both considerably shorter than the third which is distinctly longer than the snout. The first finger with a larger basal pad, subarticular tubercles prominent, rounded, fourth digit equals the diameter of the eye.

Hind limb long, the tibio-tarsal articulation reaching far beyond the tip of the snout, the heels crossing when the limbs are folded at right angles to the body, tibia more than three times as long as broad and 2 and  $1/6$  times in the length from snout to vent, slightly longer than the fore limb and longer than tarsus, fourth toe considerably longer than the thigh or tibia and slightly more than twice the length of the tarsus. All the toes are long, obtuse,  $\frac{1}{2}$  webbed, outer metatarsals united, subarticular tubercles small, not conspicuous, inner metatarsal tubercle large smaller than the first toe, inserted almost in line with the latter, outer metatarsal tubercle inconspicuous.

Skin smooth above and on the ventral surface, a fold from the hinder angle of the eye, extending over the tympanum to the shoulder, two short prominent folds from behind the eye directed slightly inwards, and a pair of less conspicuous folds on either side of the dorsal band, also starting from the upper margin of the eyelids, two short obliquely set folds forming an open U-shaped space on the back behind the shoulder as in *R. verrucosa* and *R. rufescens*. No granulation on any part of the body.

Yellowish brown above, deeper over the head and the shoulder, with a light vertebral band; cutaneous folds edged black, a broad white band from the eye to shoulder beneath the fold, lips with alternate light and dark bands; sides of body and ventral surface white; limbs barred, the stripes of the thighs oblique.

Male with two large vocal sacs, black confluent medially  
A single specimen. Type in the Central College Museum  
*Habitat* —Parambikulam forests, Cochin State, S India.

## MEASUREMENTS.

	mm.		mm.
From snout to vent	. 39.00	Fore arm	.. .. 8.00
Head ..	. 15.00	Arm ..	.. .. 9.00
Width of head ..	. 11.50	First finger ..	.. .. 6.00
Snout ..	. 6.50	Second finger ..	.. .. 5.75
Eye ..	. 5.00	Third finger ..	.. .. 2.50
Upper eyelid ..	. 3.00	Fourth finger ..	.. .. 5.00
Interorbital width ..	. 2.00	Thigh ..	.. .. 16.50
Distance between eye and nostril ..	. 2.50	Width of thigh ..	.. .. 8.00
Distance between nostril and tip of snout ..	. 3.00	Tibia ..	.. .. 18.00
Distance between nostrils ..	. 3.00	Width of tibia ..	.. .. 5.00
Distance between eye and tympanum ..	. 2.00	First toe ..	.. .. 5.00
Tympanum ..	. 2.00	Second toe ..	.. .. 9.00
Internarial distance ..	. 3.00	Third toe ..	.. .. 15.50
		Fourth toe ..	.. .. 23.50
		Fifth toe ..	.. .. 14.00
		Inner metatarsal tubercle ..	.. .. 3.00

I have compared this specimen with *R. rufescens*, *R. breviceps* and *R. dobsoni*, from which it differs almost in every character, and generally resembles *R. tigrina* in external form, though differing in details both from this species and *R. limnocharis* (*Forma typica*) through which this new species is derivable.

*Rana leucorhynchus*, n. sp.

Vomerine teeth in strong oblique series, separated only by a narrow median gap in the median line, but well separated from the choana. The choanae are oval orifices, transversely situated. Behind the vomerine teeth, a well-marked broad triangular palatine cavity. Tongue without a median papillæ, lower jaw with a single tooth-like prominence.

Head narrowly concave, slightly longer than broad; snout obtuse, truncated, projecting beyond the mouth, very slightly longer than the eye; canthus rostralis angular, loreal region oblique, broadly concave, nostril slightly nearer the tip of the snout than the eye, distance between the nostrils equals the inter-orbital width, upper eye lid  $\frac{1}{3}$  in the distance between the eyes; tympanum distinct, smaller than the diameter of the eye,  $3\frac{1}{2}$  times its distance from the latter.

Fingers rather long, pointed, first longer than the second, shorter than the third which is longer than the snout; the first exactly equals the snout and twice the width of the upper eyelid. Subarticular tubercles large and conical, tubercular pads at the base of the first and third finger.

Hind limbs short, the tibio-tarsal articulation reaching the eye, the heel separated when the limbs are folded at right angles to the body, tibia just as long as the thigh, length of tibia more than 3 times in the length of thigh, more than 2 times in its length, width of thigh equals the length of the snout and the latter  $1\frac{1}{2}$  times the width of tibia. The length of the hind limb (without digits) exceeds very slightly the distance between the snout and vent. Tarsus two times the width of tibia, toes rather short,  $\frac{1}{2}$  webbed, outer metatarsals united, subarticular tubercles small, inconspicuous, an inner tarsal fold present, inner metatarsal tubercle large, strongly compressed with a sharp edge, as long as the first toe, at the base of which it is obliquely set. A small outer metatarsal tubercle at the base of the fourth toe.

Dorsal surface of the body, throat and chest smooth, abdomen and sides of the body and lower surface and posterior margin of thighs granular, sides of body with interrupted folds, no fold from eye to the shoulder.

Pale brown above; upper surface of snout including the anterior third of the upper eyelid with a distinct broad white band. Between the eyes a dark W-shaped band, behind the eyes and shoulders white; proceeding from behind the eyes, a long oval brown mark with a pair of darker streaks running along the median line of the area. A similar mark U-shaped in the median line of the back, and U-shaped marks on the sides of the body and hip, fore arms and legs cross barred, throat and abdomen white, under-surface of thighs yellow; loral region with a dark broad band from the eye to the snout, below the eye a dark anvil-like mark, the lower lid being white, foot chocolate brown.

MEASUREMENTS

	mm.		mm.
Snout to vent	35.00	Arm	7.00
Head	18.50	First finger	6.00
Width of head	15.00	Second finger	5.00
Snout	6.00	Third finger	7.50
Eye	5.50	Fourth finger	4.75
Distance between eye and nostril	2.50	Thigh	13.50
Distance between nostril and snout	3.00	Tibia	13.50
Distance between nostrils	4.00	Tarsus	8.00
		Width of thigh	6.00
		Width of tibia	4.00

	mm.		mm.
Distance between eye and tympanum ..	1.00	Inner metatarsal tubercle ..	3.50
Interorbital width ..	4.00	First toe ..	3.50
Upper eyelid ..	3.00	Second toe ..	6.00
Tympanum ..	3.50	Third toe ..	10.00
Fore arm ..	7.25	Fourth toe ..	14.00
		Fifth toe ..	8.50
		Toes ♀ webbed	

(Linked with *Rana tigrina* through var *pantherina* resembling *R. cancrivora* at least superficially)

*Habitat* —Wattakole, Coorg, South India

A single specimen

*Type* —Central College, Bangalore

*Sub-Genus HYLORANA*

*Rana intermedius*, n. sp

Vomerine teeth in short oblique groups, arising far from choana, the distance between them and the latter is greater than the median gap separating the series of teeth. The internal bulging of the eyes into the mouth forms two large massive structures almost meeting in the median line posterior to the vomerine teeth. Tongue very large and broad, without a papilla.

Head strongly depressed, much longer than its width, snout acutely pointed, projecting well beyond the mouth, far longer than the diameter of the eye, canthus rostralis very distinct, loreal region nearly horizontal, deeply concave, nostril nearer to the tip of the snout than the eye; distance between the nostrils equal to the interorbital width, upper eyelid four-fifths in the distance between the eyes, tympanum (that on the right side is larger than the left) very distinct, smaller than the eye, separated from the latter by a distance equal to  $\frac{1}{2}$  or  $\frac{2}{3}$  its own diameter.

Fingers rather slender, terminating in discs, first longer than the second, third slightly longer than the snout, subarticular tubercles fairly large, round, a distinct horse-shoe shaped groove separating the upper half of the disc from the lower.

Hind limb rather long, tibio-tarsal articulation reaching the nostrils, heels completely overlapping when the legs are folded at right angles to the body, tibia nearly five times as long as broad and more than half the total length of the body, very greatly exceeding the fore limb, toes moderately

long, the tips dilated into discs, all bearing more or less well defined grooves, discs broader than long, web extending to the tips of all the toes except the fourth and fifth, subarticular tubercles conical, not conspicuous, outer metatarsal tubercle 2/5 in the length of the first toe, a small conical outer metatarsal tubercle at the base of the fourth toe

Skin smooth, a narrow dorso-lateral glandular fold from the eye to the groin, no fold from the eye to the tympanum to the shoulder, the distance between the dorso-lateral folds across the iliac bones is more than 1/5 in the total length of the body, humeral gland No parotoids

Uniform brown above, sides darker, limbs faintly cross-barred, abdomen and throat with irregular dark areas, under surface of thighs yellowish, posterior and anterior border of the thighs mottled

*Habitat*—Saklespur, Hassan District, Mysore State

*Type*—Central College, Bangalore

The species is intermediate between *R. gracilis* and *R. temporalis*, both of which occur in the same locality with *R. malabarica*, *R. aurantiaca* and *R. curtipes*. I have compared this with these members in almost every particular and I am convinced that though it resembles with one or the other of these species in certain characters, it is entitled to the rank of a separate species. This course is justifiable when the size and proportions of the head, the relative size of the tympanum and the eye are taken into account

MEASUREMENTS.

	mm.		mm.
Snout to vent	43.00	First finger	7.00
Head	20.00	Second finger	6.00
Width of head	13.50	Third finger	9.00
Snout	8.50	Fourth finger	7.50
From nostril to tip of snout	4.00	Thigh	20.00
Tympanum	4.00	Width of thigh	5.00
Eye	0.50	Inner metatarsal tubercle	2.00
From eye to nostril	4.75	Tarsus	12.00
Internarial width	5.00	Width of tarsus	3.00
Interorbital width	5.00	First toe	5.00
Upper eyelid	4.00	Second toe	8.50
Distance between tympanum and eye	1.00	Third toe	13.50
Arm	9.00	Fourth toe	20.00
Fore arm	8.25	Fifth toe	15.00

*Rana sauriceps*, n. sp

Vomerine teeth in short oblique series, arising from the anterior border of the choanae which are horizontal and extending slightly beyond the hinder border of the latter. Lower jaw with a small median and two lateral median tooth-like prominences. Tongue very small, without a papilla.

Head somewhat elongated, convex, longer than broad; snout acutely pointed, truncated projecting beyond the mouth, twice as long as the diameter of the eye, with a triangular or diamond-shaped pit on the terminal dorsal aspect. Behind this rostral pit there are two oval elevations with a median transverse groove and both separated from a similar elevation between the eyes. These elevations look not unlike the cephalic shields of the Saurians, behind the tip of the snout there is a pit on each side, separated by a ventral bar from the loreal region which is almost horizontal and concave, canthus rostralis obtuse, nostrils nearer to the snout than to the eye, distance between the nostrils equal to the interorbital width, tympanum slightly smaller than the eye, the latter is twice the distance between it and the former and this distance is equal to the upper eyelid.

Fingers moderate, obtuse without discs, first finger equal to the second, about  $\frac{1}{3}$  in the length of the snout, third as long as the snout, the fourth longer than the first or second and is equal to the arm. Subarticular tubercles conical, moderate.

Hind limbs rather long, slender, tibio-tarsal articulation reaching the nostril, heels very slightly overlapping when the legs are folded at right angles to the axis of the body, thigh nearly twice its width, tibia three times its width, but considerably less than half the total length of the body and slightly shorter than the third toe equal to the fifth, toes pointed, web not extending to the tip of the first phalangeal bone, rather stopping at the base, outer metatarsals not united, separated by web, inner metatarsal tubercle about  $\frac{1}{2}$  the diameter of the eye, a small rounded outer metatarsal tubercle at the base of the fourth toe; subarticular tubercles small, inconspicuous.

Upper surface of the skin slightly granulate, with short interrupted longitudinal folds with a few tubercles, curved temporal fold is present, in preserved specimens, the abdomen is thrown into fine transverse folds; throat, abdomen and undersurface of the thighs free from granulation. No dorso-lateral glandular fold. A  $\cap$ -shaped mark found on the back behind the shoulders.

Above chocolate red, sides darker, fore arm and tarsus faintly barred; throat and chest yellowish, abdomen brown, the under surface of the thighs

pale orange; upper lip with vertical bands, two of which enter the eye, the lower lip with dark and white spots or bars

## MEASUREMENTS.

	mm.		mm.
Snout to vent	30.00	Third finger	6.00
Tympanum	2.50	Fourth finger	5.00
Eye	3.00	Thigh	10.50
Distance between eye and tympanum	1.50	Width of thigh	5.00
Head	10.50	Tibia	12.00
Width of head	8.75	Width of tibia	4.00
Snout	6.00	Tarsus	9.00
From nostril to snout	2.25	Width of tarsus	3.00
From eye to nostril	3.00	Inner metatarsal tubercle	1.50
Interorbital width	3.75	First toe	4.00
Upper eyelid	1.50	Second toe	8.00
Fore arm	6.50	Third toe	11.00
Arm	5.00	Fourth toe	15.00
First finger	4.00	Fifth toe	10.00
Second finger	4.00	Outer metatarsal tubercle	0.75

*Habitat*—Wattekole, Coorg, S. India

A number of specimens of all ages

*Type*—Central College, Bangalore

## Sub-Genus DISCODELES.

*Rana tenuilingua* n. sp.

Vomerine teeth in short, transverse or slightly oblique series, arising separately from the choanae, almost on a level with the posterior border of the latter, the space between the rows exceeds the distance between the teeth and the choanae, tongue very thin, small, the posterior notch sometimes very broad, a small conical retractile papilla in the middle; the lower jaw is covered by the upper throughout and the maxillary teeth are large, without tooth-like prominence on the lower jaw

Head broadly convex, slightly wider than long; snout obtuse projecting beyond the mouth, as long as or slightly longer than the eye, canthus rostralis obtuse; loreal region oblique concave, nostril nearer to the tip of the snout than the eye; interorbital width more than twice the distance between the nostrils and is equal to the upper eyelid, tympanum  $\frac{1}{2}$  in the diameter of the eye and twice the distance between it and the latter, the frontoparietal region is excavated into a flask-like depression, occasionally traversed by a median groove and this area is further separated by a transverse groove from the nasal region where the fold of skin is elevated into scale-like patches

separated by a median groove corresponding with the bony sutures. The nasal "scales" are divided from the premaxillary area by a transverse groove, with a ridge of skin in front, connecting the nostrils. The markings on the head are exactly like those of *Rana sauriceps* but without the rostral pit.

Fingers are short, tips dilated into truncated discs which are broader than long, the groove in front of the discs is either absent or faintly marked in some, first finger as long as or very slightly longer than the second, third slightly longer than the snout, subarticular tubercles very faintly visible.

Hind limb long, tibio-tarsal articulation reaching nostril or tip of snout, the heels strongly overlap when the limbs are folded at right angles to the body, tibia about 5 times as long as broad, thigh half in the length of the body from snout to vent and longer than the fore limb, toes rather short, discs truncated, broader than long, the groove indistinct, web extending to nearly three-fourths of the first phalanx of fourth toe and second of fifth and third, web not extending to the base of the outer metatarsals, subarticular tubercles inconspicuous, inner metatarsal tubercle feebly developed, a very small outer metatarsal tubercle at the base of the fourth toe. Phalanges T-shaped.

Upper surface of skin with a series of interrupted fine folds, about 6 to 7 longitudinal series can be made out, throat and chest and undersurface of thighs smooth, abdomen finely granulate. No temporal fold.

Pale brown above, sides darker, upper surface of snout white; a dark band from the tip of the snout extending through the loreal region, and below the eyes, surrounds the tympanum, a short dark band from the tympanum to the shoulder, lower jaw with dark and white longitudinal bars, fore arm and fingers and hind limbs barred, throat pale yellow, abdomen white, undersurface of thighs reddish.

#### MEASUREMENTS

	mm		mm
From snout to vent	23.00	First finger	2.25
Tympanum	2.00	Second finger	2.00
Eye	3.00	Third finger	4.00
Distance between tympanum and eye	1.00	Fourth finger	2.50
Snout	3.50	Thigh	11.00
Distance between eye and nostril	2.25	Width of thigh	3.50
Distance between nostril and snout	1.25	Tibia	13.50
Distance between nostrils	2.75	Width of tibia	2.50
Distance between eyes	3.00	Tarsus	7.50
Upper eyelid	1.25	Width of tarsus	1.50
		Inner metatarsal tubercle	1.00
		First toe	2.50
		Second toe	5.00

	mm.		mm.
Head . .	9.00	Third toe . .	7.50
Width of head . .	9.25	Fourth toe . .	12.00
Fore limb : Arm . .	1.75	Fifth toe . .	7.00
Fore arm . .	4.75	Outer metatarsal tubercle . .	

*Habitat* — Kemphole Ghat, Hassan, Mysore, South India

*Type* — Central College, Bangalore

*Genus NYCTIBATRACHUS*

*Nyctibatrachus tytleri*, n. sp.

Vomerine teeth in two oblique rows, short series set well behind the choanae

Head as long as broad or slightly longer, depressed, projecting beyond the mouth. Snout not longer than the diameter of the eye. Inter-orbital space equals the width of the upper eyelid. Eyes lateral. Distance between the eye and the nostril equals  $2\frac{1}{2}$  times the internarial width. Tympanum partially or completely hidden. Supra tympanic fold present. Habit not stout. Canthus rostralis angular, well marked. Loreal region concave, oblique. Fingers dilated into small discs. First finger shorter than second. Third longer than snout. Subarticular tubercles moderately large, not very prominent.

Hind limbs long. The tibio-tarsal articulation reaching the posterior angle of the eye. Heels touch when the limbs are folded at right angles to the body. Thigh more than  $1\frac{1}{2}$  times longer than its width and tibia more than three times its width, and less than half in the length of the body. Toes with tips swollen into small discs, more than half-webbed. Sub-articular tubercles moderately developed. Inner metatarsal tubercle elongate, spade-like, more than half the length of the first toe, connected by web to the base of the first toe. No outer metatarsal tubercle.

Skin above with tubercles and short glandular folds. Thighs free. Tibia, tarsus and fifth toe distinctly granular. Arms feebly granular. Lower surface including the limbs perfectly smooth.

Brown above. Both limbs with faint cross bars. Ventral surface including the limbs whitish. Lower jaw with feeble white bars on a pale background of brown.

MEASUREMENTS.

	mm.		mm.
Diameter of the eye	4.00	Fourth finger	6.75
Snout . .	6.00	Thigh . .	17.75
Interorbital space . .	4.00	Thigh thickness . .	10.00
Upper eyelid . .	4.00	Shank . .	19.00

	mm.		mm.
Internarial width	2.00	Shank thickness ..	6.50
Distance between eye and nostril	5.00	Tarsus ..	11.00
Arm ..	6.75	Tarsus thickness ..	3.50
Arm thickness ..	3.50	First toe ..	5.00
Fore arm ..	8.75	Second toe ..	8.00
Fore arm thickness ..	4.00	Third toe ..	12.00
First finger ..	5.00	Fourth toe ..	17.00
Second finger ..	6.00	Fifth toe ..	12.00
Third finger ..	8.00	Inner metatarsal tubercle ..	2.75
		Total length ..	42.00

*Habitat*—Forests of Kemphole, Saklespur, Hassan, Mysore

*Type*—Central College, Bangalore

*Remarks*—There are three species of *Nyctibatrachus* known, viz., *N. pygmaeus*, *N. major* and *N. sanctipalustris*, of which we have a fairly good collection. The new species differs from the known forms in certain fundamental respects and might even appear as a connecting link with *Rana*. For instance, the tympanum is only partially hidden, the eyes are laterally directed and the body is not stout and a canthus rostralis is present. With the exception of these characteristics, its resemblance to the other species of *Nyctibatrachus* is fairly intimate. The configuration of the body and limbs of this species may perhaps be an expression of convergence towards the *Ixalus* type, evoked by the peculiar mode of life and the character of the surroundings.

#### GENUS NANNOBATRACHUS

According to Boulenger this genus is distinguished from *Nannophrys* by the character of the pupil which is vertical in *Nannobatrachus* and horizontal in *Nannophrys*. To this difference he adds a few other factors which make the two genera sufficiently distinctive. The species with which the following notes deal, possesses a horizontal pupil more or less red even in the preserved state and agrees with the other characteristics of *Nannobatrachus*. The pupil of the eye in the living specimen bears no resemblance to its shape in the preserved forms, undergoing some amount of distortion on account of contraction. This is rather an uncertain character for the purpose of differentiating genera, and even as a specific character it is not reliable. No importance is attached to this character in these notes. I have not examined *Nannophrys* and am therefore unable to suggest how far we may depend upon characters like the outer metatarsals being united at the base or separated by grooves for separating *Nannophrys* from *Nannobatrachus*.

Boulenger has described only two species of the former and one of

the latter and for purposes of the example described below, I shall adopt his classification

*Nannobatrachus kempholeyensis*, n. sp

Pupil red, horizontal, habit fairly stout, toad-like. Vomerine teeth in two short series almost transverse arising from the anterior border of the choanae, with a wide gap between, which is nearly twice as wide as the series of the teeth. Tongue moderate, nicked deeply behind, bearing in front a large spherical elevation, clearly marked off from the rest of the tongue and surrounded by a pit, whether a retractile papilla is present is doubtful. Lower jaw without a tooth-like projection

Head flat, wider than longer, snout rounded, scarcely projecting beyond the mouth, as long as the interorbital width or twice the diameter of the eye; canthus rostralis indistinct, loreal region either nearly vertical or slightly oblique, concave in the latter case, nostril equidistant between the eye and the tip of the snout, distance between the nostrils  $\frac{1}{2}$  in the interorbital width, upper eyelid equal to the distance between the nostrils or the diameter of the eye, tympanum indistinct or absent

Fingers rather small, first obtuse, second, third and fourth with ends dilated at the tips into spherical balls not very distinct from the ultimate phalangeal bone; first finger slightly smaller than the second, subarticular tubercles distinct on the third and the fourth fingers only

Hind limb short, the tibio-tarsal articulation not reaching quite the eye but only its posterior border, the heels do not meet when the limbs are folded at right angles to the body, thigh as long as the fourth toe and less than twice its own length, tibia more than twice the length of the snout and slightly exceeds the width of the head, tarsus quite as long as the thigh and more than twice its own width, tips of toes dilated into small discs, not distinct on the first, discs as long as broad, second toe very slightly exceeds the length of the first; subarticular tubercles indistinct, inner metatarsal tubercle very minute and the outer at the base of the fourth toe still more minute; no rudiment of web. No tarsal fold. Outer metatarsal separated by grooves

Skin smooth or slightly granulate behind the eyes and shoulders; no folds on the back or sides in the living forms (preserved specimens show them on the sides due to shrinkage), a short temporal fold only occasionally present on one side and in that case extending beyond the shoulder. Abdomen and lower surface of thighs finely granulate

Upper parts of the body and limbs deep bronze or black uniformly; in the males throat is yellowish, in the females the throat and abdomen and undersurface of thighs very finely marbled, hand white; foot darker; the red eyes in the living forms are very bright and conspicuous.

Males with internal vocal sacs united into a comparatively large structure extending to the breast.

Omosternum and sternum cartilaginous and slender terminal phalangeal bones of toes slightly expanded, nasal bones relatively wide, separated from one another and from the fronto-parietals by narrow grooves.

MEASUREMENTS.

	mm		mm
Snout to vent	18.00	First finger	1.50
Head	6.00	Second finger	2.00
Width of head	7.00	Third finger	3.50
Snout	3.00	Fourth finger	3.00
Distance between nostrils and tip of snout	1.50	Thigh	5.00
Distance between eye and tip of snout	1.50	Tibia	7.75
Internasal width	2.00	Width of thigh	3.00
Interorbital width	3.00	Width of tibia	2.00
Tympanum		Tarsus	5.00
Eye	2.00	Width of tarsus	2.00
Upper eyelid	2.00	First toe	1.50
Fore limb	4.50	Second toe	1.75
Fore arm	3.00	Third toe	3.00
Arm	3.00	Fourth toe	5.00
		Fifth toe	4.00

*Habitat*—Hills of Kempfoley Ghats, Hassan, Mysore, S. India  
Four specimens

*Type*.—Central College, Bangalore

The frogs were found under the rocks near waterfalls and they are incapable of burrowing. Nothing is known about their breeding habits.

This species may be distinguished from *N. beddomii* thus—

- 1 Tibio-tarsal articulation reaching the tip of the snout or not quite so far; limbs barred, white below immaculate, tongue without an elevated button-like structure in front . . . . . *N. beddomii*
- 2 Tibio-tarsal not reaching the eye, limbs without bars, beneath finely marbled; tongue with a button-like structure in front . . . . . *N. kempfoleyensis*

Genus *BUFO*.*Bufo brevirostris*, n. sp.

Crown without bony ridges Habit stout. Head perfectly flat, broader than long; snout rounded, not projecting beyond the mouth, as long as the eye; canthus rostralis angular; loreal region horizontal, broadly concave, nostril nearer to the end of the snout than to the eye, distance between the nostrils  $\frac{1}{2}$  in the distance between the eyes, upper eyelid slightly shorter than the interorbital width, tympanum small, fully developed,  $\frac{1}{2}$  the diameter of the eye, distance between the eye and the tympanum  $\frac{1}{2}$  the diameter of the latter or  $\frac{1}{2}$  the diameter of the former

Fore limb very short; fingers short, rather slightly swollen at the extremity; first equal to the second, third as long as the arm and fourth not longer than the first or the second; subarticular tubercles feebly developed. The tubercle at the base of the third finger is much larger than that at the base of the first.

Parotoids elongate, moderately prominent, as long as the arm, slightly more than twice its own width

Hind limb short, the tubio-tarsal articulation not reaching the shoulder, the heels just touch when the limbs are folded at right angles to the body, thigh nearly as long as wide; tibia about  $2\frac{1}{2}$  as long as broad and  $1\frac{1}{2}$  times longer than the tarsus; toes very short, tips rather obtuse, entirely free, subarticular tubercles not prominent, inner metatarsal tubercle about  $\frac{1}{2}$  in the length of the first toe, the outer much smaller situated at the base of the fifth toe

Upper surface of the skin covered with small uniformly distributed tubercles; with a small row of larger warts on the median line of the back, the first pair being the largest; throat and abdomen with spiny granules which are more prominent on the limbs. A dark temporal line extending to the sides.

Pale brown above, with a network of dark lines, sides lighter with similar marbling, ventral surface dirty white with dark blotches; under-surface of thighs white.

## MEASUREMENTS.

	mm.		mm.
Snout to vent ..	27.00	Fore arm ..	7.00
Eye ..	3.00	First finger ..	3.00
Tympanum ..	2.00	Second finger ..	3.00
Parotoids ..	4.50	Third finger ..	4.50
Width of Parotoids ..	2.00	Fourth finger ..	3.00

	mm.		mm.
Head ..	8.00	Thigh ..	5.00
Width of head ..	9.00	Width of thigh ..	
Snout ..	3.00	Tibia ..	8.00
Interorbital width ..	3.00	Width of tibia ..	3.00
Upper eyelid ..	2.50	Tarsus ..	6.00
Distance between eye and tympanum ..	1.00	Width of tarsus ..	2.25
Distance between eye and nostril ..	2.00	Inner metatarsal tubercle ..	1.00
Distance between eye and snout ..	1.00	Outer metatarsal tubercle ..	
Distance between nostrils ..	2.00	First toe ..	3.00
Arm ..	4.50	Second toe ..	4.00
		Third toe ..	5.75
		Fourth toe ..	8.00
		Fifth toe ..	5.00

*Habitat*—Kempholey, Hassan District, Mysore State

A single specimen

*Type*—Central College, Bangalore

#### Genus PHILAUTUS

Dr Malcolm Smith in describing some specimens of *Phslautus* (*Ixalus*) from the Malay Peninsula observes that "it is now generally recognised that the presence or absence of vomerine teeth can no longer be regarded as a character of generic distinction. Although fully accepting this principle, I am not in agreement with those authors who have hastily sunk all the species of *Philautus* under *Rhacophorus*. That some forms of *Philautus* should be placed under *Rhacophorus* is no doubt correct, but the distinctive habit of many other species, suggests that characters will yet be found to retain them apart from *Rhacophorus*. Until a proper revision of the whole group is taken, I prefer to retain the two genera as defined by Boulenger." Now the revision was undertaken by my colleague Mr J. S. Ramaswami who in reviewing the osteology of the two genera, comes to the conclusion that "after a perusal of the characters enumerated in the résumé (which embraces 21 points) it will be noticed that *Rhacophorus* and *Philautus* agree with each other in seven of the minor features referred to above. It is, therefore, expedient at the present state of our knowledge to treat *Rhacophorus* and *Philautus* as two independent genera." I have myself examined the morphological characters of the several forms of both genera and I have no hesitation in accepting the conclusion of Mr Ramaswami that *Rhacophorus* and *Philautus* should be treated as separate genera which is in accordance with the suggestion of Dr Malcolm Smith. But I am not quite sure if the diagnostic features given by Boulenger of *Phslautus* are

adequate and whether they do not overlap with those of *Micrixalus*. Omitting the common features, the only characters in the definition of the two genera, as given by Boulenger, separating them are the presence (*Philautus*) or absence (*Micrixalus*) of an intercalary ossification between the penultimate and distal phalanges and the latter being obtuse in *Philautus* and T-shaped in *Micrixalus*. In other respects the differences do not seem so fundamental as to justify the retention of the two genera as distinct entities and the forms of *Micrixalus* in our collection are too few to suggest the fusion of *Philautus* and *Micrixalus*. If the behaviour patterns of the Batrachians afford an indication, as is assumed by Noble, of phyletic relationships, then nothing can be easier than to bring *Micrixalus* under *Philautus*, for our observations on the breeding habits and on the larval forms of the members of these two genera point to a close agreement. But we know that arguments based on this correspondence may be used as a factor only in emphasising the evidence derived from other sources, but they alone cannot constitute a testimony sufficient to favour the merging of one genus into another. In this paper I retain the generic rank of *Philautus* and *Micrixalus*.

*Philautus charsus*, n. sp.

Vomerine teeth none. Papillæ on the tongue absent.

Snout acutely pointed as long as the diameter of the eye. Canthus rostralis distinct. Loral region concave, strongly obliquely disposed. Nostrils nearer to the tip of the snout than to the eye. Distance between eye and nostrils equals half the interorbital width. Interorbital space twice as broad as the upper eyelid and broader than the diameter of the eye. Tympanum distinct, about one-third the diameter of the eye, and separated from it by half its own diameter. Internarial width half the interorbital space, equals the distance between the eye and the nostril. Fingers not long; tips provided with discs with a narrow crescentic groove separating the dorsal and ventral portions. First finger smaller than the second, slightly less than half the length of the third. Subarticular tubercles moderately developed.

Hind limbs long, the tibio-tarsal articulation reaches the nostril, the heels touch when the limbs are folded at right angles to the body. Thigh less than half in the total length of the body. Tibia nearly as long as the thigh, and more than three times as long as broad. Metatarsus considerably more than half the length of the tibia. Toes rather short, tips dilated into discs, one-third to half-webbed. Metatarsals united. Subarticular tubercles feebly developed, a very minute inner metatarsal tubercle present. No tarsal fold.

Upper surface of the skin finely granulate, the granules confined to the dark median portion, sides smooth. Granules occur on the upper eyelid and on the dorsal portion of the snout. No dorso-lateral glandular fold. Throat smooth, chest and abdomen finely granulate. A supra temporal fold present.

Upper surface of the snout and sides of the body bright yellow (pale in spirit). A dark hour glass-shaped mark starting between the eyes and covering the hinder half of the eyelid, occupies the back extending nearly to the vent. A conspicuous chevron-shaped dark marking on the vent. Canthus and loreal region and upper jaw dark brown. Lower jaw with white and dark dots and vertical lines. Between the eye and the shoulder a deep black mark, covering the tympanum, the lower border of which bears a white crescentic line. Fore arm banded. Third and fourth fingers bear dark stripes and the upper surface of the discs of these digits having dots. Upper surface of thighs with cross bars, the posterior border bronzed with faint white spots. Tibia with stripes, which are inconspicuous on the metatarsus. Heel bears a dark black spot, from which a dark line runs down to more than half the distance of the last two toes, whose upper surface is striped and the top of the discs of all toes faintly dotted. The throat, belly and undersurface of thighs and arms white.

This specimen of *Philautus* is named after my colleague Mr. B. R. Seshachar who has collected a large number of South Indian Batrachians.

#### MEASUREMENTS.

	mm.		mm.
From snout to vent	23.00	Width of fore arm	2.00
Diameter of the eye	3.00	First finger	2.25
Upper eyelid	2.00	Second finger	3.00
Interorbital space	4.00	Third finger	5.00
Distance between eye and nostril	2.00	Fourth finger	4.00
Internarial distance	2.00	Thigh	10.50
Tympanum	1.00	Width of thigh	3.50
Distance between eye and tympanum	0.50	Tibia	11.00
Head	8.50	Width of tibia	3.00
Width of head	9.00	Metatarsus	7.00
Snout	3.50	Width of metatarsus	1.75
Arm	4.00	First toe	2.00
Width of arm	1.50	Second toe	3.00
Fore arm	5.00	Third toe	5.00
		Fourth toe	6.50
		Fifth toe	5.00

Locality.—Kottigehar, Kadur.

Type.—Central College, Bangalore

*Philautus elegans*, n. sp

Vomerine teeth none. A small papilla on the anterior portion of the tongue, retracted into the sheath. Head flat, longer than broad. Snout elongate, sharply rounded anteriorly, bevelled, projecting beyond the mouth. *Canthus rostralis* distinct, strongly angular. Loreal region concave, horizontal. Nostrils nearer to the eye than the snout. Distance between the nostrils slightly more than the interorbital space. Upper eyelid equals or slightly less than the interorbital space. Tympanum distinct, half the diameter of the eye, closely touching it.

First finger slightly shorter than the second, third longer than the snout. Subarticular tubercles fully developed. A shallow groove separates the dorsal and ventral portions of the discs.

Hind limb rather moderately long, the tibio-tarsal articulation reaching the eye. Heels touch each other when the limbs are folded at right angles to the body. Thigh two and a half times as long as broad and less than half the length of the body. Tibium slightly longer than thigh and is just half the length of the body and slightly less than three times its breadth. Metatarsus about three times as long as broad. Toes rather short, end in discs, about less than half-webbed. Outer metatarsals united at the base, bearing an outer fold of skin. Subarticular tubercles feebly developed. A small outer metatarsal tubercle.

Skin with minute wart-like tubercles disposed longitudinally on the back, or perfectly smooth. A supra-temporal fold absent. Dorsal-lateral glandular fold, feebly developed, extending from the posterior angle of the eye to the groin. The two are separated by a space nearly  $\frac{1}{2}$  in length of the body. Lower parts of the limbs and body smooth. Small glandular swellings behind the jaws and in front of shoulder.

Crimson above. A conicave dark streak between the eyelids. Sides of body commencing from behind the eyes black. Tympanum red. Loreal region faintly bronzed. The posterior border of upper jaw white. Lower jaw with a few black dots. Posterior border of the fore arm vermiculated. Posterior limbs with dark bands. Posterior border of thighs with white spots on a black background. The cross bars extend to the metatarsus. Last two toes with dark bands. Upper surface of discs dark. Throat and chest marbled. belly yellow (white in spirit). Thighs pinkish.

This is the prettiest specimen in the collection.

## MEASUREMENTS.

	mm.		mm.
From snout to vent	23.0	Width of fore arm	1.75
Head ..	9.00	First finger ..	3.00
Width of head	7.00	Second finger ..	3.50
Diameter of the eye	3.00	Third finger ..	4.75
Upper eyelid	2.00	Fourth finger ..	3.50
Interorbital space	2.50	Thigh ..	10.00
Snout ..	3.75	Width of thigh ..	4.00
Distance between eye and nostril	2.00	Tibia ..	11.50
Internarial width	2.75	Width of tibia ..	3.00
Tympanum	1.50	Metatarsus ..	6.00
Distance between tympanum and eye	Nil	Width of metatarsus ..	2.00
Arm ..	6.00	First toe ..	4.00
Width of the arm ..	1.75	Second toe ..	5.00
Fore arm ..	6.00	Third toe ..	7.00
		Fourth toe ..	9.00
		Fifth toe ..	7.00

Locality — Kempholey, Hassan

Type — Central College, Bangalore

*Philautus hottentotensis*, n. sp.

Vomerine teeth none Tongue with a papilla, the sac with tumid lips

Head depressed Upper surface of the snout deeply concave, bounded by ridges anteriorly and laterally Tip of snout acutely pointed, broader truncated, and projecting beyond the mouth Snout longer than the eye Canthus rostralis prominent and angular Loreal region concave, oblique, becoming continuous with the suborbital grooves. Nostrils prominent, equidistant between tip of snout and eye Internarial width equals the diameter of the eye Interorbital width less than the upper eyelid Tympanum fairly distinct, about half the diameter of the eye, which it closely touches

Fingers tipped with large discs, about twice as broad as the penultimate phalangeal segment First finger slightly longer than the second and the third very slightly longer than the snout All the fingers are free Subarticular tubercles feebly developed.

Hind limbs long. Thigh more than twice as long as broad and as long as the tibia Tibio-tarsal articulation reaches tip of the snout Metatarsus longer than arm and more than twice as long as broad Toes rather short, tip ending in discs, web extending to the discs. Outer metatarsals united

at the base or separate Subarticular tubercles feebly developed, a very minute inner metatarsal tubercle

Upper surface of body with minute tubercles, a supra-temporal fold and dorso-lateral folds are present. The latter commence well behind the eyes, and are separated from each other by a distance less than 1/5 in the total length of the body. Deep suborbital grooves confluent with the loreal region present. Undersurface of body and thighs smooth

Uniform deep bronze above and sides of head, body and limbs, hind limbs and fore arms barred by deeper stripes. Discs of fingers and toes dark on both surfaces. Web dark, and the folded toes produce a characteristic deep black colour. Lower jaw and throat bronzed—the dark colour extending sometimes to the chest and the anterior division of the abdomen, or the latter two regions may be bright orange, ventral undersurface of thighs red, the posterior border feebly marbled. Sides of body yellowish or whitish

#### MEASUREMENTS

	mm.		mm.
From snout to vent	23.00	Width of fore arm	2.00
Head	8.00	First finger	3.00
Width of head	6.00	Second finger	3.50
Snout	4.00	Third finger	4.75
Eye	3.00	Fourth finger	4.00
Upper eyelid	2.00	Thigh	15.00
Interorbital space	2.00	Width of thigh	4.50
Tympanum	1.50	Tibia	15.00
Distance between tympanum and eye	Nil	Width of tibia	3.00
Distance between eye and nostril	2.00	Metatarsus	8.00
Internasal distance	3.00	Width of metatarsus	2.25
Arm	7.00	First toe	3.25
Width of arm	2.00	Second toe	5.00
Fore arm	5.00	Third toe	7.75
		Fourth toe	11.00
		Fifth toe	8.00

*Locality*—Kottigehar, Kadur

*Type*—Central College, Bangalore

#### *Philautus swamianus*, n. sp.

Vomerine teeth none. Tongue large, bifid behind, a conical papilla nearer to the anterior end, retractile sheath an elongated slit

Head perfectly flat, depressed, snout pointed, projecting beyond the mouth, longer than the eye. Canthus rostralis prominent, angular. Loreal region concave, horizontal, separated from the narrow suborbital or upper

narial groove. Nostrils prominent, about midway between the eye and the tip of the snout. Distance between nostrils very slightly less than interorbital space, equals diameter of the eye. Interorbital space wider than the upper eyelid, and exceeds the eye by a narrow margin. Tympanum half hidden, and about half the diameter of the eye from it is separated by a distance less than its width.

Tips of fingers with discs, nearly twice as broad as the penultimate phalangeal segment. There is no groove separating the dorsal and ventral portions of the discs. First finger shorter than the second, the former about the diameter of the eye. The third slightly exceeds the length of the snout. Subarticular tubercles feebly developed on the first and the second fingers and absent from the third and the fourth fingers.

Hind limb fairly long, the tibio-tarsal articulation reaching beyond the eye, heels overlap when the limbs are folded at right angles to the body. Thigh  $2\frac{1}{2}$  times as long as broad and tibia about as long as thigh and slightly less than  $3\frac{1}{2}$  times its width. Toes rather short, web extending to the discs which are equal to twice the width of the penultimate phalangeal segment. Subarticular tubercles poorly developed. A small inner conical metatarsal tubercle present. Outer metatarsals separated at the base. No tarsal fold. Heels prominent.

Skin smooth above and also below. Supra-temporal fold inconspicuous. Dorsal-lateral folds occur, starting from behind the eyes and separated from each other by a space equal to the interorbital width. Folds may be interrupted, not quite reaching the groin. In addition there are two fairly well-developed lateral folds, converging towards the vent, beginning from about the middle of the sides of the body. The two sets of folds may become continuous.

Upper surface of the body uniformly bluish, dark bronze in spirit. Loreal region and upper jaw lighter. Thighs and tibia barred. Fore limbs without bands. On the sides of the body, commencing from the eyes, are fine rows, two or three, of white glandular bodies which meet in front of the vent. Throat pale buff, chest marbled, abdomen white. Anterior portion of the ventral surface of thighs pinkish, the posterior vermiculated. Under-surface of tarsus, foot and discs bronzed.

I have named this species after my colleague Mr. L. S. Ramaswami, whose contributions to our knowledge of the cranial morphology of the Anura of South India have been of invaluable assistance to me.

## MEASUREMENTS.

	mm		mm.
From snout to vent	29.00	Width of fore arm	2.00
Head ..	10.00	First finger ..	3.00
Width of head ..	8.00	Second finger ..	3.50
Eye ..	3.00	Third finger ..	5.00
Upper eyelid ..	2.00	Fourth finger ..	4.00
Interorbital space ..	3.50	Thigh ..	15.00
Distance between eye and nostrils ..	2.00	Width of the thigh ..	6.00
Snout ..	4.00	Tibia ..	15.00
Interorbital space ..	3.00	Width of tibia ..	4.50
Tympanum ..	1.50	Tarsus ..	6.00
Distance between eye and tympanum ..	1.00	Width of metatarsus ..	2.25
Arm ..	6.00	First toe ..	3.00
Width of arm ..	2.00	Second toe ..	5.00
Fore arm ..	5.50	Third toe ..	6.50
		Fourth toe ..	8.50
		Fifth toe ..	6.50

*Locality*.—Kottigehar, Kadur

*Type*.—Central College, Bangalore

*Philautus melanensis*, n. sp.

No vomerine teeth. Mouth large. Tongue comparatively small, a short conical papilla on the anterior half of the tongue. Lower jaw with a small anterior median tooth-like prominence, with deep indents on each side.

Head rather large, depressed nearly as broad as long, or only slightly longer. Snout blunt, rounded anteriorly, a little longer than the eye, projecting slightly beyond the mouth. Canthus rostralis prominent, broad angular. Loreal region concave, nearly horizontal or slightly oblique, continuous with the suborbital groove. Diameter of the eye equals the width of the upper eyelid. Interorbital distance greater than the eye. Nostrils nearer the tip of snout than the eye, interorbital space about half or only slightly less than half the length of the snout. Tympanum distinct, about two-thirds in the diameter of the eye, which it almost touches or may be separated by a line.

Fingers with discs which are about twice as broad as the penultimate phalangeal segment. A distinct crescentic groove separates the dorsal and ventral parts. First finger shorter than the second and the third twice as

long as the first or twice the diameter of the eye. Fourth finger equals or very slightly longer than the snout. Subarticular tubercles well developed

Hind limbs rather long, the tibio-tarsal articulation reaching the tip of the snout or beyond. Heels overlap when the limbs are folded at right angles to the body. Thigh less than four times its own width and is shorter than tibia. Tibium more than four times its own width. First three toes free, a rudimentary web at the base of the fourth and fifth toes. Discs as broad as the penultimate phalangeal segment. Subarticular tubercles poorly developed. Outer metatarsals united at the base. An inner metatarsal tubercle in an extremely rudimentary condition. No outer metatarsal tubercle. No tarsal and metatarsal folds.

Upper surface of skin with or without short longitudinal folds covered uniformly with fine white granules. These granules extend to the thighs and sometimes to the tibia. A supra-tympanic fold. Throat, chest, belly and undersurface of limbs and toes covered with large tubercles.

Uniform black above. Anterior limbs black. Thighs and tibia either black or pale yellow with cross bars. Ventral surface of body and thighs yellowish. Undersurface of tibia and foot, black or bronze.

#### MEASUREMENTS.

	mm		mm.
From snout to vent	29.00	Width of fore arm	.. 3.00
Head	13.00	First finger	.. 3.00
Width of head	12.00	Second finger	.. 4.00
Eye	3.00	Third finger	.. 6.00
Upper eyelid	3.00	Fourth finger	.. 1.50
Interorbital space	5.00	Thigh	.. 15.00
Distance between eye and nostril	3.50	Width of thigh	.. 4.00
Snout	4.00	Tibia	.. 17.00
Internarial distance	2.50	Width of tibia	.. 4.00
Tympanum	2.00	Tarsus	.. 9.00
Space between eye and tympanum	Nil	Width of tarsus	.. 3.00
Arm	8.00	First toe	.. 3.00
Width of arm	3.00	Second toe	.. 4.50
Fore arm	8.00	Third toe	.. 8.00
		Fourth toe	.. 12.00
		Fifth toe	.. 8.25

*Locality*.—Kempholey, Hassan.

*Type*—Central College, Bangalore

*Philautus narasinghi*, n. sp.

No vomerine teeth, tongue small, thin, leaf-like with a conical papilla, retractile sheath slit-like. Mouth rather small. Choana broadly separated, situated at the extreme lateral margin.

Head depressed longer than broad, snout acutely pointed, the prenasal region acutely triangular viewed from above. In front, the snout bears pits and grooves, the latter vertically disposed, the tip of snout looking like a prostomeal lobe. Snout projects beyond the mouth, a conspicuous glandular mental pad. *Canthus rostralis* prominent, ridge-like and angular. Loral region concave, horizontal, confluent with the anterior rostral pits in front and suborbital groove behind. Upper surface of the internarial region concave, bounded anteriorly by a transverse bar of the prostomial lobe; eye  $1\frac{1}{2}$  times the upper eyelid which equals the interorbital space. Snout longer than the eye and the nostrils about midway between the eye and tip of the snout. Internarial space wider than the interorbital width and equals the diameter of the eye. Tympanum moderately distinct, about  $\frac{1}{3}$  the diameter of the eye from which it is narrowly separated.

Fingers rather short with discs which are small on the first and the second, larger on the fourth which is twice as wide as the penultimate phalangeal segment. First finger shorter than the second. The fourth finger a little longer than the eye. A single subarticular tubercle on the first finger, others nearly free. Grooves of the discs not clear.

Hind limb rather long, the tibio-tarsal articulation reaching beyond the tip of the snout. Heels overlap when the limbs are folded at right angles to the axis of the body. Thigh more than three times as long as wide, tibia longer than the thigh and about 5 times as long as broad. Toes rather short, discs small. Web extending upto the discs of first two toes, but stops at the base of the penultimate phalangeal bone of the fourth toe, and at the base of the phalangeal bone of the third and the fifth. Outer metatarsal free at base. Subarticular tubercle feebly developed. No inner or outer metatarsal tubercle. Heel not marked, no dermal appendage. No tarsal and metatarsal folds.

Skin smooth above and below. No supra-temporal fold. The place of dorso-lateral folds is taken by two short folds commencing from behind the eyes, terminate on the back, a little behind the shoulder, about  $1\frac{1}{2}$  times as long as the snout, and separated from each other by a space equal to the interorbital width.

Brownish red above including the limbs. The anterior border of the thighs and the upper part of tibia very feebly banded. The posterior border of thighs and the inner border of tibia feebly vermiculated. Throat

and chest buff coloured. Belly dirty white. Lower surface of thighs and the tibia whitish.

This species is named after my colleague Mr A Narayana Rao, who has added considerably to our batrachian collection.

MEASUREMENTS

	mm.		mm.
From snout to vent	29.00	Width of fore arm	2.00
Head	10.00	First finger	2.00
Width of head	7.50	Second finger	3.25
Diameter of the eye	3.00	Third finger (Injured in both arms)	
Upper eyelid	2.00	Fourth finger	3.50
Interorbital space	3.00	Thigh	13.00
Snout	4.00	Width of thigh	4.00
Distance between eye and nostril	2.00	Tibia	15.00
Internasal space	3.00	Width of tibia	3.00
Tympanum	1.50	Tarsus	6.25
Distance between tympanum and eye	0.50	Width of tarsus	2.00
Arm	7.00	First toe	3.00
Width of arm	2.00	Second toe	5.00
Fore arm	5.00	Third toe	8.00
		Fourth toe	10.75
		Fifth toe	8.00

Locality —Kottigehar, Kadur

Type —Central College, Bangalore.

*Phelsuma longicrus*, n. sp.

Vomerine teeth none. Tongue without papilla. Choana laterally situated. Head moderately depressed, as long as broad. Snout obtusely pointed, a little longer than the eye, scarcely projecting beyond the mouth. Canthus rostralis prominent. Loral region slightly concave and horizontal. Nostrils midway between the eye and the tip of the snout. Distance between the nostrils equals the diameter of the eye or the interorbital space. Upper eyelid about  $\frac{1}{3}$  in the interorbital space. Tympanum distinct, about half the diameter of the eye, being separated from it very narrowly.

Fingers moderately developed. Discs not broad, slightly wider than the terminal phalangeal segment. First finger shorter than the second. Subarticular tubercles fairly well developed. Length of arm equals the length of the snout.

Hind limb very long. The tibio-tarsal articulation reaching far beyond the tip of the snout. Heels strongly overlap when the limbs are folded at right angles to the body. Tibium more than four times as long as broad and is longer than the thigh. Tips of toes dilated into discs, half-webbed.

Outer metatarsals united at the base. A minute elongate inner metatarsal tubercle. Subarticular tubercles fairly well-developed. A faint outer metatarsal fold. No tarsal fold.

Upper surface of the skin with faint folds. Sides with short glandular folds, serially arranged. Minute tubercles occurring between the series. Ventral surface of body and thighs smooth. Upper surface of snout pale grey—a dark band between the nostril and eye over canthus rostralis. Loreal and suborbital region yellow, extending as far behind as the angles of the mouth. A brown mark over the supra-tympanic fold. Tympanum reddish. Upper and lower jaw with dark vertical bands, the upper series terminating just below the middle of the eye. Interorbital space with a faint transverse band. Upper surface of body olive brown. Thighs with cross bars on the anterior border, the posterior border minutely marbled. Tibium also barred anteriorly, but whitish posteriorly. A dark line stretching from heel to foot. Lower surface of the body and thighs white.

## MEASUREMENTS.

	mm.		mm.
From snout to vent	20.00	Width of fore arm	2.00
Head	8.50	First finger	2.75
Width of head	8.50	Second finger	3.00
Snout	4.00	Third finger	3.75
Eye	3.00	Fourth finger	2.50
Upper eyelid	2.00	Thigh	10.00
Interorbital space	3.00	Width of thigh	4.00
Distance between eye and nostril	2.00	Tibia	13.00
Internasal distance	3.00	Width of tibia	3.00
Tympanum	1.50	Tarsus	6.25
Distance between tympanum and eye	0.75	Width of tarsus	2.00
Arm	4.00	First toe	2.00
Width of arm	2.00	Second toe	4.00
Fore arm	5.00	Third toe	6.00
		Fourth toe	10.00
		Fifth toe	5.50

Locality.—Kempholey, Hassan.

Type.—Central College, Bangalore.

*Phlaeius montanus*, n. sp.

Vomerine teeth absent. Tongue with a short papilla situated rather anteriorly and in the spirit specimen retracted into a pit, strongly bifid. Head depressed as long as broad. Snout rounded, broadly truncate, not projecting beyond the mouth. Snout longer than the eye. Canthus rostralis vertical, loreal region concave and horizontal. Nostrils nearer to

the tip of snout than to eye. Internarial distance equals the space between the eye and the nostrils. Diameter of the eye equals the interorbital width and is more than twice the tympanum. Upper eyelid less than interorbital space. Tympanum not prominent, about half the diameter of the eye from which it is separated by a narrow space.

Fingers moderately long, tips provided with discs bearing the groove separating the upper from the lower portion. No web. Discs small, smaller than the tympanum. First finger shorter than the second. Third finger longer than the snout. Subarticular tubercles moderately developed.

Hind limb long; the tibio-tarsal articulation reaching the eye. Heels do not overlap when the limbs are folded at right angles to the body. Thigh more than three times its width, tibia as long as the thigh and more than four times its own width. Metatarsus equals or slightly longer than the third or fifth toe. Toes not long, about two-thirds webbed. Discs smaller than those of the fingers. A small oval inner metatarsal tubercle present. Base of toes united, outer metatarsal without a fold. No outer metatarsal tubercle. Subarticular tubercles moderately developed.

Skin smooth above. Throat, chest and abdomen and undersurface of the arms and hand finely granulate. A feebly developed supra tympanic fold present. A faintly V-shaped fold of skin on the occiput, commencing from the middle of the eyelids. No dorso-lateral fold.

Bright red above in the living condition, brownish red in spirit, with or without white spots on the body. Throat and upper part of chest brownish, speckled with white. Rest of the ventral surface yellowish, the dark bands meeting on the anterior border of thighs give rise to ocellus-like spots when viewed ventrally. A brown band between the eye-lids, faint in spirit. Thighs barred, a single broad band on the shank. Undersurface of hind limbs yellow with brown marbling.

#### MEASUREMENTS.

	mm		mm
From snout to vent	. 37.00	Width of arm	. 3.00
Head ..	. 14.00	Width of fore arm	. 2.75
Width of head	. 14.00	First finger	. 3.25
Internarial distance	. 3.00	Second finger	. 4.50
Distance between eye and nostril	. 3.00	Third finger	. 7.50
Eye ..	. 5.00	Fourth finger	. 6.25
Nostril ..	. 6.00	Thigh ..	. 17.00
Interorbital width	. 5.00	Width of thigh ..	. 5.00
Upper eyelid ..	. 3.00	Shank ..	. 17.00
		Width of shank ..	. 4.00

	mm		mm.
Tympanum ..	2 50	First toe ..	3 50
Distance between tympanum and eye ..	1 00	Second toe ..	5 00
Arm ..	6 00	Third toe ..	9 00
Fore arm ..	8 00	Fourth toe ..	12 00
		Fifth toe ..	9 00

*Locality* —Hills of Kempholey, Hassan

*Type* —Central College, Bangalore

*Family MICROHYLIDAE*

*Ramanella minor*, n. sp.

Post-narial ridges strongly developed, obliquely set, meeting dorsally, with an extremely narrow ventral cleft. Anterior pharyngeal fold inconspicuous. Snout short, broadly truncated. Distance between nostrils equals width of the upper eyelid. Interorbital space nearly twice the width of the upper eyelid. Diameter of the eye less than the length of the snout. Canthus rostralis obtuse. Loreal region almost vertical. Distance between eye and nostril equals the diameter of the eye.

Tibio-tarsal articulation touches middle of the body. Toes entirely free, tips almost pointed. Subarticular tubercles well-developed. Inner metatarsal tubercle conical, moderately developed. Outer metatarsal tubercle rounded, inconspicuous. When the hind limbs are folded at right angles to the body, the heels touch.

Fingers with triangular dilatations, nearly twice as broad as the penultimate joint.

Skin smooth above, slightly pustular on the sides. A fold from the posterior angle of the eye to the shoulder. No occipital fold. Lower surface smooth.

Upper surface of snout olive, with a median dark band, short not reaching the interorbital space. Sides of the snout dark which is continuous with the dark on the sides and ventral surface of the body. A dark spot between the eyes on the upper surface of the head. A dark broad band on the dorsum, flanked by olive bands. The latter do not extend to the sides of the body. The former stops at the coccyx, which is olive mottled, with dark irregular markings. The anterior and posterior limbs entirely black. The ventral surface including the throat black. The abdominal region and thighs bearing white spots.

## MEASUREMENTS.

4.—25 mm.

Diameter of the eye	..	2.00	Third toe	..	..	6.00
Length of snout	..	2.75	Fourth toe	..	..	9.00
Distance between the nostrils	..	1.50	Fifth toe	..	..	6.00
Interorbital width	..		Thigh across the anterior border	..	..	7.00
Distance between eye and nostril	..	2.00	Thigh across the anterior border ventrally	..	..	9.00
First toe	..	2.00	Shank	..	..	9.75
Second toe	..	4.00	Tarsus	..	..	5.50

*Locality*—Saklespur, Hassan*Type*—Central College, Bangalore*Ramanella triangularis rufoventris*, n. var

In the plantain gardens behind the houses in Saklespur, there is a small variety of *R. triangularis*, occurring fairly in large numbers. These are found in the whorls of the leaf stalks, and are mostly solitary. This variety possesses the power of climbing smooth surfaces like *R. variegata*.

This variety differs from the *R. triangularis* in two characteristics:

1 The snout is pointed, tips obliquely truncated, prominent

2 Tibio-tarsal articulation stands well behind the shoulder.

In regard to colour, almost every specimen in the collection bears an occipital dark band continuous at the sides with the tympanoloreal band. This cross band may or may not be continuous with the dorso-median band. Even in the spirit specimens, the lower surface is reddish (in the living state, the red is bright and warm) which extends over the undersurface of the anterior and posterior limbs. The ventral white spots are confined to the abdomen and the lower surface of thighs. A short dark streak on the rostrum is always present. The olive is replaced in this variety by grey. Limbs barred with dark red.

*Ramanella triangularis* of the plains is not met with in the Malnad areas and it is represented by this local variety which may be designated *Ramanella triangularis rufoventris*.

Length 22 mm. Specimens, Central College, Bangalore.

*Locality*.—Mudigere, Kadur, Saklespur, Hassan.

*Ramanella marmorata*, n. sp.

Post-choanal ridges transversely disposed, well developed, a broad median gap between the two ridges. The anterior pharyngeal ridge strongly arched, the limbs dilated into thick lateral swellings. The fimbriae of the posterior pharyngeal ridges not conspicuous. Snout short, truncated, not prominent, as long as the diameter of the eye. Canthus rostralis rounded. Loreal region oblique. Rostralis rounded. Inter-orbital space much broader than the upper eyelid which equals the internarial distance.

Fingers with wide triangular dilation, which are twice as large as the penultimate joints.

Toes ending obtusely, with a suspicion of a web at the base, sub-articular tubercles moderately developed. Inner metatarsal tubercle shovel shaped. Outer metatarsal tubercle well developed, conical, situated at the base of the fifth toe. Tibio-tarsal articulation does not reach the shoulder, but stands well behind the axilla. Heels stand apart when the hind limbs are flexed behind at right angles to the body. Skin on the upper surface is warty in the young specimens, tubercles becoming confined to the posterior half of the body, sometimes extending to the dorsal surface of the hind limbs in the mature forms. Lower surface perfectly smooth. Supra tympanic fold inconspicuous. No occipital fold.

Immature specimens pale olive or pale reddish brown above. Mature males bright reddish brown above, mature females deep olive above. A dark broad band between the eyes, sometimes involving the lids. Snout bearing a triangular or V-shaped dark mark. A broad angular dark band between the shoulders. Behind this region, dark spots or irregular markings invariably occur which may sometimes extend to the sides. No lateral dark bands on the body. Loreal region free. Limbs bright yellow or deep olive green according to the sex. The posterior border of the arm bears a dark band. Fore arm, hand, fingers, thigh, shank, tarsus, foot and toes barred. The crural band usually extending into the groin, may be reduced to a spot continuous with the prevailing colour of the upper surface. Lower surface including the throat and the limbs beautifully marked in the male with yellow and reddish brown and in the female with dark blue and moss green.

## MEASUREMENTS

	mm.		mm.
Diameter of the eye . . .	3.00	Thigh . . .	7.00
Distance between eye and nostril . . .	2.75	Width of thigh . . .	5.00
Snout . . .	3.00	Shank . . .	9.00
		Width of shank . . .	3.00

Internarial distance	2.00	Tarsus	..	4.00
Interorbital space	3.75	Width of tarsus	..	2.00
Upper eyelid	2.00	Width of foot	..	3.00
Upper arm	4.00	First toe	..	2.00
Fore arm	6.00	Second toe	..	4.25
First finger	2.50	Third toe	..	6.00
Second finger	3.00	Fourth toe	..	8.50
Third finger	5.00	Fifth toe	..	6.00
Fourth finger	4.25	Total length	..	26.00

*Locality*.—Saklespur, Hassan District, Mysore

*Type*—Central College, Bangalore

*Remarks*—The specimens were found in the whorls of the plantain leaves. They usually occur solitary. Occasionally two or three specimens may be found together in the same cramped space.

*Ramanella anamalaiensis*, n. sp.

Post-narial ridges incompletely developed, that on the right-side is absent. Post-pharyngeal ridge fimbriated. Snout broadly triangular, truncated. Head depressed. Diameter of the eye greater than the length of the snout. Interorbital space twice the width of the upper eyelid. Distance between the eye and nostril equals width of upper eyelid. Internarial distance slightly more than half the length of the snout. Canthus rostralis rounded. Loral region oblique. Supra-tympanic fold present. No occipital fold. Tips of fingers truncated, not discoidal, about the same width as the penultimate joints. Fore arm slightly longer than the third finger and twice as long as the first. Toes perfectly free, ending obtusely. Thigh slightly shorter than the third toe, shank shorter than the fourth toe. Shank more than twice as long as wide. Inner metatarsal tubercle moderately developed, shovel shaped. Outer obtusely conical placed between the bases of the fourth and fifth toes. Sub-articular tubercles moderately developed. Tibio-tarsal articulation reaches the shoulder. Heels do not touch when the legs are folded behind the vent at right angles to the body.

Upper surface of the skin warty. Lower surface smooth. Bright olive above with a broad dark median band commencing from behind the occipital region and extending upto the coccygeal region, behind this band a few irregularly shaped black-markings. Between the eyes a squarish dark mark and on each upper eyelid a dark spot. Upper surface of the snout olive. Loral region and sides of body reddish brown. Upper surface of arms olive blotched with brown. Fore arm reddish brown. Upper surface of thighs and shanks reddish brown, throat and abdomen and lower surface

of limbs pale brown. A few indistinct white spots on the belly and foot brown

Hand

## MEASUREMENTS

	mm		mm
Diameter of the eye	4.00	Shank. Length	9.50
Snout ..	3.00	Width ..	4.00
Interorbital space	4.25	Tarsus Length ..	8.00
Upper eyelid	2.00	Width ..	3.00
Distance between nostrils	1.75	First toe ..	2.00
Distance between eye and nostril	2.00	Second toe ..	4.50
Arm ..	6.00	Third toe ..	7.75
Fore arm ..	8.00	Fourth toe ..	11.0
First finger ..	4.00	Fifth toe ..	6.50
Second finger ..	5.00	Broadest part of the foot (between 2nd and 5th toes)	
Third finger ..	7.50	Total length ..	28.00
Fourth finger ..	5.50		
Thigh. Length	7.25		
Width	5.50		

Locality—Base of Anamalai Hills, Coimbatore District

Type—Central College, Bangalore

## TADPOLES

*Philautus leucophincus*

These tadpoles are small. Head and body rather narrowly oval. Upper surface convex, ventral rather flat. The snout is rounded and slopes down. Diameter of the mouth smaller than the convex interorbital space, which is as wide as the internasal breadth. Nostrils nearer to the eyes than to the tip of snout. Eyes and nostrils dorso-lateral. Spiracular tube short, not prominent. Spiracular orifice directed upwards and backwards nearer to the eye than to the root of the hind limbs. Mouth disc provided with upper and lower lips. The latter is notched in the middle. Both lips and lateral lobate borders fringed with papillae. The upper beak with a smooth border, the lower with serrated margin. No teeth. The tail is long. The upper fin commences far beyond the root of the tail, and is strongly arched. The ventral fin commences well behind the dorsal and has a straight border. The dorsal fin is deeper. Tip of tail pointed. The limbless forms have an yellow dorso-median area, which in the preorbital region becomes white which is characteristic mark of the adult. Sometimes a brown dorsal band. Sides and belly blackish. Throat whitish, muscular part of the tail yellowish with black marks. In the four-legged forms, the colour becomes uniformly

rufous, the snout remaining whitish. The dark dorsal band becomes more marked. Anal tube dextral.

MEASUREMENTS.

	mm.		mm.
Total length .	26.00	Breadth of body .	5.50
Length of head and body .	9.00	Depth of body .	4.00
Length of tail .	17.00	Depth of tail .	3.75

Locality.—Streams of Kempholey, Hassan

*Philautus hypomelas*

These tadpoles have an elongated body, and are not large. The head and body are elliptical. Snout broadly rounded, mouth ventral. Hind end of body more or less tapering. Both surfaces of body convex. Mouth small, shorter than the interorbital space. Internasal width slightly greater than the interorbital breadth. Eyes and nostrils dorso-lateral. Spiracle broad at the base, directed upwards, orifice dorso-lateral, nearer to the eye than to the root of the hind limb. Mouth disc small, provided with well-developed upper and lower lips. The latter multilobed. The lower lip and the lobate sides are free from papillae which occur only on the upper lip. The inner border of the lower lip with a crenulate edge. The upper beak is smooth, the lower serrated. No teeth. The tail is well developed. The upper fin begins well behind the root of the tail. Both fins are of equal depth and are broadly curved. Tip of tail pointed. Skin smooth. Upper surface of the body purplish or reddish brown. Preorbital region of the head transparent. Throat whitish. Belly black. Muscular part of tail reddish, with black markings. Fins grey. Anal tube dextral.

MEASUREMENTS

	mm.		mm.
Total length .	32.00	Breadth of Body .	8.00
Length of head and body .	10.00	Depth of body .	5.00
Length of tail .	22.00	Depth of tail .	6.50

Locality.—Streams of Kempholey, Hassan

*Philautus nassatus*

The tadpoles are not large. The head and body rather elongated and narrowly oval. Both surfaces are convex. The snout is rounded, slopes downwards. Dorso-median line grooved in some cases. Mouth small, shorter than the interorbital space which equals the internasal width. Eyes and nostrils dorso-lateral. Latter nearer to the eyes than the tip of the snout. Spiracle tubular, lateral pointing backwards. Spiracular opening nearer to eyes than the root of the hind limbs. The mouth disc is

small. Both lips well developed. Upper fringed with papillæ. Sides lobate, borders with more than one row of papillæ. Lower lip provided with three lobes, covered with numerous papillæ. Upper beak broadly V-shaped with a serrated border. No teeth. The tail is long and powerfully developed. The dorsal and ventral fins attain maximum depth in the posterior half of the tail. The muscular portion broader. The dorsal fin does not extend beyond the root of tail. Tail pointed. Anal tube dextral. Skin smooth. Upper surface bright yellow, with dark irregularly distributed dots. The muscular part of tail yellow, with clusters of black spots, sometimes running together to form bands. Under surface of body white. Caudal fins grey.

## MEASUREMENTS

	mm.		mm.
Total length . . . . .	32 00	Greatest width of body . . . . .	11 00
Length of head and body . . . . .	12 00	Greatest depth of body . . . . .	9 00
Length of Tail . . . . .	21 00	Greatest depth of tail . . . . .	8 00

*Locality*—Streams of Kempholey, Hassan

*Philautus pulcher*

The tadpoles are comparatively small. Head and body narrowly oval, moderately flattened above and below. In a few cases the upper surface is convex. The snout is rounded, sloping downwards. Diameter of the mouth about  $\frac{1}{4}$  in the interorbital width, which is broader than the interorbital space. Eyes dorso-lateral. Nostrils point upwards, nearer to the eyes than to the tip of snout. Spiracle distinctly tubular, directed upwards. Spiracular orifice nearer to the eye than to the root of the hind limb. The mouth disc is small. Upper lip not developed, but bears a lobe provided with a double row of papillæ border. Sides lobate, lower lip multilobed, both bearing minute papillæ. The upper beak broadly crescentic with a smooth border, lower serrated. Tail strongly developed. Upper fin commences well behind the root of the tail, both fins are well developed and are of the same depth. The upper broadly arched, the lower being nearly straight. Tip of tail pointed. Skin smooth. Upper surface of head and body slate coloured. Undersurface grey. Muscular part of tail yellow, with black markings, running into longitudinal or vertical streaks. Latter marking prominent in the hinder half of the tail. Anal tube median.

## MEASUREMENTS.

	mm.		mm.
Total length . . . . .	27 00	Width of body . . . . .	5 00
Length of head and body . . . . .	9 00	Depth of body . . . . .	4 00
Length of tail . . . . .	18 00	Depth of tail . . . . .	4 00

*Locality*—Streams of Kempholey, Hassan

*Phalaenoides variabilis*

The tadpoles are of moderately large size, the head and body being oval. Dorsal surface slightly flat, ventral distinctly convex. The snout is rounded, sloping down. Mouth ventral. Nostrils nearer to the tip of the snout, internasal space almost equal to interorbital width, eyes dorso-lateral, nostrils point upwards. Eyes nearer to the tip of the snout than to the spiracular opening. The spiracle is lateral, sinistral, orifice directed upwards and backwards. Mouth disc small, surrounded by broad lobes fringed with rows of papillæ. The upper lip has two rows of papillæ. The lower lip is divided into four lobes, the hinder border bearing small papillæ. The upper beak is broadly U-shaped and the lower V-shaped, having a serrated margin. Dorsal rows absent. The tail is well developed. The upper lobe does not extend beyond the roof of the tail. The upper fin is deeper than the lower, but less deep than the muscular part. Tip of tail pointed. Skin smooth, dorsal surface yellowish, with numerous dark dots. The muscular part of tail yellow with dark irregular blotches. Sometimes the inferior border of the lower fin, may bear fine black dots, confined to the posterior division. No preorbital and frontal glands. No sensory pits

## MEASUREMENTS

	mm	mm.
Total length . . . . .	40.00	Breadth of body . . . . .
Length of head and body . . . . .	13.00	Depth of body . . . . .
Length of tail . . . . .	27.00	Depth of tail . . . . .

Locality—Streams of Kempholev, Hassan

*Rana aurantiaca*

These tadpoles by their uniform brown colour are apt to be mistaken for those of *Bufo*, and it is interesting that the larvae of *Rana aurantiaca* the adult of which is beautifully coloured, are so drab. The size of these tadpoles, their oral and caudal characters, distinguish them at once from the larvae of *Bufo*.

The tadpoles are of moderate size. Head and body oval, not flattened dorso-ventrally, both surfaces being convex. Snout rounded, mouth small, ventral. Eyes dorsal or dorso-lateral. Nostrils pointing laterally. Width of mouth equals or less than the interorbital width which is broader than the internasal spaces. Nostrils equidistant between tip of snout and eyes. Spiracle lateral, sinistral, somewhat broadly tubular, pointing slightly upwards. The mouth disc is small, lower lip better developed. The upper free from papillæ which occur on the sides and on the outer borders of the

lower lip, with a distinct median space free from papillæ. The upper beak is broadly crescentic with a smooth border, the lower is broadly V-shaped, finely serrated. Dental formula  $1\frac{1}{2}$ . The tail is broad. The dorsal fin extends beyond the root of the tail. Both fins are almost equal, but slightly deeper than the muscular portion. Pointed at the tip. Anal tube slightly dextral. Both surfaces of the body and the muscular portion of the tail uniformly brown. Tail fins lightly coloured.

## MEASUREMENTS

	mm		mm
Total length	31.00	Greatest width of body	5.75
Length of head and body	12.00	Greatest depth of body	5.75
Length of tail	19.00	Greatest depth of tail	7.50

The tadpoles live near the water margins on the tanks at the base of hills and occur in small numbers. Mostly they rest on the floor.

*Locality*—Base of Anamalai Hills, Coimbatore District.

*Bufo brevirostris*

Generally speaking, the tadpoles of the genus *Bufo* are comparatively small and those of *B. brevirostris* are extremely so. These were taken from rain water puddles from which the adults were obtained. Besides these tadpoles those of *B. melanostictus* and *B. microtympanum* also were collected from the same situation. It is interesting to note that the shoals of tadpoles belonging to the different species keep together separately and it is this distinctive feature of their habits that led to their examination. Like the other members of the genus, the body is obcordate the greater width being at the pectoral region. The snout slopes and is rounded. The diameter of the mouth equals or is greater than the interorbital space. The eyes and nostrils are distinctly dorsal, looking upward. Interorbital width (1.5 mm) is greater than the internasal space (1.0 mm), the nostrils nearer to the eyes than to the tip of the snout. Upper lip devoid of papillæ which fringe the sides and the outer borders of the lower lip. Both lips are equally developed. Dental formula  $1\frac{1}{2}$ . The second series in the upper lip is interrupted in the middle, the three lower series are continuous almost equal to one another. The jaws are finely serrated. Skin smooth. A pair of preorbital glands present. Sensory pits on the back absent. A frontal gland is prominent. Spiracular tube opens slightly dorsalward situated about the middle of the body, not visible from below. Length of tail about  $3\frac{1}{2}$  times its total length. Both fins are poorly developed, the dorsal being slightly arched.

Tips of tail pointed. The colour of the dorsal surface is brownish. Under-surface uniformly whitish. Fins of tail grey.

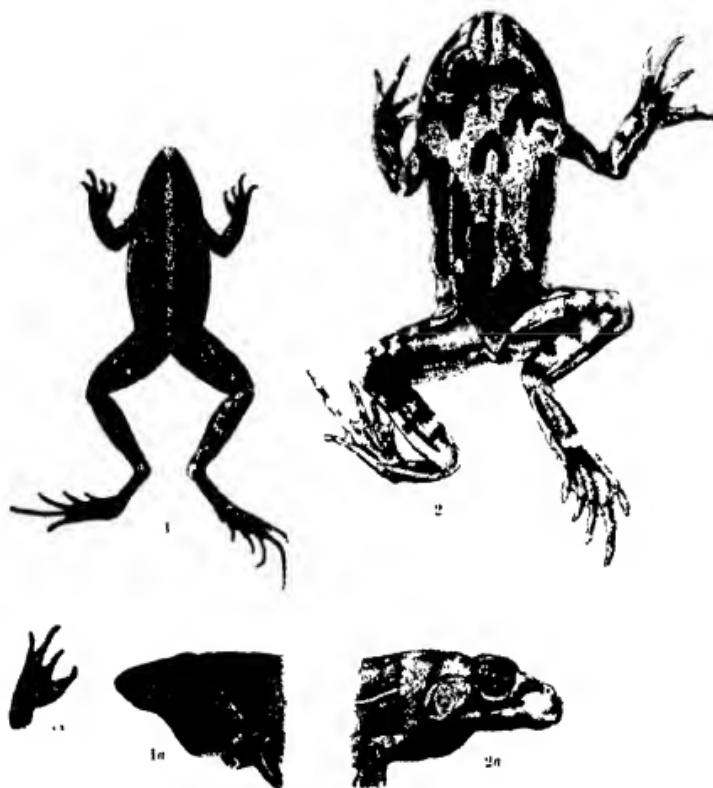
#### MEASUREMENTS

	mm		mm
Total length	13.00	Maximum breadth of body	3.75
Length of head and body	6.00	Maximum depth of body	2.75
Length of tail	7.00	Maximum depth of tail	2.00

The tadpoles of this species of *Bufo* can be easily recognised from the other described forms by the oral papillæ and serrated jaws.

#### EXPLANATION OF FIGURES

FIG. 1. *Rana parambikulamana*, actual size.  
 FIG. 1a. Side view of head  $\times 2$ .  
 FIG. 1b. Undersurface of head  $\times 2$ .  
 FIG. 2. *Rana luerhynca*  $\times 2$ .  
 FIG. 2a. Side view of head  $\times 2$ .  
 FIG. 3. *Rana intermedia*  $\times 1\frac{1}{2}$ .  
 FIG. 3a. Side view of head  $\times 1\frac{1}{2}$ .  
 FIG. 4. *Rana sauriceps*  $\times 2$ .  
 FIG. 4a. Side view of head  $\times 2$ .  
 FIG. 4b. Undersurface of head  $\times 3$ .  
 FIG. 4c. Foot  $\times 3$ .  
 FIG. 5. *Rana tenuilungua*  $\times 3$ .  
 FIG. 5a. Side view of head  $\times 3$ .  
 FIG. 6. *Nyctibatrachus sylvaticus*  $\times 1\frac{1}{2}$ .  
 FIG. 6a. Side view of head  $\times 1\frac{1}{4}$ .  
 FIG. 7. *Nannobatrachus hempholeensis*  $\times 3$ .  
 FIG. 7a. Side view of head  $\times 3$ .  
 FIG. 8. *Bufo brevirostris*  $\times 2$ .  
 FIG. 8a. Side view of head  $\times 2$ .  
 FIG. 9. *Philautus charius*  $\times 2$ .  
 FIG. 9a. Side view of head.  
 FIG. 10. *Philautus elegans*  $\times 3$ .  
 FIG. 11. *Philautus kottayachensis*  $\times 2$ .  
 FIG. 11a. Side view of head  $\times 3$ .  
 FIG. 12. *Philautus swaminanus*  $\times 2$ .  
 FIG. 12a. Side view of head  $\times 4$ .  
 FIG. 13. *Philautus melanensis*  $\times 2$ .  
 FIG. 13a. Side view of head  $\times 2$ .  
 FIG. 14. *Philautus narainensis*  $\times 2$ .  
 FIG. 14a. Side view of head  $\times 3$ .























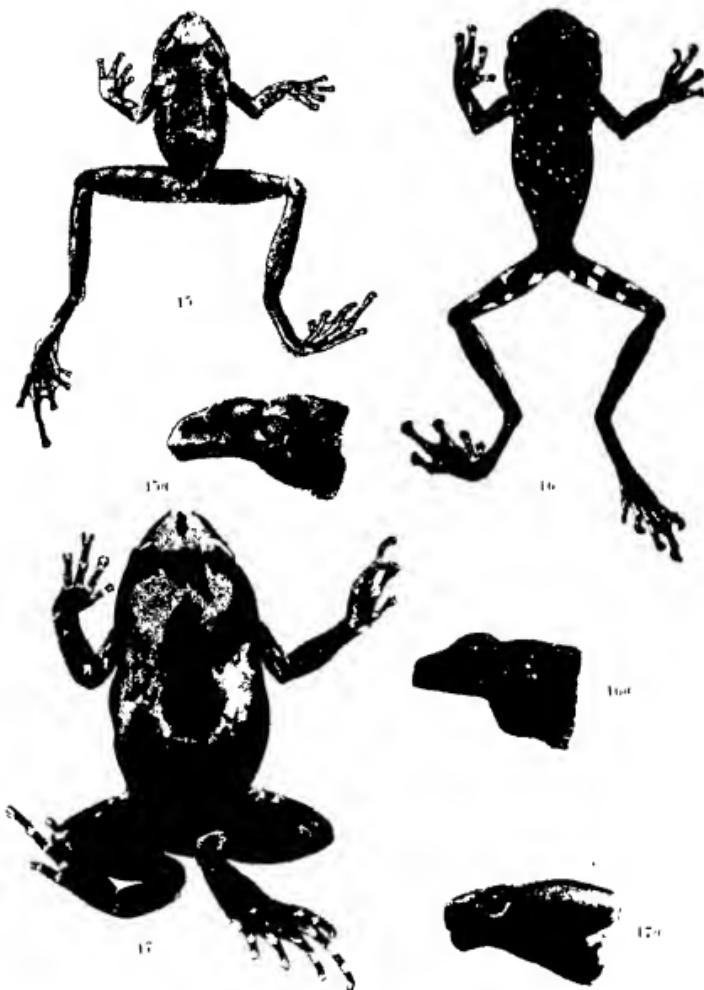




14a

14













21a



21d





25a



26a



27a



27b



FIG. 15 *Philautus longicrus*  $\times 2$   
FIG. 15a Side view of head  $\times 3$ .  
FIG. 16 *Philautus montanus*  $\times 1\frac{1}{2}$   
FIG. 16a. Side view of head  $\times 2$   
FIG. 17 *Ramanella minor*  $\times 3$   
FIG. 17a Side view of head  $\times 3$   
FIG. 18. *Ramanella triangularis rufiventris*  $\times 2$   
FIG. 18a Side view of head  $\times 3$   
FIG. 19 *Ramanella marmorata*  $\times 2$   
FIG. 19a Side view of head  $\times 3$   
FIG. 20. *Ramanella anamalaiensis*  $\times 3$

## TADPOLE\*

FIG. 21 *Philautus leucorhynchus*  $\times 3$   
FIG. 21a Mouth disc  $\times 5$   
FIG. 22 *Philautus hypomelas*  $\times 3$   
FIG. 22a Mouth disc  $\times 10$   
FIG. 23 *Philautus nasutus*  $\times 3$   
FIG. 23a. Mouth disc  $\times 10$   
FIG. 24 *Philautus pulcher*  $\times 3$   
FIG. 24a Mouth disc  $\times 5$   
FIG. 25. *Philautus taimabalis*  $\times 2$   
FIG. 25a Mouth disc  $\times 10$   
FIG. 26 *Rana aurantiaca*  $\times 3$   
FIG. 26a Mouth disc  $\times 10$   
FIG. 27 *Bufo brevirostris*  $\times 5$   
FIG. 27a Mouth disc  $\times 15$

## ON THE OCCURRENCE OF WINGED SPORES IN THE LOWER GONDWANA ROCKS OF INDIA AND AUSTRALIA.

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Received December 27, 1937

(Communicated by Prof. B. Sahni, F.R.S.)

DURING an investigation of fossil cuticles from some carbonaceous shales in the Lower Gondwana rocks of the Salt-Range, Punjab, numerous spores were discovered, some winged, others unwinged. Some of the two-winged spores showed a striking resemblance to Professor Seward's *Pityosporites antarcticus* which was suspected by him to be a pollen-grain of *Glossoptris*<sup>1,2</sup>. Two of these spores are shown in Fig. 3 A, B. The wings in both the spores are reticulately marked and are of the same size measuring about 37  $\mu$ . The striped body in each slightly differs in size—A measures about 4  $\mu$  and B about 10  $\mu$ .

The geological section from which these and other spores were obtained is a typical Lower Gondwana section with the Talchir Boulder bed at its base, and it includes more than one horizon containing *Glossoptris*, *Gangamopteris*, and other forms. The first spores were found in some specimens presented to Professor B. Sahni by Mr. E. R. Gee of the Geological Survey of India. Subsequently similar specimens were collected by myself at the same locality.<sup>3</sup>

After this a piece of shale from the Permo-Carboniferous rocks of Newcastle, New South Wales, was examined for cuticles as well as spores. This shale is so rich in the fronds of *Glossoptris Browniana* Brong. that rarely a bare face of the rock is exposed. Though I did not succeed in obtaining good samples of cuticles, numerous two-winged pollen-grains, all showing a general resemblance with *Pityosporites antarcticus* were found, some in groups, but chiefly lying free. A few spores were even seen adhering to small pieces of cuticle (see Fig. 1 C, D). No clear evidence of sporangia is available. Some of these Australian spores are shown in Figs. 1 and 2 lying in different postures, mostly dorsal, but rarely lateral, ventral and polar.

There are two sizes of spores represented in these figures. The smaller size (Fig. 1 A-C, Fig. 2 A-D) possesses reticulately marked wings, about 27  $\mu$  across, which in some spores appear to be united on the dorso-lateral

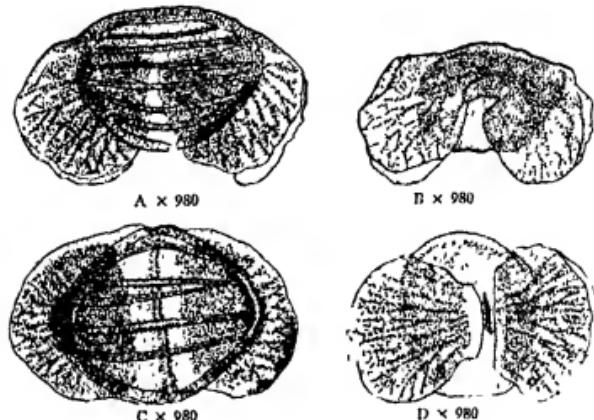


FIG. 2

Camera-lucida drawings of *Pityosporites Sewardi* sp. nov. Permo-Carboniferous of Newcastle, N.S.W., Australia. From a piece of shale crowded with *Glossoptris Browniana* Brong.

A. *Pityosporites Sewardi* sp. nov. Lateral view of the spore in Fig. 1,  $1 \times 980$

B. *P. Sewardi* sp. nov. The union of the two wings on the dorso-lateral side of spore, in Fig. 1, B  $\times 980$

C. *P. Sewardi* sp. nov. Dorsal view. Striped body seen above and the wings below  $\times 980$

D. *P. Sewardi* sp. nov. Ventral view showing the two wings  $\times 980$

side (Fig. 1 B, Fig. 2 B). The body, with a relatively thick wall, measures about  $36 \mu$  in diameter, and shows a number of well-marked horizontal stripes (Fig. 1 A, Fig. 2 A, C). On the other hand, the wings and the body of the larger spores (Fig. 1 D-F) measures about  $30 \mu$  and  $50 \mu$  respectively. In their striped body and reticulate wings these larger, lighter coloured spores show almost identical structure with the smaller ones. Even in living plants it is not uncommon to find different sizes of spores within the same sporangium. That the difference in preservation may also be responsible for this difference in size seems not impossible. However, after a careful examination of a large number of these spores I am inclined to refer them provisionally to two separate species.

The parentage of none of these spores, Indian or Australian, can be determined with certainty. The circumstantial evidence, however, strongly favours the view that they both represent the pollen-grains of *Glossoptris*

The Newcastle spores were obtained from the surface of the cuticle of *Glossopteris Browniana* Brong., from a shale which consists solely of fronds of this species, maceration of the rock matrix did not yield spores. The spores were obtained in hundreds from a small bit of this shale. As stated above they show a striking resemblance with *Pityosporites antarcticus*. As already stated, spores of a similar though not identical kind occur in the Permio-Carboniferous rocks of the Salt-Range, again in association with *Glossopteris*. Lastly, from Professor Seward's paper<sup>1</sup> it appears that Hamshaw Thomas has found evidence of winged spores of *Glossopteris* in South Africa. If his spores too resemble the others it can be said that winged spores of the general *Pityosporites* type occur in the *Glossopteris*-bearing rocks in widely scattered parts of Gondwanaland, namely, Antarctica, South Africa, India and Australia.

It must, however, be mentioned that the possibility of these spores being other than those of *Glossopteris* cannot be overlooked. No organic continuity is seen between the frond and these spores. Nor is there any clear evidence of the sporangia which must have contained them. Unless more light is thrown in this direction it cannot be said with any certainty that these are the pollen-grains of *Glossopteris*. But on the evidence available at present one is inclined to believe that these winged pollen-grains most probably belong to *Glossopteris* and that Professor Seward's unfortunate *Pityosporites antarcticus*, which met with adverse criticism, is after all a pollen-grain and most probably of *Glossopteris*. I have pleasure in naming the Australian spores of the smaller size shown in Fig. 1 A-C, Fig. 2 A-D, after the discoverer of the antarctic pollen-grain, as *Pityosporites Sewardi* sp. nov.

A detailed description of all these spores and others, both winged and unwinged, found in the Lower Gondwana rocks of the Salt-Range, will appear in a subsequent paper.

I wish to express my grateful thanks to Professor B. Sahni, F.R.S., for his ready help, invaluable criticism and never failing encouragement.

#### REFERENCES

- 1 Seward, A. C., "Antarctic fossil plants," *British Antarctic Expedition, 1910, Nat. Hist. Report*, London, 1914
- 2 Seward, A. C., "An Antarctic pollen-grain, fact or fancy?" *New Phyt.*, 1933, **32**, 4, 311-13.
- 8 Virkki, C., "A Lower Gondwana flora from the Salt Range, Punjab," *Proc. 25th Ind. Sci. Congress, Calcutta, 1938, Pt. III, 150-51 (Advance Abstract)*.

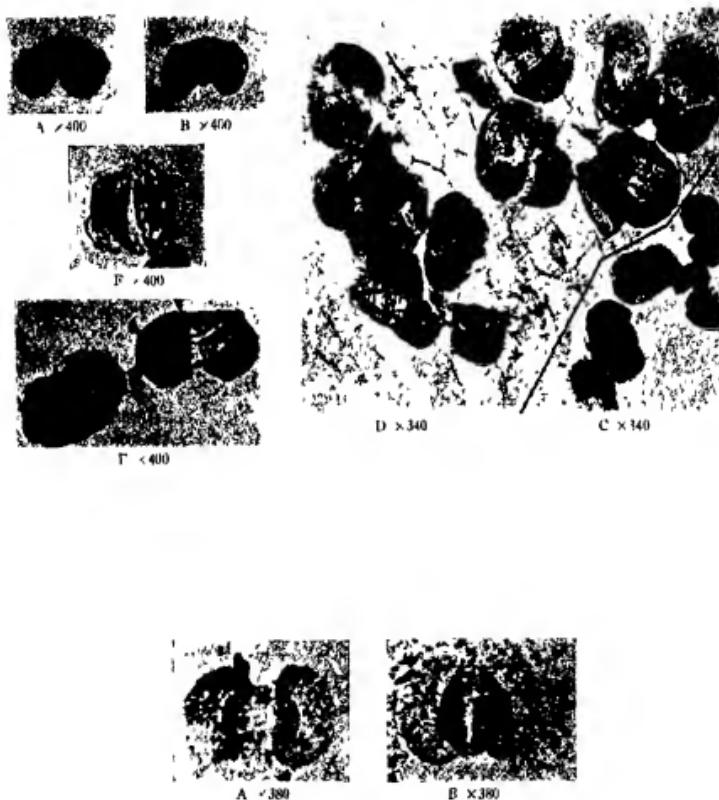


FIG. 3



EXPLANATION OF PLATE

Fig 1 Untouched photographs of *Pityosporites Sewardi* sp nov and *Pityosporites* sp. (larger spores) Permo-Carboniferous of Newcastle, N.S.W., Australia. From a piece of shale crowded with *Glossopeltis Browniana* Brong.

- A *Pityosporites Sewardi* sp nov Lateral view with the two wings attached to the striped body  $\times 400$  See also Fig 2 A
- B *P. Sewardi* sp nov The union of the two wings on the dorso-lateral side  $\times 400$   
The same spore is drawn in Fig 2 B
- C *P. Sewardi* sp nov Adhering to a piece of cuticle of *G. Browniana* Brong  $\times 340$
- D *Pityosporites* sp Adhering to the same piece of cuticle as C, mostly showing their dorsal aspect  $\times 340$
- E *Pityosporites* sp The one on the right side shows the two wings attached to the striped body  $\times 400$
- F *Pityosporites* sp Ventral view.  $\times 400$

Fig 2. A, B Ventral view of the two-winged spores with striped body and reticulate wings. From a horizon 1½ ft above the Takchir Boulder Bed Loc. Kathwai, Salt-Ridge, Punjab  $\times 180$  (Coll C Virkki)



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